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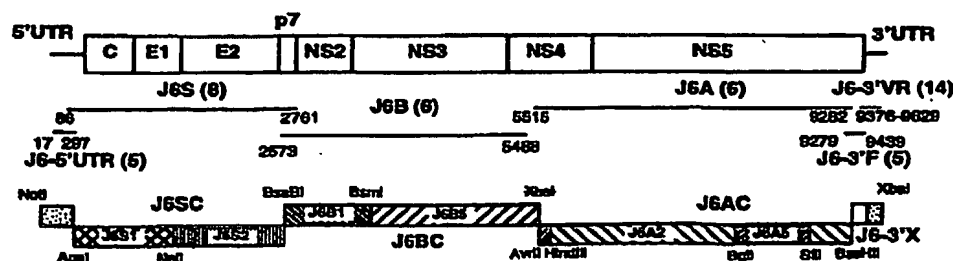
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(54) Title: CLONED GENOME OF INFECTIOUS HEPATITIS C VIRUS OF GENOTYPE 2a AND USES THEREOF



(57) Abstract: The present invention discloses nucleic acid sequence which encodes infectious hepatitis C virus of strain HC-J6_{CH}, genotype 2a, and the use of the sequence, and polypeptides encoded by all or part of the sequence, in the development of vaccines and diagnostics for HCV and in the development of screening assays for the identification of antiviral agents for HCV.

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Title Of Invention

Cloned Genome Of Infectious
Hepatitis C Virus of Genotype 2a And Uses Thereof

Field Of Invention

The present invention relates to molecular approaches to the production of nucleic acid sequence which comprises the genome of infectious hepatitis C virus. In particular, the invention provides a nucleic acid sequence which comprises the genome of an infectious hepatitis C virus of genotype 2a. The invention therefore relates to the use of the nucleic acid sequence and polypeptides encoded by all or part of the sequence in the development of vaccines and diagnostic assays for HCV and in the development of screening assays for the identification of antiviral agents for HCV.

Background Of Invention

Hepatitis C virus (HCV) has a positive-sense single-strand RNA genome and is a member of the genus *Hepacivirus* within the *Flaviviridae* family of viruses (Rice, 1996). As for all positive-stranded RNA viruses, the genome of HCV functions as mRNA from which all viral proteins necessary for propagation are translated.

The viral genome of HCV is approximately 9600 nucleotides (nts) in length and consists of a highly conserved 5' untranslated region (UTR), a single long open reading frame (ORF) of approximately 9,000 nts and a complex 3' UTR. The 5' UTR contains an internal ribosomal entry site (Tsukiyama-Kohara et al., 1992;

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° Honda et al., 1996). The 3' UTR consists of a short variable region, a polypyrimidine tract of variable length and, at the 3' end, a highly conserved region of approximately 100 nucleotides (Kolykhalov et al., 1996; 5 Tanaka et al., 1995; Tanaka et al., 1996; Yamada et al., 1996). The last 46 nucleotides of this conserved region were predicted to form a stable stem-loop structure thought to be critical for viral replication (Blight and 10 Rice, 1997; Ito and Lai, 1997; Tsuchihara et al., 1997). The ORF encodes a large polypeptide precursor that is cleaved into at least 10 proteins by host and viral proteinases (Rice, 1996). The predicted envelope proteins contain several conserved N-linked 15 glycosylation sites and cysteine residues (Okamoto et al., 1992a). The NS3 gene encodes a serine protease and an RNA helicase and the NS5B gene encodes an RNA-dependent RNA polymerase.

20 A remarkable characteristic of HCV is its genetic heterogeneity, which is manifested throughout the genome (Bukh et al., 1995). The most heterogeneous regions of the genome are found in the envelope genes, in particular the hypervariable region 1 (HVR1) at the 25 N-terminus of E2 (Hijikata et al., 1991; Weiner et al., 1991). HCV circulates as a quasispecies of closely related genomes in an infected individual. Globally, six major HCV genotypes (genotypes 1-6) and multiple 30 subtypes (a, b, c, etc.) have been identified (Bukh et al., 1993; Simmonds et al., 1993).

The nucleotide and deduced amino acid sequences among isolates within a quasispecies generally differ by < 2%, whereas those between isolates of 35 different genotypes vary by as much as 35%. Genotypes

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1, 2 and 3 are found worldwide and constitute more than 90% of the HCV infections in North and South America, Europe, Russia, China, Japan and Australia (Forns and Bukh, 1998). Throughout these regions genotype 1 accounts for the majority of HCV infections but genotypes 2 and 3 each account for 5-15%.

At present, more than 80% of individuals infected with HCV become chronically infected and these chronically infected individuals have a relatively high risk of developing chronic hepatitis, liver cirrhosis and hepatocellular carcinoma (Hoofnagle, 1997). The only effective therapy for chronic hepatitis C, interferon (IFN), alone or in combination with ribavirin, induces a sustained response in less than 50% of treated patients (Davis et al., 1998; McHutchinson et al., 1998). Consequently, HCV is currently the most common cause of end stage liver failure and the reason for about 30% of liver transplants performed in the U.S. (Hoofnagle, 1997). In addition, a number of recent studies suggested that the severity of liver disease and the outcome of therapy may be genotype-dependent (reviewed in Bukh et al., 1997). In particular, these studies suggested that infection with HCV genotype 1b was associated with more severe liver disease (Brechot, 1997) and a poorer response to IFN therapy (Fried and Hoofnagle, 1995). As a result of the inability to develop a universally effective therapy against HCV infection, it is estimated that there are still more than 25,000 new infections yearly in the U.S. (Alter 1997). Moreover, since there is no vaccine for HCV, HCV remains a serious public health problem.

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Despite the intense interest in the development of vaccines and therapies for HCV, progress has been hindered by the absence of a useful cell culture system and the lack of any small animal model for laboratory study. For example, while replication of HCV in several cell lines has been reported, such observations have turned out not to be highly reproducible. In addition, the chimpanzee is the only animal model, other than man, for this disease. Consequently, HCV has been studied only by using clinical materials obtained from patients or experimentally infected chimpanzees, an animal model whose availability is very limited.

However, several researchers have recently reported the construction of infectious cDNA clones of HCV, the identification of which would permit a more effective search for susceptible cell lines and facilitate molecular analysis of the viral genes and their function. For example, Yoo et al., and Dash et al., (1997) (1995) reported that RNA transcripts from cDNA clones of HCV-1 (genotype 1a) and HCV-N (genotype 1b), respectively, resulted in viral replication after transfection into human hepatoma cell lines. Unfortunately, the viability of these clones was not tested in vivo and concerns were raised about the infectivity of these cDNA clones in vitro (Fausto, 1997). In addition, both clones did not contain the terminal 98 conserved nucleotides at the very 3' end of the UTR.

Kolykhalov et al., (1997) and Yanagi et al. (1997, 1998) reported the derivation from HCV strains H77 (genotype 1a) and HC-J4 (genotype 1b) of cDNA clones

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° of HCV that are infectious for chimpanzees. However, while these infectious clones will aid in studying HCV replication and pathogenesis and will provide an important tool for development of in vitro replication and propagation systems, it is important to have infectious clones of more than one genotype, given the extensive genetic heterogeneity of HCV and the potential impact of such heterogeneity on the development of effective therapies and vaccines for HCV.

10 In addition, synthetic chimeric viruses can be used to map the functional regions of viruses with different phenotypes. In flaviviruses and pestiviruses, infectious chimeric viruses have been successfully engineered to express different functional units of related viruses (Bray and Lai, 1991; Pletnev et al., 1992, 1998; Vassilev et al., 1997) and in some cases it has been possible to make chimeras between non-related or distantly related viruses. For instance, the IRES element of poliovirus or bovine viral diarrhoea virus has been replaced with IRES sequences from HCV (Frolov et al., 1998; Lu and Wimmer, 1996; Zhao et al., 1999). Recently, the construction of an infectious chimera of two closely related HCV subtypes has been reported. The chimera contained the complete ORF of a genotype 1b strain but had the 5' and 3' termini of a genotype 1a strain (Yanagi et al., 1998).

30 It is important to determine whether chimeras constructed from more divergent HCV strains are infectious because such chimeras could be used to define the functions of viral units and to dissect the immune response.

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Summary Of The Invention

The present invention relates to nucleic acid sequence which comprises the genome of infectious hepatitis C virus and in particular, nucleic acid sequence which comprises the genome of infectious hepatitis C virus of genotype 2a. It is therefore an object of the invention to provide nucleic acid sequence which encodes infectious hepatitis C virus. Such nucleic acid sequence is referred to throughout the application as "infectious nucleic acid sequence".

For the purposes of this application, nucleic acid sequence refers to RNA, DNA, cDNA or any variant thereof capable of directing host organism synthesis of a hepatitis C virus polypeptide. It is understood that nucleic acid sequence encompasses nucleic acid sequences, which due to degeneracy, encode the same polypeptide sequence as the nucleic acid sequences described herein.

The invention also relates to the use of the infectious nucleic acid sequences to produce chimeric genomes consisting of portions of the open reading frames of nucleic acid sequences of other genotypes (including, but not limited to, genotypes 1, 2, 3, 4, 5 and 6) and subtypes (including, but not limited to, subtypes 1a, 1b, 2a, 2b, 2c, 3a, 4a-4f, 5a and 6a) of HCV. For example, infectious nucleic acid sequence of the 2a strain HC-J6, described herein can be used to produce chimeras with sequences from the genomes of other strains of HCV from different genotypes or subtypes. Nucleic acid sequences which comprise sequences from two or more HCV genotypes or subtypes are designated "chimeric nucleic acid sequences".

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° The invention further relates to mutations of the infectious nucleic acid sequence of the invention where mutation includes, but is not limited to, point mutations, deletions and insertions. In one embodiment, a gene or fragment thereof can be deleted to determine the effect of the deleted gene or genes on the properties of the encoded virus such as its virulence and its ability to replicate. In an alternative embodiment, a mutation may be introduced into the infectious nucleic acid sequences to examine the effect of the mutation on the properties of the virus.

 The invention also relates to the introduction of mutations or deletions into the infectious nucleic acid sequence in order to produce an attenuated hepatitis C virus suitable for vaccine development.

 The invention further relates to the use of the infectious nucleic acid sequence to produce attenuated viruses via passage in vitro or in vivo of the viruses produced by transfection of a host cell with the infectious nucleic acid sequence.

 The present invention also relates to the use of the nucleic acid sequence of the invention or fragments thereof in the production of polypeptides where "nucleic acid sequence of the invention" refers to infectious nucleic acid sequence, mutations of infectious nucleic acid sequence, chimeric nucleic acid sequence and sequences which comprise the genome of attenuated viruses produced from the infectious nucleic acid sequence of the invention. In one embodiment, said polypeptide or polypeptides are fully or partially purified from hepatitis C virus produced by cells transfected with nucleic acid sequence of the invention.

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° In another embodiment, the polypeptide or polypeptides are produced recombinantly from a fragment of the nucleic acid sequences of the invention. In yet another embodiment, the polypeptides are chemically synthesized.

5 The polypeptides of the invention, especially structural polypeptides, can serve as immunogens in the development of vaccines or as antigens in the development of diagnostic assays for detecting the presence of HCV in biological samples.

10 The invention therefore also relates to vaccines for use in immunizing mammals especially humans against hepatitis C. In one embodiment, the vaccine comprises one or more polypeptides made from the nucleic acid sequence of the invention or fragment thereof. In 15 a second embodiment, the vaccine comprises a hepatitis C virus produced by transfection of host cells with the nucleic acid sequences of the invention.

20 The present invention therefore relates to methods for preventing hepatitis C in a mammal. In one embodiment the method comprises administering to a mammal a polypeptide or polypeptides encoded by the nucleic acid sequence of the invention in an amount 25 effective to induce protective immunity to hepatitis C. In another embodiment, the method of prevention comprises administering to a mammal a hepatitis C virus of the invention in an amount effective to induce protective immunity against hepatitis C.

30 In yet another embodiment, the method of protection comprises administering to a mammal the nucleic acid sequence of the invention or a fragment thereof in an amount effective to induce protective 35 immunity against hepatitis C.

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The invention also relates to hepatitis C viruses produced by host cells transfected with the nucleic acid sequence of the present invention.

The invention therefore also provides pharmaceutical compositions comprising the nucleic acid sequence of the invention and/or the encoded hepatitis C viruses. The invention further provides pharmaceutical compositions comprising polypeptides encoded by the nucleic acid sequence of the invention or fragments thereof. The pharmaceutical compositions of the invention may be used prophylactically or therapeutically.

The invention also relates to antibodies to the hepatitis C virus of the invention or their encoded polypeptides and to pharmaceutical compositions comprising these antibodies.

The invention also relates to the use of the nucleic acid sequences of the invention to identify cell lines capable of supporting the replication of HCV in vitro.

The invention further relates to the use of the nucleic acid sequences of the invention or their encoded viral enzymes (e.g. NS3 serine protease, NS3 helicase, NS5B RNA polymerase) to develop screening assays to identify antiviral agents for HCV.

Brief Description Of Figures

Figure 1 shows the amplification and cloning of hepatitis C virus genotype 2a (strain HC-J6_{CH}). The nucleotide positions correspond to the sequence of PJ6CF, a full length cDNA clone of hepatitis C virus, genotype 2a, strain HC-J6_{CH}. Products from polymerase

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chain reaction are also shown. The names of the clones obtained from these products are indicated (number of clones sequenced are shown in parenthesis). The composition of the full-length cDNA clone is shown at the bottom. The restriction enzymes used for cloning are indicated. An *Xba*I site in HC-J6_{CH} was eliminated by a silent substitution at position 5494.

Figure 2 shows tree analysis of clones amplified from an infectious acute phase plasma pool generated in a chimpanzee inoculated with human plasma containing strain HC-J6 (Okamoto *et al.*, 1991) as well as a tree of the predicted polyprotein sequence of HC-J6_{CH} and the infectious HC-J6_{CH} cDNA clone (pJ6CF). The nucleotide positions with deletions or insertions were stripped in the analysis of the clones. Multiple sequence alignments and tree analyses were performed with GeneWorks (Oxford Molecular Group) (Bukh *et al.*, 1995). Genotype designations are indicated. Other sequences included in the analysis are HC-J8 (Okamoto *et al.*, 1992), genotype 1a infectious clone BEBE1 (Nakao *et al.*, 1996), H77C (Yanagi *et al.*, 1997); genotype 1b infectious clone J4L6S (Yanagi *et al.*, 1998). The scale in each tree indicates the calculated genetic distance.

Figure 3 shows the alignment of the hypervariable region 1 sequences from 8 J6S clones of strain HC-J6_{CH}. HC-J6_{CH} represents the consensus amino acid sequence of the infectious plasma pool from an experimentally infected chimpanzee. HC-J6 is the published amino acid sequence of the original inoculum (Okamoto *et al.*, 1991).

Figure 4 shows the construction of four intertypic chimeric cDNA clones. White boxes are

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sequences derived from genotype 2a clone pJ6CF, and
black boxes are sequences derived from genotype 1a clone
pCV-H77C (Yanagi et al., 1997). An NdeI site (mutation
at position 9158 of pCV-H77C) was eliminated and an
artificial NdeI site (mutation at position 2765 of
pCV-H77C) was created by site-directed mutagenesis;
silent mutations are underlined.

Figures 5A and 5B show the alignment of the
nucleotide sequences of the 5' (Fig. 5A) and 3' UTRs
(Fig. 5B) and the amino acid sequences of E2/p7/NS2
junctions (Fig. 5B) in the intertypic 1a, 2a chimeric
cDNA clones. In the 5' UTR alignment, the first 39 nts
of core believed to be important for the IRES function
were included (Lemon and Honda, 1997). Top line: the
sequence of the infectious genotype 1a clone pCV-H77C
(Yanagi et al., 1997). Bottom line: the sequence of the
infectious genotype 2a clone pJ6CF. Dot: identity with
the sequence of H77C. Capital letter: different from the
sequence of H77C. Dash: deletion. Bold face: initiation
or stop codon of the ORF. Underlined: AgeI cleavage
site. Arrow: putative sites in the HCV polyprotein
cleaved by host signal peptidases. Numbering
corresponds to the sequence of pCV-H77C.

Figures 6A-6F show the nucleotide sequence of
the infectious hepatitis C virus clone of genotype 1a
strain H77C and Figures 6G-6H show the amino acid
sequence encoded by the clone.

Figures 7A-7F show the nucleotide sequence of
the infectious hepatitis C virus clone of genotype 1b
strain HC-J4 and Figures 7G-H show the amino acid
sequence encoded by the clone.

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DESCRIPTION OF THE INVENTION

The present invention relates to nucleic acid sequence which comprises the genome of an infectious hepatitis C virus. More specifically, the invention relates to nucleic acid sequence which encodes infectious hepatitis C virus of strain HC-J6_{CH}, genotype 2a. The infectious nucleic acid sequence of the invention is shown in SEQ ID NO:1 and is contained in a plasmid construct deposited with the American Type Culture Collection (ATCC) on May 28, 1999 and having ATCC accession number PTA-153.

The invention also relates to "chimeric nucleic acid sequences" where the chimeric nucleic acid sequences consist of open-reading frame sequences and/or 5' and/or 3' untranslated sequences taken from nucleic acid sequences of hepatitis C viruses of different genotypes or subtypes.

In one embodiment, the chimeric nucleic acid sequence consists of sequence from the genome of infectious HCV of genotype 2a which encodes structural polypeptides and sequence from the genome of a HCV of a different genotype or subtype which encodes nonstructural polypeptides.

Alternatively, the nonstructural region of infectious HCV of genotype 2a and structural region of a HCV of a different genotype or subtype may be combined. This will result in a chimeric nucleic acid sequence consisting of sequence from the genome of infectious HCV of genotype 2a which encodes nonstructural polypeptides and sequence from the genome of a HCV of a another genotype or subtype which encodes structural polypeptides.

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Preferably, the nucleic acid sequence from the genome of the infectious HCV clone of genotype 1a (deposited with the ATCC on June 2, 1999 ; Figures 6A-6F), or the nucleic acid sequence from the genome of the infectious HCV clone of genotype 1b (ATCC accession number 209596; Figures 7A-7F) is used to construct the chimeric nucleic acid sequence with the HCV of genotype 2a of the invention.

10 It is believed that the construction of such chimeric nucleic acid sequences will be of importance in studying the growth and virulence properties of hepatitis C virus and in the production of candidate hepatitis C virus vaccines suitable to confer protection against multiple genotypes of HCV. For example, one might produce a "multivalent" vaccine by putting epitopes from several genotypes or subtypes into one clone. Alternatively one might replace just a single gene from an infectious sequence with the corresponding gene from the genomic sequence of a strain from another genotype or subtype or create a chimeric gene which contains portions of a gene from two genotypes or subtypes. Examples of genes which could be replaced or which could be made chimeric, include, but are not limited to, the E1, E2 and NS4 genes.

25 The invention further relates to mutations of the infectious nucleic acid sequences where "mutations" include, but are not limited to, point mutations, deletions and insertions. Of course, one of ordinary skill in the art would recognize that the size of the insertions would be limited by the ability of the resultant nucleic acid sequence to be properly packaged within the virion. Such mutations could be produced by

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techniques known to those of skill in the art such as site-directed mutagenesis, fusion PCR, and restriction digestion followed by religation.

In one embodiment, mutagenesis might be undertaken to determine sequences that are important for viral properties such as replication or virulence. For example, one may introduce a mutation into the infectious nucleic acid sequence which eliminates the cleavage site between the NS4A and NS4B polypeptides to examine the effects on viral replication and processing of the polypeptide.

Alternatively, one may delete all or part of a gene or of the 5' or 3' nontranslated region contained in an infectious nucleic acid sequence and then transfect a host cell (animal or cell culture) with the mutated sequence and measure viral replication in the host by methods known in the art such as RT-PCR. Preferred genes include, but are not limited to, the P7, NS4B and NS5A genes. Of course, those of ordinary skill in the art will understand that deletion of part of a gene, preferably the central portion of the gene, may be preferable to deletion of the entire gene in order to conserve the cleavage site boundaries which exist between proteins in the HCV polyprotein and which are necessary for proper processing of the polyprotein.

In the alternative, if the transfection is into a host animal such as a chimpanzee, one can monitor the virulence phenotype of the virus produced by transfection of the mutated infectious nucleic acid sequence by methods known in the art such as measurement of liver enzyme levels (alanine aminotransferase (ALT) or isocitrate dehydrogenase (ICD)) or by histopathology

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° of liver biopsies. Thus, mutations of the infectious nucleic acid sequences may be useful in the production of attenuated HCV strains suitable for vaccine use.

The invention also relates to the use of the
5 infectious nucleic acid sequence of the present invention to produce attenuated viral strains via passage in vitro or in vivo of the virus produced by transfection with the infectious nucleic acid sequence.

10 The present invention therefore relates to the use of the nucleic acid sequence of the invention to identify cell lines capable of supporting the replication of HCV.

15 In particular, it is contemplated that the mutations of the infectious nucleic acid sequence of the invention and the production of chimeric sequences as discussed above may be useful in identifying sequences critical for cell culture adaptation of HCV and hence, may be useful in identifying cell lines capable of
20 supporting HCV replication.

Transfection of tissue culture cells with the nucleic acid sequences of the invention may be done by methods of transfection known in the art such as
25 electroporation, precipitation with DEAE-Dextran or calcium phosphate or liposomes.

In one such embodiment, the method comprises the growing of animal cells, especially human cells, in vitro and transfecting the cells with the nucleic acid
30 of the invention, then determining if the cells show indicia of HCV infection. Such indicia include the detection of viral antigens in the cell, for example, by immunofluorescence procedures well known in the art; the
35 detection of viral polypeptides by Western blotting

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° using antibodies specific therefor; and the detection of newly transcribed viral RNA within the cells via methods such as RT-PCR. The presence of live, infectious virus particles following such tests may also be shown by
5 injection of cell culture medium or cell lysates into healthy, susceptible animals, with subsequent exhibition of the signs and symptoms of HCV infection.

Suitable cells or cell lines for culturing HCV include, but are not limited to, lymphocyte and
10 hepatocyte cell lines known in the art.

Alternatively, primary hepatocytes can be cultured, and then infected with HCV; or, the hepatocyte cultures could be derived from the livers of infected
15 chimpanzees. In addition, various immortalization methods known to those of ordinary skill in the art can be used to obtain cell lines derived from hepatocyte cultures. For example, primary hepatocyte cultures may be fused to a variety of cells to maintain stability.
20

The present invention further relates to the in vitro and in vivo production of hepatitis C viruses from the nucleic acid sequences of the invention.

In one embodiment, the sequences of the
25 invention can be inserted into an expression vector that functions in eukaryotic cells. Eukaryotic expression vectors suitable for producing high efficiency gene transfer in vivo are well known to those of ordinary skill in the art and include, but are not limited to,
30 plasmids, vaccinia viruses, retroviruses, adenoviruses and adeno-associated viruses.

In another embodiment, the sequences contained in the recombinant expression vector can be transcribed
35 in vitro by methods known to those of ordinary skill in

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the art in order to produce RNA transcripts which encode the hepatitis C viruses of the invention. The hepatitis C viruses of the invention may then be produced by transfecting cells by methods known to those of ordinary skill in the art with either the in vitro transcription mixture containing the RNA transcripts or with the recombinant expression vectors containing the nucleic acid sequences described herein.

The hepatitis C viruses produced from the sequences of the invention may be purified or partially purified from the transfected cells by methods known to those of ordinary skill in the art. In a preferred embodiment, the viruses are partially purified prior to their use as immunogens in the pharmaceutical compositions and vaccines of the present invention.

The present invention therefore relates to the use of the hepatitis C viruses produced from the nucleic acid sequences of the invention as immunogens in live or killed (e.g., formalin inactivated) vaccines to prevent hepatitis C in a mammal.

In an alternative embodiment, the immunogen of the present invention may be an infectious nucleic acid sequence, a chimeric nucleic acid sequence, or a mutated infectious nucleic acid sequence which encodes a hepatitis C virus. Where the sequence is a cDNA sequence, the cDNAs and their RNA transcripts may be used to transfect a mammal by direct injection into the liver tissue of the mammal as described in the Examples.

Alternatively, direct gene transfer may be accomplished via administration of a eukaryotic expression vector containing a nucleic acid sequence of the invention.

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In yet another embodiment, the immunogen may be a polypeptide encoded by the nucleic acid sequences of the invention. The present invention therefore also relates to polypeptides produced from the nucleic acid sequences of the invention or fragments thereof. In one
5 embodiment, polypeptides of the present invention can be recombinantly produced by synthesis from the nucleic acid sequences of the invention or isolated fragments thereof, and purified, or partially purified, from
10 transfected cells using methods already known in the art. In an alternative embodiment, the polypeptides may be purified or partially purified from viral particles produced via transfection of a host cell with the
15 nucleic acid sequences of the invention. Such polypeptides might, for example, include either capsid or envelope polypeptides prepared from the sequences of the present invention.

20 When used as immunogens, the nucleic acid sequences of the invention, or the polypeptides or viruses produced therefrom, are preferably partially purified prior to use as immunogens in pharmaceutical compositions and vaccines of the present invention.
25 When used as a vaccine, the sequences and the polypeptide and virus products thereof, can be administered alone or in a suitable diluent, including, but not limited to, water, saline, or some type of
30 buffered medium. The vaccine according to the present invention may be administered to an animal, especially a mammal, and most especially a human, by a variety of routes, including, but not limited to, intradermally, intramuscularly, subcutaneously, or in any combination
35 thereof.

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Suitable amounts of material to administer for prophylactic and therapeutic purposes will vary depending on the route selected and the immunogen (nucleic acid, virus, polypeptide) administered. One
5 skilled in the art will appreciate that the amounts to be administered for any particular treatment protocol can be readily determined without undue experimentation. The vaccines of the present invention may be
10 administered once or periodically until a suitable titer of anti-HCV antibodies appear in the blood. For an immunogen consisting of a nucleic acid sequence, a suitable amount of nucleic acid sequence to be used for prophylactic purposes might be expected to fall in the
15 range of from about 100 µg to about 5 mg and most preferably in the range of from about 500 µg to about 2mg. For a polypeptide, a suitable amount to use for prophylactic purposes is preferably 100 ng to 100 µg and
20 for a virus 10^2 to 10^6 infectious doses. Such administration will, of course, occur prior to any sign of HCV infection.

A vaccine of the present invention may be employed in such forms as capsules, liquid solutions,
25 suspensions or elixirs for oral administration, or sterile liquid forms such as solutions or suspensions. An inert carrier is preferably used, such as saline or phosphate-buffered saline, or any such carrier in which
30 the HCV of the present invention can be suitably suspended. The vaccines may be in the form of single dose preparations or in multi-dose flasks which can be utilized for mass-vaccination programs of both animals
35 and humans. For purposes of using the vaccines of the present invention reference is made to Remington's

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° Pharmaceutical Sciences, Mack Publishing Co., Easton, Pa., Osol (Ed.) (1980); and New Trends and Developments in Vaccines, Voller et al. (Eds.), University Park Press, Baltimore, Md. (1978), both of which provide much
5 useful information for preparing and using vaccines. Of course, the polypeptides of the present invention, when used as vaccines, can include, as part of the composition or emulsion, a suitable adjuvant, such as
10 alum (or aluminum hydroxide) when humans are to be vaccinated, to further stimulate production of antibodies by immune cells. When nucleic acids, viruses or polypeptides are used for vaccination purposes, other specific adjuvants such as CpG motifs (Krieg, A.K. et
15 al. (1995) and (1996)), may prove useful.

When the nucleic acids, viruses and polypeptides of the present invention are used as vaccines or inocula, they will normally exist as physically discrete units suitable as a unitary dosage
20 for animals, especially mammals, and most especially humans, wherein each unit will contain a predetermined quantity of active material calculated to produce the desired immunogenic effect in association with the
25 required diluent. The dose of said vaccine or inoculum according to the present invention is administered at least once. In order to increase the antibody level, a second or booster dose may be administered at some time after the initial dose. The need for, and timing of,
30 such booster dose will, of course, be determined within the sound judgment of the administrator of such vaccine or inoculum and according to sound principles well known in the art. For example, such booster dose could
35 reasonably be expected to be advantageous at some time

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- ° between about 2 weeks to about 6 months following the initial vaccination. Subsequent doses may be administered as indicated.

The nucleic acid sequences, viruses and polypeptides of the present invention can also be administered for purposes of therapy, where a mammal, especially a primate, and most especially a human, is already infected, as shown by well known diagnostic measures. When the nucleic acid sequences, viruses or polypeptides of the present invention are used for such therapeutic purposes, much of the same criteria will apply as when it is used as a vaccine, except that inoculation will occur post-infection. Thus, when the nucleic acid sequences, viruses or polypeptides of the present invention are used as therapeutic agents in the treatment of infection, the therapeutic agent comprises a pharmaceutical composition containing a sufficient amount of said nucleic acid sequences, viruses or polypeptides so as to elicit a therapeutically effective response in the organism to be treated. Of course, the amount of pharmaceutical composition to be administered will, as for vaccines, vary depending on the immunogen contained therein (nucleic acid, polypeptide, virus) and on the route of administration.

The therapeutic agent according to the present invention can thus be administered by subcutaneous, intramuscular or intradermal routes. One skilled in the art will certainly appreciate that the amounts to be administered for any particular treatment protocol can be readily determined without undue experimentation. Of course, the actual amounts will vary depending on the route of administration as well as the sex, age, and

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° clinical status of the subject which, in the case of human patients, is to be determined with the sound judgment of the clinician.

The therapeutic agent of the present invention
5 can be employed in such forms as capsules, liquid solutions, suspensions or elixirs, or sterile liquid forms such as solutions or suspensions. An inert carrier is preferably used, such as saline, phosphate-buffered saline, or any such carrier in which the HCV of the
10 present invention can be suitably suspended. The therapeutic agents may be in the form of single dose preparations or in the multi-dose flasks which can be utilized for mass-treatment programs of both animals and
15 humans. Of course, when the nucleic acid sequences, viruses or polypeptides of the present invention are used as therapeutic agents they may be administered as a single dose or as a series of doses, depending on the situation as determined by the person conducting the
20 treatment.

The nucleic acids, polypeptides and viruses of the present invention can also be utilized in the production of antibodies against HCV. The term
25 "antibody" is herein used to refer to immunoglobulin molecules and immunologically active portions of immunoglobulin molecules. Examples of antibody molecules are intact immunoglobulin molecules, substantially intact immunoglobulin molecules and
30 portions of an immunoglobulin molecule, including those portions known in the art as Fab, F(ab')₂ and F(v) as well as chimeric antibody molecules.

Thus, the polypeptides, viruses and nucleic
35 acid sequences of the present invention can be used in

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° the generation of antibodies that immunoreact (i.e., specific binding between an antigenic determinant-containing molecule and a molecule containing an antibody combining site such as a whole antibody molecule or an active portion thereof) with antigenic determinants on the surface of hepatitis C virus particles.

The present invention therefore also relates to antibodies produced following immunization with the nucleic acid sequences, viruses or polypeptides of the present invention. These antibodies are typically produced by immunizing a mammal with an immunogen or vaccine to induce antibody molecules having immunospecificity for polypeptides or viruses produced in response to infection with the nucleic acid sequences of the present invention. When used in generating such antibodies, the nucleic acid sequences, viruses, or polypeptides of the present invention may be linked to some type of carrier molecule. The resulting antibody molecules are then collected from said mammal. Antibodies produced according to the present invention have the unique advantage of being generated in response to authentic, functional polypeptides produced according to the actual cloned HCV genome.

The antibody molecules of the present invention may be polyclonal or monoclonal. Monoclonal antibodies are readily produced by methods well known in the art. Portions of immunoglobulin molecules, such as Fabs, as well as chimeric antibodies, may also be produced by methods well known to those of ordinary skill in the art of generating such antibodies.

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° The antibodies according to the present invention may also be contained in blood, plasma, serum, hybridoma supernatants, and the like. Alternatively, the antibody of the present invention is isolated to the extent desired by well known techniques such as, for example, using DEAE Sephadex. The antibodies produced according to the present invention may be further purified so as to obtain specific classes or subclasses of antibody such as IgM, IgG, IgA, and the like. Antibodies of the IgG class are preferred for purposes of passive protection.

 The antibodies of the present invention are useful in the prevention and treatment of diseases caused by hepatitis C virus in animals, especially mammals, and most especially humans.

 In providing the antibodies of the present invention to a recipient mammal, preferably a human, the dosage of administered antibodies will vary depending on such factors as the mammal's age, weight, height, sex, general medical condition, previous medical history, and the like.

 In general, it will be advantageous to provide the recipient mammal with a dosage of antibodies in the range of from about 1 mg/kg body weight to about 10 mg/kg body weight of the mammal, although a lower or higher dose may be administered if found desirable. Such antibodies will normally be administered by intravenous or intramuscular route as an inoculum. The antibodies of the present invention are intended to be provided to the recipient subject in an amount sufficient to prevent, lessen or attenuate the severity, extent or duration of any existing infection.

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° The antibodies prepared by use of the nucleic acid sequences, viruses or polypeptides of the present invention are also highly useful for diagnostic purposes. For example, the antibodies can be used as in
5 vitro diagnostic agents to test for the presence of HCV in biological samples taken from animals, especially humans. Such assays include, but are not limited to, radioimmunoassays, EIA, fluorescence, Western blot analysis and ELISAs. In one such embodiment, the
10 biological sample is contacted with antibodies of the present invention and a labeled second antibody is used to detect the presence of HCV to which the antibodies are bound.

15 Such assays may be, for example, direct where the labeled first antibody is immunoreactive with the antigen, such as, for example, a polypeptide on the surface of the virus; indirect where a labeled second
20 antibody is reactive with the first antibody; a competitive protocol such as would involve the addition of a labeled antigen; or sandwich where both labeled and unlabeled antibody are used, as well as other protocols well known and described in the art.

25 In one embodiment, an immunoassay method would utilize an antibody specific for HCV envelope determinants and would further comprise the steps of contacting a biological sample with the HCV-specific
30 antibody and then detecting the presence of HCV material in the test sample using one of the types of assay protocols as described above. Polypeptides and antibodies produced according to the present invention may also be supplied in the form of a kit, either
35 present in vials as purified material, or present in

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° compositions and suspended in suitable diluents as previously described.

In a preferred embodiment, such a diagnostic test kit for detection of HCV antigens in a test sample
5 comprises in combination a series of containers, each container a reagent needed for such assay. Thus, one such container would contain a specific amount of HCV-specific antibody as already described, a second
10 container would contain a diluent for suspension of the sample to be tested, a third container would contain a positive control and an additional container would contain a negative control. An additional container could contain a blank.

15 For all prophylactic, therapeutic and diagnostic uses, the antibodies of the invention and other reagents, plus appropriate devices and accessories, may be provided in the form of a kit so as to facilitate ready availability and ease of use.

20 The present invention also relates to the use of nucleic acid sequences and polypeptides of the present invention to screen potential antiviral agents for antiviral activity against HCV. Such screening
25 methods are known by those of skill in the art.

Generally, the antiviral agents are tested at a variety of concentrations, for their effect on preventing viral replication in cell culture systems which support viral
30 replication, and then for an inhibition of infectivity or of viral pathogenicity (and a low level of toxicity) in an animal model system.

In one embodiment, animal cells (especially human cells) transfected with the nucleic acid sequences
35 of the invention are cultured in vitro and the cells are

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° treated with a candidate antiviral agent (a chemical, peptide etc.) by adding the candidate agent to the medium. The treated cells are then exposed, possibly under transfecting or fusing conditions known in the art, to the nucleic acid sequences of the present invention. A sufficient period of time would then be allowed to pass for infection to occur, following which the presence or absence of viral replication would be determined versus untreated control cells by methods known to those of ordinary skill in the art. Such methods include, but are not limited to, the detection of viral antigens in the cell, for example, by immunofluorescence procedures well known in the art; the detection of viral polypeptides by Western blotting using antibodies specific therefor; the detection of newly transcribed viral RNA within the cells by RT-PCR; and the detection of the presence of live, infectious virus particles by injection of cell culture medium or cell lysates into healthy, susceptible animals, with subsequent exhibition of the signs and symptoms of HCV infection. A comparison of results obtained for control cells (treated only with nucleic acid sequence) with those obtained for treated cells (nucleic acid sequence and antiviral agent) would indicate, the degree, if any, of antiviral activity of the candidate antiviral agent. Of course, one of ordinary skill in the art would readily understand that such cells can be treated with the candidate antiviral agent either before or after exposure to the nucleic acid sequence of the present invention so as to determine what stage, or stages, of viral infection and replication said agent is effective against.

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In an alternative embodiment, viral enzyme
such as NS3 protease, NS2-NS3 protease, NS3 helicase or
NS5B RNA polymerase may be produced from a nucleic acid
sequence of the invention and used to screen for
5 inhibitors which may act as antiviral agents. The
structural and nonstructural regions of the HCV genome,
including nucleotide and amino acid locations, have been
determined, for example, as depicted in Houghton, M.
(1996), Fig. 1; and Major, M.E. et al. (1997), Table 2.

10 Such above-mentioned protease inhibitors may
take the form of chemical compounds or peptides which
mimic the known cleavage sites of the protease and may
be screened using methods known to those of skill in the
15 art (Houghton, M. (1996) and Major, M.E. et al. (1997)).
For example, a substrate may be employed which mimics
the protease's natural substrate, but which provides a
detectable signal (e.g. by fluorimetric or colorimetric
20 methods) when cleaved. This substrate is then incubated
with the protease and the candidate protease inhibitor
under conditions of suitable pH, temperature etc. to
detect protease activity. The proteolytic activities of
the protease in the presence or absence of the candidate
25 inhibitor are then determined.

In yet another embodiment, a candidate
antiviral agent (such as a protease inhibitor) may be
directly assayed in vivo for antiviral activity by
30 administering the candidate antiviral agent to a
chimpanzee transfected with a nucleic acid sequence of
the invention or infected with a virus of the invention
and then measuring viral replication in vivo via methods
such as RT-PCR. Of course, the chimpanzee may be
35 treated with the candidate agent either before or after

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transfection with the infectious nucleic acid sequence or infected with a virus of the invention so as to determine what stage, or stages, of viral infection and replication the agent is effective against.

The invention also provides that the nucleic acid sequences, viruses and polypeptides of the invention may be supplied in the form of a kit, alone or in the form of a pharmaceutical composition.

All scientific publication and/or patents cited herein are specifically incorporated by reference. The following examples illustrate various aspects of the invention but are in no way intended to limit the scope thereof.

EXAMPLES

Materials and Methods

Source of HCV

An infectious plasma pool of HCV genotype 2a (HC-J6_{CH}) prepared from acute phase plasma of a chimpanzee experimentally inoculated with plasma from a Japanese patient infected with strain HC-J6 (Okamoto et al., 1991) was used for cloning. An infectious cDNA clone of HCV strain H77, genotype 1a was also used (pCV-H77C; Yanagi et al., 1997).

Amplification, cloning and sequence analysis

Viral RNA was extracted from 100 µl aliquots of the HC-J6_{CH} plasma pool with the TRIzol system (GIBCO/BRL) (Yanagi et al., 1997). Primers used in cDNA synthesis and PCR amplification were based on the genomic sequence of strain HC-J6 (Okamoto et al., 1991) and from the conserved region (3'X) of the 3' UTR of HCV genotype 2a (Tanaka et al., 1996) (Table 1). The RNA

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was denatured at 65°C for 2 min, and cDNA was synthesized at 42°C for 1 hour with Superscript II reverse transcriptase (GIBCO/BRL) and specific reverse primers in 20 µl reaction volumes. The cDNA mixtures were treated with RNase H and RNase T1 (GIBCO/BRL) at 37°C for 20 min.

TABLE 1

Oligonucleotides used for amplification and cloning of strain HC-J6_{CH}, genotype 2a

Designation	Sequence (5' → 3')a
2427S-H77	ACTGGACACGGAGGTGGCCGCGTC
2426S-H77	TTGTTCTTGTCGGGTTAATGGCGC
2645R-H77	GGGTGTACTACACACATGAGTAAG
2832R-H77	AAGCGCCCTAACTGATGATG
H2751SII	CGTCATCGATA CCTCAGCGGGCATATGCACTGGACACGGA
H2786R	GTCCAGTGCAATGCCCCGCTGAGG
H2870R	CATGCACCAGCTGATATAGCGCTTGTAAATATG
H7851S	TCCGTAGAGGAAGCTTGCAGCCTGACGCCC
H9140S (M)	CAGAGGAGGCAGGGTGCTATATGTGGCAAGTAC
H9173R (M)	GTA CTTGCCACATATAGCAGCCCTGCCTCCTCTG
H9471R	CGTCTCTAGAC AGAAATGGCTTAAGAGGCCGAGTGTTACC
J6-H2556S	TTATGGATGCTCATCTTGTTGGGCCAGGCCGAAGCAGCTTGGAGAACCTCGTAATACTCAATGC
356RF-J6H	AGGATTGTGCTCATGGTGACGGTCTACGAG
1S-J6F ^b	TTTTTTTTCGGCCGC TAATACGACTCACTATAGACCCGCCCTAATAGG
333S-J6	CCGTGCACCATGAGCACAAATCCTAAACCTC
753R-J6	GGATGTACCCCATGAGGTCGGCAAAG
2543S-J6F	GTTTGCGCCTGCTTATGGATGCTCATCTTG
2787R-J6 (26)	GCGTCATAAGCATATGCTGTTGGGG
3329R-J6	CCCTCAGCACTGGAGTACATCTG
5487-J6F	CGTCATGCATA CCCTAGGGCGGCTCTCATTTGAAGAGGG
5518R-J6F	CGTCCCCTCTTCAATGAGAGCCGCTCTAGA
9251S-J6F	GCGGTGAAGACCAAGCTCAAACCTCACTC
9305R-J6F	AATCTAGA AGGCGCGCTTCGGGCAATGGAGTGAGTTTGAGC
9310R-J6F	CGTCTCTAGAG GATAAATCCAGGAGGCGGCTTCCGGC
9399S-J6F	TACTTTTGTAGGGGTAGGCCTTTTCC
9464-J6F	CGTCTCTAGAG TGTAGCTAATGTGTGCCGCTCTA
9470 (24)-J6	CTATGGAGTGTAGCTAATGTGTGC
J6-3' XR	CGTCTCTAGAC ATGATCTGCAGAGAGACCAGTTACGGCACTCTCTGFCAGTCATGCGGC
	TCACGGACCTTTCACAGCTAGCCGTGACTAGGGCTAAGATGGAGCCACC

a HCV-specific sequences are shown in plain text, non HCV-specific sequences are shown in bold face, and cleavage sites used for cDNA cloning are underlined.

b The core sequence of the T7 promoter is shown in italics.

The strategy used to amplify and clone the full-length HC-J6_{CH} sequence is shown in Fig. 1.

Nucleotide positions correspond to those of the 2a

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° infectious clone (pJ6CF) that is described herein. The 5' end of HC-J6_{CH} (nts. 17-297, excluding primer sequences) was amplified from 2 µl of cDNA synthesized with primer a-2 (Yanagi et al., 1996). PCR was performed with *AmpliTaq Gold* DNA polymerase (Perkin-Elmer) as described previously (Yanagi et al., 1996) using primers 1S-J6F and a-2. After purification, the amplified products were cloned into pGEM-T Easy vector (Promega) using standard procedures and 5 clones (pJ6-5'UTR) were sequenced.

The 3' end of HC-J6_{CH} was amplified in 3 overlapping pieces. RT-PCR of a short fragment of NS5B (nts. 9279-9439) was performed with primers 9251S-J6F and 9464R-J6F as described above. The PCR products were cloned into pGEM-T Easy vector and sequence analysis was performed from 5 pJ6-3'F clones. A second region spanning from NS5B to the conserved region of the 3' UTR (nts. 9376-9629) was amplified in RT-nested PCR (external primers H9261F and H3'X58R, internal primers H9282F and H3'X45R) (Yanagi et al., 1997). The amplified products were cloned into pGEM-9zf(-) by using *HindIII* and *XbaI* sites and 14 pJ6-3'VR clones were sequenced. The third fragment, which included the 3' terminal sequence was amplified with primers 9399S-J6F and J6-3'XR from one of the pJ6-3'VR clones, and cloned into one of the pJ6-3'F clones by using *StuI* and *XbaI* sites (pJ6-3'X).

The ORF of HCV HC-J6_{CH} was amplified by long RT-PCR in 3 overlapping pieces. The amplification was performed on 2 µl of the cDNA mixtures with the Advantage cDNA polymerase mix (Clontech) (Yanagi et al., 1997). The J6S fragment (nts. 86-2761) was amplified

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with primers a-1 (Yanagi et al., 1996) and J6-2787R from
cDNA synthesized with primer J6-3329R. A single PCR
round was performed in a Robocycler thermal cycler
(Stratagene), and consisted of denaturation at 99°C for
5 35 sec, annealing at 67°C for 30 sec and elongation at
68°C for 4 min 30 sec during the first 5 cycles, 5 min
during the next 10 cycles, 5 min 30 sec during the
following 10 cycles and 6 min during the last 10 cycles.
10 The J6B fragment (nts. 2573-5488) was amplified with
primers 2543S-J6F and 5518R-J6F from cDNA synthesized
with primer 5518R-J6F. Finally, the J6A fragment (nts.
5515-9282) was amplified with primers 5487S-J6F and
9310R-J6F from cDNA synthesized with primer
15 9470R(24)-J6F. PCR amplifications of fragments J6B and
J6A consisted of denaturation at 99°C for 35 sec,
annealing at 67°C for 30 sec and elongation at 68°C for 6
min during the first 5 cycles, 7 min during the next 10
cycles, 8 min during the following 10 cycles and 9 min
20 during the last 10 cycles.

After purification of the long PCR products
with QIAquick PCR purification kit (QIAGEN), A-tailing
reactions were performed with *AmpliTaq* DNA polymerase
25 (Perkin Elmer) at 72 °C for 1 hour. The gel-purified
A-tailed PCR products were cloned into pCR2.1 vector
(Invitrogen) or pGEM-T Easy vector (Promega). DH5-alpha
competent cells (GIBCO BRL) were transformed and
30 selected on LB agar plates containing 100 µg/ml
ampicillin (SIGMA) and amplified in LB liquid cultures
at 30°C for 18 - 20 hrs (Yanagi et al., 1997). Midiprep
was performed using Wizard *Plus* Midipreps DNA
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Purification System (Promega). Multiple clones of the J6S, J6A and the J6B fragments were sequenced.

The consensus sequence of strain HC-J6_{CH} (nts. 17-9629) was determined by direct sequencing of PCR products (nts. 297-3004 and nts. 4893-5762) and by sequence analysis of the TA clones (nts. 17-5488 and nts. 5515-9629) (Fig. 1). Both strands of DNA were sequenced in all cases. Analyses of genomic sequences, including multiple sequence alignments and tree analyses, were performed with GeneWorks (Oxford Molecular Group) (Bukh et al., 1995).

Construction of chimeric cDNA clones of genotypes 1a & 2a

Four full-length intertypic chimeric cDNA clones were constructed (Figs. 4, 5A, 5B). In each clone the C, E1 and E2 genes encoded the consensus amino acid sequence of HC-J6_{CH}. The p7 protein was encoded either by the HC-J6_{CH} or pCV-H77C consensus sequence, and the NS proteins were all encoded by pCV-H77C genes. To engineer these cDNA clones, an NdeI site from pCV-H77C was first eliminated by a silent substitution (C to T) at position 9158. In brief, two fragments were amplified from pCV-H77C with primers H7851S and H9173R(M) and with primers H9140S(M) and H9417R (Table 3), gel-purified and used for fusion PCR with primers H7851S and H9417R. The fusion PCR products were cloned into pCV-H77C by using HindIII and AflIII sites. A new artificial NdeI site was introduced by a silent substitution (C to T) at position 2765. PCR products, which were amplified from pCV-H77C with primer H2751SII containing artificial ClaI and NdeI sites and primer H2870R, were cloned into the modified pCV-H77C by using

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*Cla*I and *Eco*47III sites. The final construct (pH77CV) was used as a cassette vector to construct the intertypic chimeric HCV cDNA clones.

The four chimeric cDNA clones were constructed as follows. pH77CV-J6S (nucleotide sequence shown in SEQ ID No:3 and amino acid sequence shown in SEQ ID No:4): The *Age*I/*Bsm*I fragment of clone J6S2 and the *Bsm*I/*Nde*I fragment of clone J6S1, were cloned into pH77CV by using *Age*I and *Nde*I sites; pH77 (p7)CV-J6S (nucleotide sequence shown in SEQ ID No:5 and amino acid sequence shown in SEQ ID No:6): A fragment of pH77CV-J6S was replaced with a fragment amplified from pCV-H77C with primers J6-H2556S and H2786R by using *Bsa*BI and *Nde*I sites; J6S (nucleotide sequence shown in SEQ ID No:7 and amino acid sequence shown in SEQ ID No:8): A fragment amplified from pH77pCV-H77C with primers a-1 and 356RF-J6H77 and another fragment amplified from pH77CV-J6S with primers 333S-J6 and 753R-J6 were gel-purified and a fusion-PCR was performed with primers a-1 and 753R-J6. The *Age*I/*Cla*I fragment of the subcloned fusion PCR products and the *Cla*I/*Nde*I fragment of pH77CV-J6S were cloned into pH77CV-J6S by using *Age*I and *Nde*I sites; pH77 (p7)-J6S (nucleotide sequence shown in SEQ ID No:9 and amino acid sequence shown in SEQ ID No:10): The *Age*I/*Cla*I fragment of J6S and the *Cla*I/*Nde*I fragment of (p7)CV-J6S were cloned into pH77 (p7)CV-J6S by using *Age*I and *Nde*I sites.

Each intertypic chimeric cDNA clone was retransformed to select a single clone, and large-scale preparation of plasmid DNA was performed with a QIAGEN plasmid Maxi kit as described previously (Yanagi et al., 1997). Each of the four cDNA clones was completely

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sequenced before inoculation. Each clone was genetically stable since the digestion pattern was as expected following retransformation and the complete sequence was the expected one.

5 Construction of full-length cDNA clone HC-J6_{CH}

An overview of the full-length HC-J6_{CH} clone is presented in Fig. 1. In the final construct pJ6CF, which encodes the consensus polyprotein of HC-J6_{CH}, an XbaI site was eliminated by a silent substitution (A to G) at position 5494. Digested fragments containing the consensus sequence were purified from the appropriate subclones and ligated using the sites indicated. The full-length cDNA clone (pJ6CF) was retransformed to select a single clone, and large-scale preparation of plasmid DNA followed by the complete sequence analysis was performed. Clone pJ6CF was genetically stable.

20 Intrahepatic transfection of chimpanzee with transcribed RNA

In duplicate 100 µl reactions, RNA was transcribed *in vitro* with T7 RNA polymerase (Promega) from 10 µg of template plasmid linearized with XbaI (Promega) as described previously (Yanagi et al., 1997). The integrity of the RNA was checked by electrophoresis through agarose gel stained with ethidium bromide (Yanagi et al., 1997). Each transcription mixture was diluted with 400 µl of ice-cold phosphate-buffered saline without calcium or magnesium and then immediately frozen on dry ice and stored at -80°C. Within 24 hours, both transcription mixtures were injected into the same chimpanzee by percutaneous intrahepatic injection guided by ultrasound (Yanagi et al., 1998, 1999). If the

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chimpanzee did not become infected, the same transfection was repeated once. After two negative results, the next clone was inoculated into the same chimpanzee following the same protocol. Injections were performed at weeks 0 and 2 with pH77CV-J6S, at weeks 5 and 8 with pH77(p7)CV-J6S, at weeks 14 and 16 with pH77-J6S, at weeks 19 and 23 with pH77(p7)-J6S, at week 28 with pJ6CF, and finally at week 34 with pCV-H77C. The chimpanzee was maintained under conditions that met or exceeded all requirements for its use in an approved facility.

Serum samples were collected weekly from the chimpanzee and monitored for liver enzyme levels by standard procedures, anti-HCV antibodies by the second-generation ELISA (Abbott) and HCV RNA by a sensitive RT-nested PCR assay with *AmpliTag Gold* DNA polymerase using primers from the 5' UTR (Yanagi et al., 1996). Samples were scored as negative for HCV RNA if two independent tests on 100 µl of serum were negative. The genome equivalent (GE) titer of HCV in positive samples was determined by RT-nested PCR on 10-fold serial dilutions of the extracted RNA (Bukh et al., 1998). The consensus sequence of the complete ORF from the chimpanzee infected with RNA transcripts of pJ6CF was determined by direct sequencing of overlapping PCR products obtained by long RT-nested PCR as previously described (Yanagi et al., 1997) with HC-J6 specific primers. After the intrahepatic transfection with RNA transcripts of pCV-H77C, we performed H77(genotype 1a)-specific RT-nested PCR with primers 2427S-H77 and 2832R-H77 for the 1st round and with primers 2462S-H77 and 2645R-H77 for the 2nd round (Table 3). The

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° sensitivity of this assay was equivalent to that of the assay using 5' UTR primers when testing serum containing only H77, genotype 1a. The genome titer of genotype 1a was determined by using this specific RT- nested PCR on
5 10-fold serial dilutions of the extracted RNA.

EXAMPLE 1

Sequence analysis of HCV strain HC-J6_{CH}

10 As minor deviations from the consensus amino acid sequence were found previously to render full-length HCV cDNA clones noninfectious (Yanagi et al., 1997, 1998), the consensus sequence of the cloning source of genotype 2a (strain HC-J6_{CH}) was determined
15 prior to constructing any full-length clones. In brief, a plasma pool containing strain HC-J6_{CH} was prepared from acute phase plasmapheresis units collected from a chimpanzee experimentally infected with HC-J6 (Okamoto et al., 1991). The HCV genome titer of this pool was
20 10^{5.4} genome equivalents (GE)/ml (Quantiplex HCV RNA bDNA 2.0, Chiron) and the infectivity titer was 10⁴ chimpanzee infectious doses/ml.

25 The consensus sequence of the 5' UTR of HC-J6_{CH} (nts. 17-340) was deduced from 5 clones containing nts. 17-297 and 8 clones containing nts. 86-340. The 5' UTR of the various clones was highly conserved, but the consensus sequence of HC-J6_{CH} differed by 2 nucleotides from that published previously for HC-J6 (Okamoto et
30 al., 1991: C to T at position 36 and T to C at position 222).

35 The consensus sequence of 14 clones of the 3' UTR of HC-J6_{CH} indicated that the 39 nucleotide long variable region was highly conserved in this strain and

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° was identical to that previously published for HC-J6 (Okamoto et al., 1991). The polypyrimidine tract varied greatly in length (84-164 nucleotides), and contained some conserved A residues. In the conserved region, the proximal 16 nucleotides were identical to those previously published for isolates of different HCV genotypes (Kolykhalov et al., 1996; Tanaka et al., 1996; Yamada et al., 1996). The remaining 82 nucleotides of the conserved region were determined for other genotype 2a strains (Tanaka et al., 1996) but not for HC-J6 or HC-J6_{CH}.

The ORF of HC-J6_{CH} was amplified in 3 fragments by RT-PCR (Fig. 1). Eight clones of the J6S fragment (nts. 86-2761), 6 clones of the J6B fragment (nts. 2573-5488) and 6 clones of the J6A fragment (nts. 5515-9298) were sequenced. PCR fragments containing nts. 5489-5514 were sequenced directly. A quasispecies was found at 243 nucleotide (2.7%) and 69 amino acid (2.3%) positions, scattered throughout the 9099 nts (3033 aa) of the ORF. However, the majority, 231 nucleotide substitutions, were detected only once and 71.6 % of these represented silent mutations. The 12 remaining nucleotide substitutions were each restricted to 2 clones and only 4 of these resulted in amino acid changes. The nucleotide difference among the J6S clones ranged from 0.1 - 1.3%, among the J6B clones it ranged from 0.1 - 0.3%, and it ranged from 0.2 - 4.0% among the J6A clones (Fig. 2). Three of 8 J6S clones, 4 of 6 J6B clones, and all 6 J6A clones had defective polyproteins due to nucleotide deletions, insertions or substitutions.

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• The sequences of clones of strain HC-J6_{CH} were relatively homogeneous. This was highlighted by the high degree of conservation among clones of the HVR1 (Fig. 3), a region frequently used to study the quasispecies of HCV (Bukh et al., 1995). An exception was the sequence of clone J6A1, which differed by about 4% from the other clones of this region (Fig. 2). Importantly, the consensus sequence of strain HC-J6_{CH} (nts. 17-9629) could be determined with no ambiguity at the nucleotide or deduced amino acid level. The difference between the consensus ORF sequence of HC-J6_{CH} from the experimentally infected chimpanzee and that of HC-J6 of the inoculum (Okamoto et al., 1991) was 4.1 % and 2.2 % at the nucleotide and deduced amino acid levels, respectively (Fig. 2, Table 2). Moreover, we found that 12 (44.4%) of the 27 amino acids constituting HVR1 differed between HC-J6_{CH} and HC-J6 (Fig. 3). Such diversities are greater than the < 2 % generally considered to comprise a quasispecies. In fact, these differences are equivalent to those found between the two prototype strains of HCV genotype 1a [strains HCV-1 (Choo et al., 1991) and H77 (Yanagi et al., 1997)]. These results indicated that HC-J6_{CH}, which represented the major species in the experimentally infected chimpanzee, was a minor species in the original inoculum.

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- 40 -

TABLE 2

Percent difference of nucleotide and predicted amino acid sequences between strain HC-J6 (Okamoto et al., 1991) and strain HC-J6_{CH} from acute phase plasma pool of a chimpanzee inoculated with HC-J6

Genome Region	nt.position ^a	% nt. difference	% a.a. difference
ORF	341-9439	4.1 (373/9099) ^b	2.2 (66/3033) ^b
5' UTR	17-340	0.6 (2/324)	
Core	341-913	0.5 (3/573)	0 (0/191)
E1	914-1489	4.3 (25/576)	2.1 (4/192)
HVR1	1490-1570	24.7 (20/81)	44.4 (12/27)
E2-HVR1	1571-2590	3.9 (40/1020)	3.2 (11/340)
p7	2591-2779	3.7 (7/189)	3.2 (2/63)
NS2	2780-3430	4.0 (26/651)	2.8 (6/217)
NS3	3431-5323	4.0 (76/1893)	0.8 (5/631)
NS4A	5324-5485	4.3 (7/162)	1.9 (1/54)
NS4B	5486-6268	3.7 (29/783)	0.4 (1/261)
NS5A	6269-7666	5.4 (75/1398)	3.4 (16/466)
NS5B	7667-9439	3.7 (65/1773)	1.4 (8/591)
3' UTR	9440-9481	0 (0/42)	

a The nucleotide positions correspond to those of the infectious full-length genotype 2a clone (pJ6CF).

b The numbers in parenthesis indicate the nucleotide or amino acid differences for each region.

Example 2

Chimeric molecular clones

As chimeric flaviviruses with substituted structural genes have been useful in defining the biological function of viral sequences or proteins, in analyzing immune responses and in generating attenuated vaccine candidates (Bray and Lai, 1991; Chambers et al., 1999; Pletnev et al., 1992, 1993, 1998). The consensus sequence of the 2a structural genes and surrounding region was substituted for that of the infectious 1a cDNA clone. In the genotype 1a backbone, two silent mutations were introduced for cloning purposes [at positions 2765 (p7) and 9158 (NS5B) of pCV-H77C] (Fig. 4). The complete sequence of each chimera was verified. Infectivity of RNA transcripts from four different

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intertypic chimeric clones (Figs. 4, 5A, 5B) was evaluated by consecutive intrahepatic transfections of a chimpanzee. Clones were considered not to be viable if viral RNA was not detected in the serum within two weeks of the repeat transfection. All chimeric clones contained the C, E1 and E2 genes of genotype 2a. The two chimeric clones tested initially differed from each other in that one had the p7 gene of 2a (pH77CV-J6S) and the other [pH77(p7)CV-J6S] the p7 gene of 1a. They differed from the two other clones in that the 186 nucleotides of the 5' UTR just upstream of the initiation codon were from the 2a genotype. Since neither clone containing the chimeric 5' UTR was infectious, the chimeric 5' UTR was replaced with the consensus genotype 1a 5' UTR to generate the two p7 varieties [pH77-J6S and pH77(p7)-J6S]. After consecutive transfection of the four clones, no HCV RNA, anti-HCV or ALT elevation was detected in the chimpanzee during 28 weeks of follow-up, suggesting that RNA transcripts from these intertypic chimeric clones were not viable *in vivo*.

This finding that the intertypic clones between genotypes 1a and 2a were not viable was surprising since flavivirus chimeras containing the structural region of dengue virus type 1 or 2 or of tick-borne encephalitis virus and the nonstructural region of an infectious dengue type 4 virus were viable (Bray and Lai, 1991; Pletnev et al., 1992, 1993). While considerable sequence variation exists between the infectious genotype 1a and 2a clones of HCV (Table 3), these viruses exhibit a higher degree of genetic heterogeneity than do the major genotypes of HCV. For other flaviviruses, however, it was possible to obtain

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infectious chimeric clones only if the capsid region was derived from the backbone cDNA clone (Chambers et al., 1999; Pletnev and Men, 1998).

TABLE 3

Percent difference of the amino acid sequences between the infectious clone of genotype 1a (pCV-H77C; Yanagi et al., 1997) and the infectious clone of genotype 2a (pJ6CF) of hepatitis C virus

Genome Region ^a	% difference
Polyprotein	27.9 (839/3007) ^b
Core	8.9 (17/191)
E1	37.0 (71/192)
HVR1	59.3 (16/27)
E2-HVR1	27.1 (91/336)
p7	38.1 (24/63)
NS2	41.9 (91/217)
NS3	19.2 (121/631)
NS4A	33.3 (18/54)
NS4B	26.8 (70/261)
NS5A	38.5 (171/444)
NS5B	25.2 (149/591)

^a Genome regions defined as in Table 1.

^b The numbers in parenthesis indicate the amino acid differences for each region. Positions with deletions or insertions in E2 (4 aa positions) and NS5A (26 aa positions) were not considered.

Trivial explanations may account for the lack of viability of these intertypic chimeras. First, the two silent mutations introduced in the genotype 1a backbone (one in p7 and one in NS5B) for cloning purposes could potentially eliminate infectivity. This is, however, very unlikely since mutations at these positions exist among field isolates of HCV including strain HC-J6_{CH} (Bukh et al., 1998). Also, it is noteworthy that the three previously published infectious clones of strain H77 had numerous silent nucleotide differences (Hong et al., 1999; Kolykhalov et al., 1997; Yanagi et al., 1997). Second, signal peptidases might not cleave the chimeric E2/p7 or p7/NS2

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junction. This seems unlikely, however, since
eukaryotic signal peptidases typically recognize the
amino acid sequences upstream of the cleavage site [the
(-3, -1) rule] (Nielsen et al., 1997) and the amino
acids at these two sites are conserved between genotypes
1a and 2a (Fig. 5B). Finally, the E2/p7 and/or p7/NS2
gene junctions could differ between genotypes 1a and 2a.
The junctions determined for genotypes 1a and 1b were
used (Lin et al., 1994; Mizushima et al., 1994; Selby et
al., 1994) because those for genotype 2a have not been
identified. In the latter two cases, further analyses
of genotype 2a should eventually provide sufficient data
to overcome such potential problems and it would most
likely be possible to construct a viable chimera.

More complicated explanations for the lack of
viability of the chimeras might be required if critical
genotype-specific interactions occur as regards the
structural proteins, the nonstructural proteins and the
genomic RNA. For instance, one cannot rule out that the
chimeras were not viable because the IRES function was
compromised. In *in vitro* studies the IRES activity
depended on RNA sequences not only in the 5' UTR but
also extending 3' of the translation initiation site
(Hahm et al., 1998; Lemon and Honda, 1997; Reynolds et
al., 1995). Although the 3' border of the HCV IRES is
still controversial it is believed to involve at most
the first 39 nts of the core gene (Lemon and Honda,
1997). The 5' UTR of the intertypic chimeras was either
a chimera of genotype 1a and 2a sequences or the entire
5' UTR was derived from the 1a clone (Figs. 4, 5A).
Importantly, the 5' end of core is conserved among
genotypes 1a and 2a (Fig. 5A). Thus, the predicted

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- ° IRES-like secondary structure is maintained in these chimeras, suggesting that the IRES activity most likely was maintained.

Possible interactions between the structural proteins and the nonstructural proteins and/or the genomic RNA, which involve RNA packaging, replication or translation are conceivable. In poliovirus, which is another positive-sense RNA virus, functional coupling of RNA packaging to RNA replication and of RNA replication to translation have been suggested (Novak and Kirkegaard, 1994 ; Nugent et al., 1999). Similar to other viruses of the *Flaviviridae* family, a membrane-associated replicase complex is thought to initiate replication at the 3' end of HCV and to synthesize a complementary negative-strand RNA (Rice, 1996). The putative cis-acting elements at the 5' and 3' termini which are believed to be important for viral genome replication (Rice 1996; Frolov et al., 1998) should be maintained in the intertypic HCV chimeras at least in the two constructs with the authentic 1a 5'UTR. However, it is conceivable that the viral packaging system was interrupted (Frolov et al., 1998). Studies using a Kunjin flavivirus replicon system and providing the structural proteins *in trans* suggested that the essential encapsidation signals did not reside in the structural region of the genome (Khromykh et al., 1997, 1998). The location of the packaging signals of HCV is not known. However, if the structural proteins encapsidate viral RNA via genotype-specific sequences outside of the structural region, the chimeras would be unable to package the RNA and it might be extremely

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difficult to construct viable chimeras between highly divergent strains.

Example 3

5 A consensus molecular clone of
 genotype 2a is infectious in vivo

10 In order to prove that the genotype 2a portion
 used in the 4 intertypic chimeric cDNA clones indeed
 represented the infectious sequence, a consensus full-
15 length cDNA clone of HC-J6_{CH} (pJ6CF) was constructed.
 The core sequence of the T7 promoter, a 5' guanosine
 residue and the full-length sequence of HC-J6_{CH} (9711
 nts) were cloned into pGEM-9Zf vector using NotI/XbaI
20 sites. Within the HCV sequence there were no deduced
 amino acid differences and only 4 nucleotide differences
 (at nucleotide positions 1822, 5494, 9247 and 9289) from
 the consensus sequence of HC-J6_{CH} as determined in the
 present study. The silent mutation at position 1822 was
25 within the structural region and so was also present in
 the four intertypic chimeras. The 5' terminal 16 nts
 and the 3' terminal 82 nts were deduced from previously
 published HCV genotype 2a sequences (Okamoto et al.,
30 1991, Tanaka et al., 1996). The full-length cDNA clone
 of genotype 2a contained a 5' UTR of 340 nts, an ORF of
 9099 nts encoding 3033 amino acids and a 3' UTR
 consisting of a variable region of 39 nts followed by a
 132 nucleotide-long polypyrimidine tract interrupted
 with 3 A residues and the 3' terminal conserved region
 of 98 nts.

35 RNA transcripts from pJ6CF were injected into
 the same chimpanzee used for injection of the 4
 intertypic chimeras. The chimpanzee became infected at

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the first attempt with an HCV titer of 10^2 GE/ml at week 1 post inoculation (p.i.), and 10^3 - 10^4 GE/ml during weeks 2 to 6 p.i. The consensus sequence of PCR products of the complete ORF, amplified from serum obtained during week 5 p.i., was identical to the sequence of pJ6CF and there was no evidence of a quasispecies. Since RNA transcripts of this infectious genotype 2a clone were infectious *in vivo*, and it shared an exact sequence with the non-infectious intertypic chimeric clones, their failure to replicate must have been the result of incompatibilities between the genotype 1a and 2a sequences.

To confirm that the chimpanzee used was susceptible also to infection by genotype 1a, which comprised most of the intertypic chimeras, the chimpanzee was subsequently inoculated with RNA transcripts from the infectious genotype 1a clone (pCV-H77C). Serum samples were tested in an H77-specific RT-PCR assay to identify super-infection with genotype 1a. At week 1 p.i. the total HCV genome titer was 10^4 GE/ml and the H77-specific (1a) genome titer was 10^2 GE/ml. The H77-specific genome titer increased to 10^3 GE/ml at week 2 p.i., and reached 10^4 GE/ml during weeks 3-6 p.i. The consensus sequence of PCR products amplified with H77-specific primers at weeks 1-6 p.i. were found to be identical to that of pCV-H77C. However, the direct sequences of PCR products amplified with the 5' UTR primers at weeks 1-2 after inoculation of pCV-H77C were identical to that of pJ6CF indicating that the 2a genotype was still present and represented the majority species. These experiments confirmed that the inability of the intertypic 1a, 2a

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cDNA clones to infect the chimpanzee was not the result of protective immune responses in the chimpanzee but represented deficiencies intrinsic to the chimeras.

Discussion

The published infectious cDNA clones of HCV represent the two most important subtypes of genotype 1 (Hong et al., 1999; Kolykhalov et al., 1997; Yanagi et al., 1997, 1998). However, 5 more major genotypes of HCV are recognized. In the above Examples, the infectivity of a cDNA clone of a second major HCV genotype was demonstrated. As in previous studies, the infectivity of RNA transcripts was demonstrated *in vivo* by intrahepatic transfection of a chimpanzee. This new infectious clone (pJ6CF) encodes the consensus polyprotein of HCV strain HC-J6_{CH}, genotype 2a. Its encoded polyprotein differs from those of the infectious clones of genotypes 1a and 1b by approximately 30% (Table 2). Genotype 2 strains, in particular subtypes 2a and 2b, have a worldwide distribution and important differences between genotypes 1 and 2 with respect to pathogenesis and treatment were indicated in previous studies. The availability of an infectious clone representing a second major genotype of HCV should permit new ways of studying the molecular biology and immunopathology of this important and genetically quite different human pathogen.

The 5' and 3' UTRs of HCV are believed to be critical for viral replication, translation and viral packaging (Rice, 1996). The 5' 203 terminal nucleotides and the 3' 101 terminal nucleotides of the published infectious clones of genotypes 1a and 1b were identical.

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However, the sequences of UTRs of the genotype 2a clone differ from those of the genotype 1 clones. Overall, the 5' UTR of the genotype 2a clone has 17 nt differences and a single nucleotide deletion compared with the infectious clones of genotype 1a (Fig. 5A). Five of these differences and the deletion are within the first 30 nucleotides, whereas the remainder are found within the predicted IRES structure. Differences also exist between the 3' UTR of the genotype 2a clone and the clones of genotype 1a (Fig. 5B). The sequences of the variable region are very different. Recent study has shown this region is not critical for infectivity *in vivo* (Yanagi et al., 1999). Within the regions which are critical for infectivity *in vivo* (Yanagi et al., 1999), the 132 nucleotide-long polypyrimidine tract of the genotype 2a clone has 3 unique A residues interspersed and the 3' terminal conserved region of 98 nts has 4 nt differences within the 3' terminal stable stem-loop structure (Fig. 5B) (Kolykhalov et al., 1996; Tanaka et al., 1996). Since the 2a clone was infectious these sequence differences are apparently real and are compatible with infectivity. Further studies are required to determine whether these represent critical genotype-specific sequences.

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WHAT IS CLAIMED IS:

1. A purified and isolated nucleic acid molecule which encodes human hepatitis C virus of genotype 2a, said molecule capable of expressing said virus when transfected into cells.
2. The nucleic acid molecule of claim 1, wherein said molecule encodes the amino acid sequence of SEQ ID NO:2.
3. The nucleic acid molecule of claim 2, wherein said molecule comprises the nucleic acid sequence of SEQ ID NO:1.
4. A DNA construct comprising a nucleic acid molecule according to claim 1.
5. A DNA construct comprising a nucleic acid molecule according to claim 3.
6. An RNA transcript of the DNA construct of claim 4.
7. An RNA transcript of the DNA construct of claim 5.
8. A cell transfected with the DNA construct of claim 4.
9. A cell transfected with the DNA construct of claim 5.
10. A cell transfected with RNA transcript of claim 6.

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11. A cell transfected with RNA transcript of claim 7.
12. A hepatitis C virus polypeptide produced by the cell of claims 8 or 9.
- 5
13. A hepatitis C virus polypeptide produced by the cell of claims 10 or 11.
14. A hepatitis C virus produced by the cell of claims 8 or 9.
- 10
15. A hepatitis C virus produced by the cell of claims 10 or 11.
16. A hepatitis C virus whose genome comprises a nucleic acid molecule according to claim 1.
- 15
17. A hepatitis C virus whose genome comprises a nucleic acid molecule according to claim 3.
- 20
18. A method for producing a hepatitis C virus comprising transfecting a host cell with the RNA transcript of claims 6 or 7.
19. A polypeptide encoded by a nucleic acid sequence according to claim 1.
- 25
20. A polypeptide encoded by a nucleic acid sequence according to claim 3.
- 30
21. The polypeptide of claim 19, wherein said polypeptide is selected from the group consisting of NS3 protease, E1 protein, E2 protein or NS4 protein.

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22. The polypeptide of claim 20, wherein said polypeptide is selected from the group consisting of NS3 protease, E1 protein, E2 protein or NS4 protein.

23. A method for assaying candidate antiviral agents for activity against HCV, comprising:

- a) exposing a cell containing the hepatitis C virus of claims 16 or 17 to the candidate antiviral agent; and
- b) measuring the presence or absence of hepatitis C virus replication in the cell of step (a).

24. The method of claim 23, wherein said replication in step (b) is measured by at least one of the following: negative strand RT-PCR, quantitative RT-PCR, Western blot, immunofluoresence, or infectivity in a susceptible animal.

25. A method for assaying candidate antiviral agents for activity against HCV, comprising:

- a) exposing an HCV protease encoded by a nucleic acid sequence according to claims 1 or 3 or a fragment thereof to the candidate antiviral agent in the presence of a protease substrate; and
- b) measuring the protease activity of said protease.

26. The method of claim 25, wherein said HCV protease is selected from the group consisting of an NS3 domain protease, an NS3-NS4A fusion polypeptide, or an NS2-NS3 protease.

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27. An antiviral agent identified as having antiviral activity for HCV by the method of claim 23.

28. An antiviral agent identified as having antiviral activity for HCV by the method of claim 25.

29. Antibody to the polypeptide of claim 19.

30. Antibody to the polypeptide of claim 20.

31. Antibody to the hepatitis C virus of claim 16.

32. Antibody to the hepatitis C virus of claim 17.

33. A method for determining the susceptibility of cells *in vitro* to support HCV infection, comprising the steps of:

- a) growing animal cells *in vitro*;
- b) transfecting into said cells the nucleic acid of claim 1; and
- c) determining if said cells show indicia of HCV replication.

34. The method according to claim 33, wherein said cells are human cells.

35. A composition comprising a polypeptide of claim 19 suspended in a suitable amount of a pharmaceutically acceptable diluent or excipient.

36. A composition comprising a polypeptide of claim 20 suspended in a suitable amount of a pharmaceutically acceptable diluent or excipient.

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37. A composition comprising a nucleic acid molecule of claim 1 suspended in a suitable amount of a pharmaceutically acceptable diluent or excipient.

5

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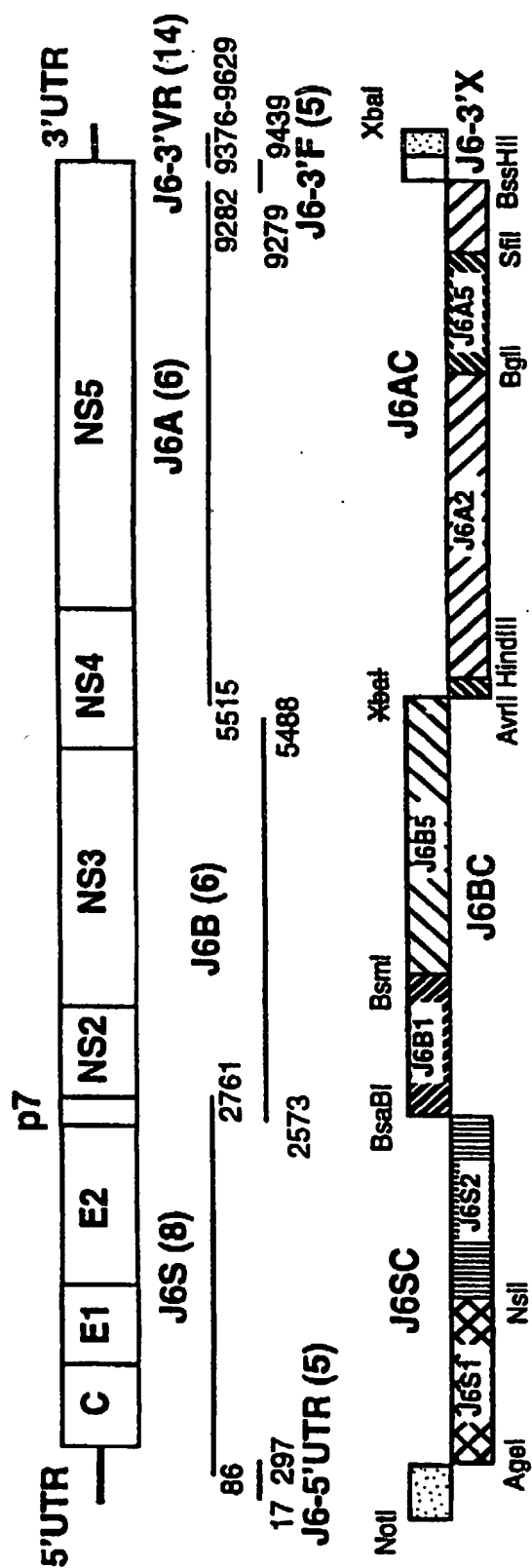
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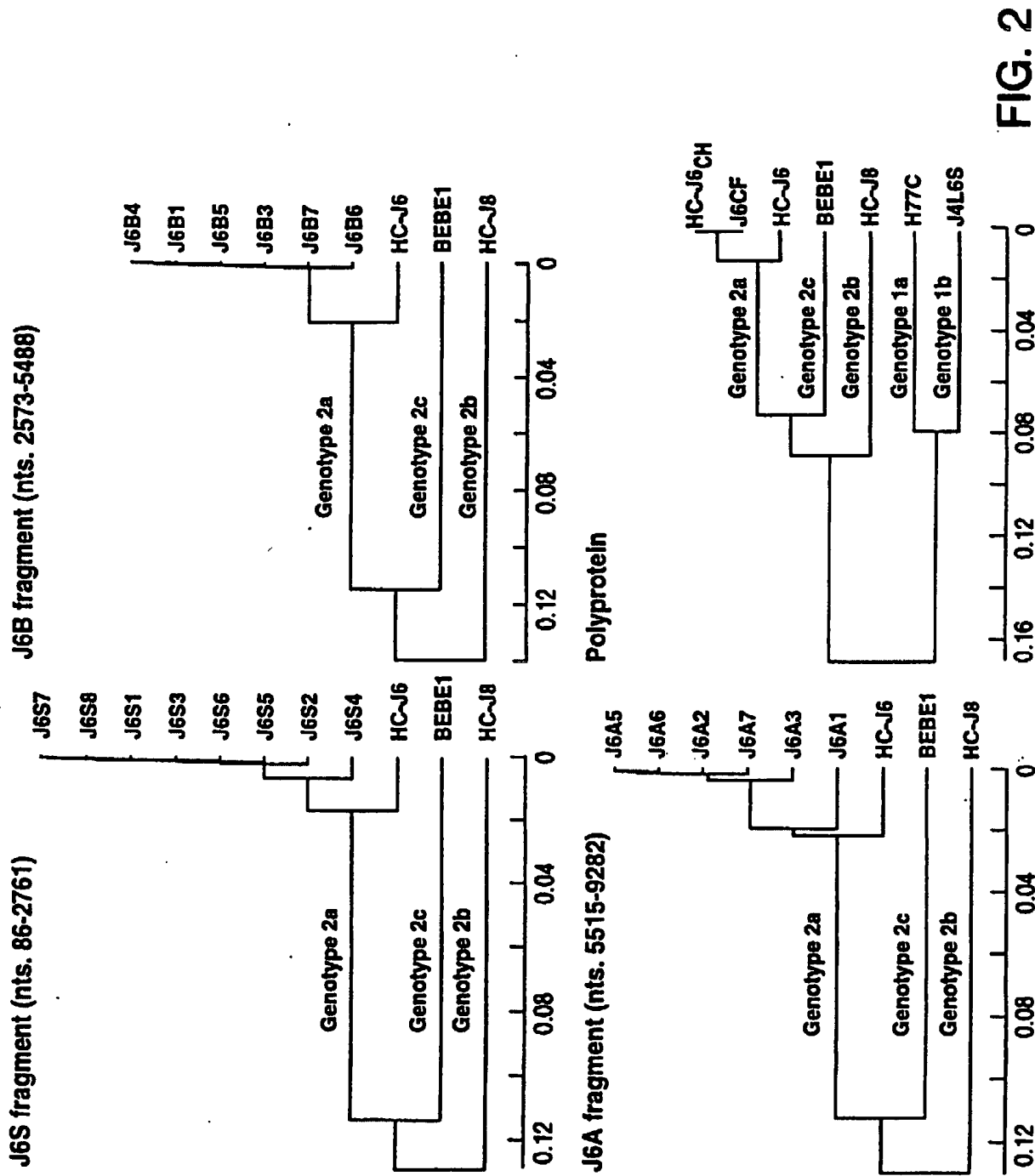
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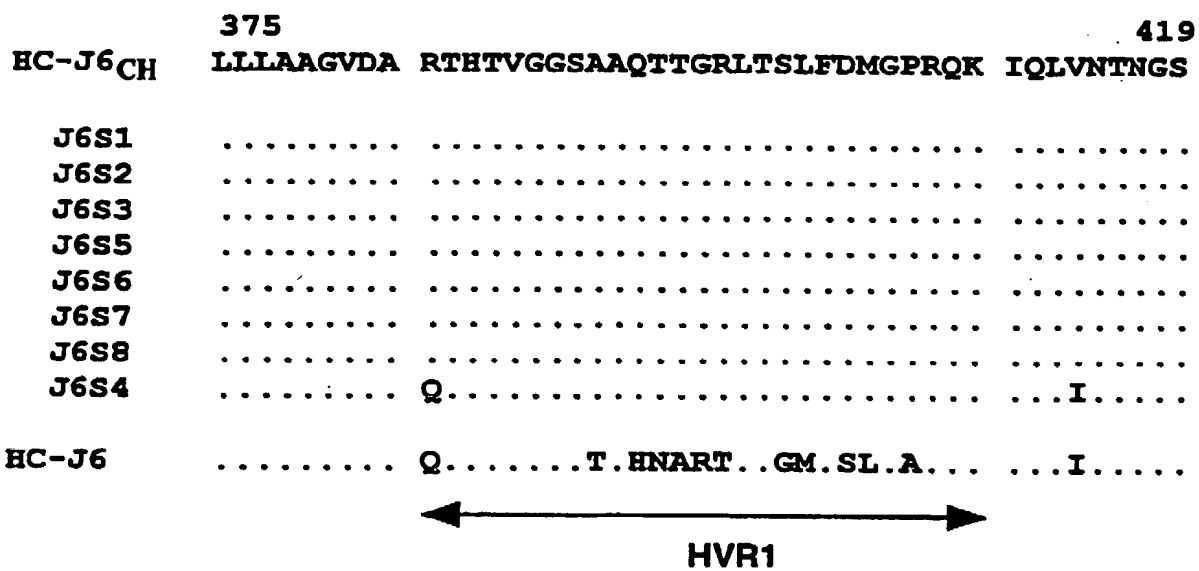


FIG. 3

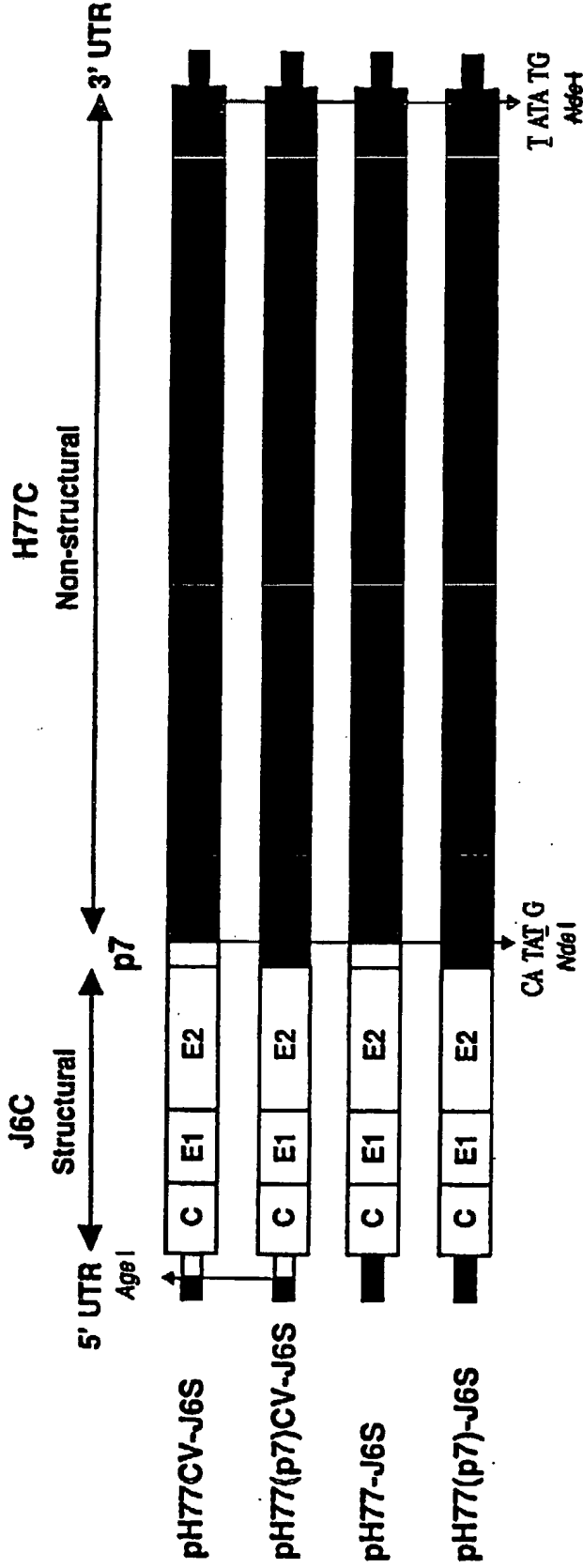


FIG. 4

FIG. 5B

3' Untranslated Region

9375
 H77C TGAAGTTGG GGTAAACACT CCGGCTCTT AAGCCATTTC CTG (Polypyrimidine tract)81 AATGCTGGCT CCATCTTAGC 9518
 H77CV-J6S
 H77(p7)CV-J6S (Polypyrimidine tract)81
 H77-J6S (Polypyrimidine tract)81
 H77(p7)-J6S (Polypyrimidine tract)81
 J6CF .AG..CGGCA CAC.TTAG.. A.ACT.CA.A GCTAAC.G.. .C- (Polypyrimidine tract)132 ---

9519
 H77C OCTAGTCACG GCTAGCTGTG AAAGTCCGT GAGCCGATG ACTGCAGAGA GTGCTGATAC TGGCTCTCT GCAGATCATG T 9599
 H77CV-J6S
 H77(p7)CV-J6S
 H77-J6S
 H77(p7)-J6S
 J6CFC.TA.. .T.....

E2/p7/NS2 Region

730
 H77C RVCSCLMMLLISQAEA ALENLVIILNAASLAGTHGLVSFLVFFCFAMYLKGRWPGVAVYALYGMWPLILLALLPQRAYA LDTEVAASCGGVVLVG 825
 H77CV-J6S ...A...LI.LG... ..K...H...A.SCN.FLY.VI..VA...I...V..L.T.S.T.L.SFS.....Q...
 H77(p7)CV-J6S ...A...LI.LG... ..K...H...A.SCN.FLY.VI..VA...I...V..L.T.S.T.L.SFS.....Q...
 H77-J6S ...A...LI.LG... ..K...H...A.SCN.FLY.VI..VA...I...V..L.T.S.T.L.SFS.....Q...
 H77(p7)-J6S ...A...LI.LG... ..K...H...A.SCN.FLY.VI..VA...I...V..L.T.S.T.L.SFS.....Q...
 J6CF ...A...LI.LG... ..K...H...A.SCN.FLY.VI..VA...I...V..L.T.S.T.L.SFS.....Q... Y.AS.HGQI.AAL..M

H77C

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
GCCAGCCCC	TGATGGGGC	GACACTCCAC	CATGAATCAC	TCCCCGTGA	50
GGAACTACTG	TCTTCACGCA	GAAAGCGICT	AGCCATGGCG	TTAGTATGAG	100
TGTCGTGCAG	CCTCCAGGAC	CCCCCCTCC	GGGAGAGCCA	TAGTGGTCTG	150
CGGAACCGGT	GAGTACACCG	GAATTGCCAG	GACGACCGGG	TCCTTTCTTG	200
GATAAACCCG	CTCAATGCT	GGAGATTGCG	GCGTGGCCCC	GCAAGACTGC	250
TAGCCGAGTA	GTGTGCGGTC	GCGAAAGGCG	TTGTGGTACT	GCTGTATAGG	300
GTGCTTGCGA	GTGCCCCGGG	AGGTCCTGTA	GACCGTGCAC	CATGAGCAAG	350
AATCCTAAAC	CTCAAAGAAA	AACCAACGCT	AACACCAACC	GTGCCCCACA	400
GGACGTCAAG	TTCCCCGGTG	GCGGTACAGT	CGTTGGTGGG	GTTTACTTGT	450
TGCCCGCGAG	GGGCCCCAGA	TGGGGTGTGC	GCGCGACGAG	GAAGACTTCC	500
GAGCGGTCCG	AACCTCGAGG	TAGAAGTCAG	CCTATCCCCA	AGGCAGCTCG	550
GCCCCAGGGC	AGGACCTGGG	CTCAGCCCCG	GTACCCCTTG	CCCCCTCTATG	600
GCAATGAGGG	TTGCGGGTGG	GCGGGATGGC	TCCTGTCTCC	CCGTGGCTCT	650
CGGCCCTAGCT	GGGGCCCCAC	AGACCCCCCG	CGTAGGTGCG	GCAATTTGGG	700
TAAGGTCATC	GATACCCCTA	CGTGCCTGCT	CGCCGACCTC	ATGGGGTACA	750
TACCGCTCGT	CGGGCCCCCT	CTTGGAGGCG	CTGCCAGGCG	CCTGGCGCAT	800
GGCGTCCGGG	TTCTGGAAGA	CGGCGTGAAC	TATGCAACAG	GGAACTTCC	850
TGGTGTGCTCT	TTCTCTATCT	TCCTTCTGGC	CCTGTCTCTCT	TGCTTACTG	900
TGCCCCGCTTC	AGCCTACCAA	GTGCGCAATT	CCTCGGGGCT	TTACCATGTC	950
ACCAATGATT	GCCCTAAGTC	GAGTATTGTG	TACGAGGCGG	CCGATGCCAT	1000
CCTGCACACT	CCGGGGTGTG	TCCCTTGGGT	TGCGAGGGGT	AACGCTCGA	1050
GGTGTGTTGGT	GGGGGTGACC	CCACCGGTGG	CCACCAGGGA	CGGCAAGTC	1100
CCCACAACGC	AGCTTCGACG	TCATATCGAT	CTGCTTGTGG	GGAGCGGCAC	1150
CCTCTGCTCG	GCCCCCTACG	TGGGGGACCT	GTGCGGGTCT	GTCTTTCTTG	1200
TTGGTCAACT	GTTTACCTTC	TCTCCAGGC	GCCACTGGAC	GACGCAAGAC	1250
TGCAATTGTT	CTATCTATCC	CGGCATATA	ACGGGTGATC	GATGGCATG	1300
GGATATGATG	ATGAAGTGGT	CCCCTACGGC	AGCGTTGGTG	GTAGCTCAGC	1350
TGCTCCGGAT	CCCACAAGCC	ATCATGGACA	TGATCGCTGG	TGCTCACTGG	1400
GGAGTCCCTGG	CGGGCATAGC	GTATTTCTCC	ATGGTGGGGA	ACTGGGGGAA	1450
GGTCCCTGGTA	GTGCTGCTGC	TATTTGCGCG	CGTCGACGCG	GAAACCCACG	1500
TCACCGGGGG	AAATGCGGGC	CGCACCAAGG	CTGGGCTTGT	TGGTCTCTCT	1550
ACACCAGGCG	CCAAGCAGAA	CATCCAAGTC	ATCAACACCA	ACGGCAGTTG	1600
GCACATCAAT	AGCACGGGCT	TGAATTGCAA	TGAAAGCCTT	AACACCGGCT	1650
GGTTAGCAGG	GCTCTTCTAT	CAACACAAAT	TCAACTCTTC	AGGCTGTCTT	1700
GAGAGGTTGG	CCAGCTGCCG	ACGCTTACCC	GATTTTGGCC	AGGGCTGGGG	1750
TCCTATCAGT	TATGCCAAGC	GAAGCGGCTT	CGACGAACGC	CCCTACTGCT	1800
GGCACTACCC	TCCAAGACCT	TGTGGCATGT	TGCCCCGAAA	GAGCGTGTGT	1850
GGCCCCGTAT	ATTGCTTCAC	TCCCAGCCCC	GTGGTGGTGG	GAACGACCGA	1900

FIG. 6A

SUBSTITUTE SHEET (RULE 26)

H77C

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
CAGGTCGGGC	GCGCCTACCT	ACAGCTGGGG	TGCAAATGAT	ACGGATGICT	1950
TCGTCTTAA	CAACACCAGG	CCACCGCTGG	GCAATTGGTT	CGGTGTGACC	2000
TGGATGAACT	CAACTGGATT	CACCAAAGTG	TGCGGAGOGC	COOCTTGTTG	2050
CATCGGAGGG	GTGGGCACAA	ACACCTTGCT	CTGCCCCACT	GATTGCTTCC	2100
GCAAACATCC	GGAAGCCACA	TACTCTUGGT	GCGGCTCOGG	TCCCTGGATT	2150
ACACCCAGGT	GCATGGTCCA	CTACCGGTAT	AGGCTTTGGC	ACTATCCTTG	2200
TACCATCAAT	TACACCATAT	TCAAAGTCAG	GATGTACGTG	GGAGGGGTGG	2250
AGCACAGGCT	GGAAGCGGOC	TGCAACTGGA	CGCGGGGGGA	ACGCTGTGAT	2300
CTGGAAGACA	GGGACAGGTC	CGAGCTCAGC	CGGTGCTGTC	TGTCCACAC	2350
ACAGTGGCAG	GTCTTTCGT	GTCTTTTCAC	GACCTTGCCA	GCTTGTGCA	2400
CCGGCCTCAT	CCACCTCCAC	CAGAACATTG	TGGACGTGCA	GTAATTGTAC	2450
GGGGTAGGGT	CAAGCATCGC	GTCTTGGGCC	ATTAAGTGGG	AGTACGTCTG	2500
TCTCCTGTTT	CTTCTGCTTG	CAGACGCGCG	CGTCTGCTCC	TGCTGTGGA	2550
TGATGTACT	CATATCCCAA	GCGGAGGCGG	CTTTGGAGAA	CCTCGTAATA	2600
CTCAATGCAG	CATCCCTGGC	CGGGACGCAC	GGTCTGTGTT	CCTTCTCTCT	2650
GTCTTTCTGC	TTTGCGTGGT	ATCTGAAGGG	TAGGTGGGTG	CCCGGAGCGG	2700
TCTACGCCCT	CTACGGGATG	TGGCCTCTCC	TCCTGCTCCT	GCTGGGGTTG	2750
CCTCAGCGGG	CATACGCACT	GGACACGGAG	GTGGCCGCGT	CGTGTGGCGG	2800
CGTTGTTCCT	GTGGGGTAA	TGGCGCTGAC	TCTGTGCGCA	TATTAACAAGC	2850
GCTATATCAG	CTGGTGCAATG	TGGTGGCTTC	AGTATTTTCT	GACCAGAGTA	2900
GAAGCGCAAC	TGCACGTGTG	GGTCCCCCC	CTCAACGTCC	GGGGGGGGCG	2950
CGATGCCGTC	ATCTTACTCA	TGTGTGTAGT	ACACCCGACC	CTGGTATTTG	3000
ACATCACCAA	ACTACTCCTG	GCCATCTTCC	GACCCCTTTG	GATTCTTCAA	3050
GCCAGTTTGC	TAAAGTCCC	CTACTTGTG	CGGTTCAAG	GCTTCTCCG	3100
GATCTGCGCG	CTAGCGCGGA	AGATAGCGCG	AGGTCATTAC	GTGCAAATGG	3150
CCATCATCAA	GTAGGGGCG	CTTACTGGCA	CCTATGTGTA	TAAOCATCTC	3200
ACCCCTCTTC	GAGACTGGGC	GCACAACGGC	CTGGGAGATC	TGGCGGTGGC	3250
TGTGGAACCA	GTGCTCTTCT	CCCGAATGGA	GACCAAGCTC	ATCAGTGGG	3300
GGGCAGATAC	CGCCGCGTGC	GGTGACATCA	TCAAAGGCTT	GCCGCTCTCT	3350
GCCCGTAGGG	GCCAGGAGAT	ACTGCTTGGG	CCAGCCGACG	GAATGGTCTC	3400
CAAGGGGTGG	AGGTTGCTGG	CGCCCATCAC	GGCGTAAGCC	CAGCAGACGA	3450
GAGGCCCTCT	AGGGTGTATA	ATCACCAGCC	TGACTGGCCG	GGACAAAAAC	3500
CAAGTGGAGG	GTGAGGTCCA	GATGTGTGTA	ACTGCTAACCC	AAACCTTCTT	3550
GGCAACGTGC	ATCAATGGGG	TATGCTGGAC	TGCTTACCAC	GGGGCGGGAA	3600
CGAGGACCAT	CGCATCACCC	AAGGGTCTCG	TCATCCAGAT	GTATACCAAT	3650
GTGGACCAAG	ACCTTGTTGG	CTGGCCCGCT	CCTCAAGGTT	CCCGCTCATT	3700
GACACCTGT	ACCTGCGGCT	CCTCGGACCT	TTACCTGGTC	ACGAGGCACG	3750
CCGATGTCAT	TCCCGTGGC	CGCGAGGGTG	ATAGCAGGGG	TAGCTGCTT	3800

FIG. 6B

SUBSTITUTE SHEET (RULE 26)

H77C

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
TGCCCCCGGC	CCATTTCCTA	CTTGAAAGGC	TCCTCGGGGG	GTCCGCTGTT	3850
GTGCCCCGCG	GGACACGCGG	TGGGOCATTT	CAGGGGCGCG	GIGTGCAACC	3900
GTGGAGTGGC	TAAAGCGGTG	GACTTTATCC	CTGTGGAGAA	CCTAGGGACA	3950
ACCATGAGAT	CCCCGGTGT	CACGGACAAC	TCCTCTCCAC	CAGCAGTGCC	4000
CCAGAGCTTC	CAGGTGGGCC	ACCTGCATGC	TCCCACCGGC	AGCGGTAAAG	4050
GCACCAAGGT	CCCGGCTGGG	TACGCAGGCC	AGGGCTACAA	GGTGTGGGTG	4100
CTCAACCCCT	CTGTTCCTGC	AACGCTGGGC	TTTGGTGCCT	ACATGTCCAA	4150
GGCCCATGGG	GTGATCCCTA	ATATCAGGAC	CGGGGTGAGA	ACAATTACCA	4200
CTGGCAGGCC	CATCAOGTAC	TCCACCTAAG	GCAAGTTCTT	TGCGGACGGC	4250
GGGTGCTCAG	GAGGTGCTTA	TGACATAATA	ATTTGTGACG	AGTGCCACTC	4300
CACGGATGCC	ACATCCATCT	TGGGCATCGG	CACTGTCTTT	GACCAAGCAG	4350
AGACTGCGGG	GGCGAGACTG	GTGTGTCTCG	CCACTGCTAC	CCCTCCGGGC	4400
TCCGTCACTG	TGTCCCATCC	TAACATCGAG	GAGGTGTCTC	TGTCCACCAC	4450
CGGAGAGATC	CCCTTTTACG	GCAAGGCTAT	CCCCCTCGAG	GTGATCAAGG	4500
GGGGAAGACA	TCTCATCTTC	TGCCACTCAA	AGAAGAAGTG	CGACGAGCTC	4550
GCCGCGAAGC	TGGTCCCATT	GGGCATCAAT	GCCGTGGCCT	ACTACCGCGG	4600
TCTTGACGTG	TCTGTCTATC	CGACCAGCGG	CGATGTGTGC	GTGCTGTCCA	4650
CCGATGCTCT	CATGACTGGC	TTTACCGGGG	ACTTCGACTC	TGTGATAGAC	4700
TGCAACACGT	GTGTCACTCA	GACAGTCGAT	TTCAGCCTTG	ACCTTACCTT	4750
TACCATTGAG	ACAACCACGC	TCCCCCAGGA	TGCTGTCTCC	AGGACTCAAC	4800
GCCGGGGCAG	GACTGGCAGG	GGGAAGCCAG	GCATCTATAG	ATTTGTGGCA	4850
CCGGGGGAGC	GCCCCCTCCG	CATGTTCGAC	TGTCGGTCC	TCTGTGAGTG	4900
CTATGACGGG	GGCTGTGCTT	GGTATGAGCT	CAOGCCGGC	GAGACTACAG	4950
TTAGGCTACG	AGCGTACATG	AACACCCCGG	GGCTTCCCGT	GTGCCAGGAC	5000
CATCTTGAAT	TTTGGGAGGG	CGTCTTTAAG	GGCTCACTC	ATATAGATGC	5050
CCACTTTTAA	TCCCAGACAA	AGCAGAGTGG	GGAGAACTTT	OCTTACCTGG	5100
TAGCGTACCA	AGCCACCGTG	TGCGCTAGGG	CTCAAGCCCC	TCCCCCATCG	5150
TGGGACCAGA	TGTGGAAGTG	TTTGATCCGC	CTTAAACCCA	CCCTCCATGG	5200
GCCAACACCC	CTGCTATACA	GACTGGGGGC	TGTTCAAGAAT	GAAGTCACCC	5250
TGACGCACCC	AATCACCAAA	TACATCATGA	CATGCATGTC	GGCGGACCTG	5300
GAGGTGCTCA	CGAGCACCTG	GGTGCTCGTT	GGGGGGGTCC	TGGCTGCTCT	5350
GGCGGGGTAT	TGCTGTCAA	CAGGCTGGGT	GGTCATAGTG	GGCAGGATCG	5400
TCTTGTCCGG	GAAGCCGGCA	ATTATACCTG	ACAGGGAGGT	TCTCTACCAG	5450
GAGTTCGATG	AGATGGAAGA	GTGCTCTCAG	CACTTACCGT	ACATCGAGCA	5500
AGGGATGATG	CTCGCTGAGC	AGTTCAGCA	GAAGGCGCTC	GGCTCCCTGC	5550
AGACCGCGTC	CCGCCATGCA	GAGGTATCA	CCCTGCTGT	CCAGACCAAC	5600
TGGCAGAAAC	TGGAGGTCTT	TTGGGGCAAG	CACATGTGGA	ATTTTCATCAG	5650
TGGGATACAA	TACTTGGCGG	GCCTGTCAAC	GCTGCCCTGGT	AACCCCGCCA	5700

FIG. 6C

SUBSTITUTE SHEET (RULE 26)

H77C

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1234567890	1234567890	1234567890	1234567890	1234567890	
TTGCTTCATT	GATGGCTTTT	ACAGCTGCGG	TCACCAGCCC	ACTAACCCT	5750
GGCCAAACCC	TCCTCTTCAA	CATATTGGGG	GGGTGGGTGG	CTGCCCAGCT	5800
CGCCGCCCCC	GGTGGCGCTA	CTGCCCTTGT	GGGTGCTGGC	CTAGCTGGCG	5850
CGCCCATCGG	CAGCGTTGGA	CTGGGGAAGG	TCCTGGTGGG	CATTCTTGCA	5900
GGGTATGGCG	CGGGGCTGGC	GGGAGCTCTT	GTAGCATTCA	AGATCATGAG	5950
CGGTGAGGTC	CCCTCCACGG	AGGACCTGGT	CAATCTGCTG	CCCGCATCC	6000
TCTCGGCTGG	AGCCCTTGTA	GTCGGTGTGG	TCTGGGCAGC	AATACTGGCG	6050
CGGCACGTTG	GCCCCGGCGA	GGGGGCAGTG	CAATGGATGA	ACCGGCTAAT	6100
AGCCTTCGCC	TCCCCGGGGA	ACCATGTTTC	CCCCAAGCAC	TACGTGCGCG	6150
AGAGCGATGC	AGCGGCCCCG	GTCAC TGCA	TACTCAGCAG	CCTCACTGTA	6200
ACCCAGCTCC	TGAGGGGACT	GCATCAGTGG	ATAAGCTGGG	AGTGTACAC	6250
TCCATGCTCC	GGTTCCTGGC	TAAAGGACAT	CTGGGACTGG	ATATGCGAGG	6300
TGCTGAGCGA	CTTFAAGACC	TGGCTGAAAG	CCAAGCTCAT	GCCACAAC TG	6350
CCTGGGATTC	CCTTTGTGTC	CTGCCAGCGC	GGGTATAGGG	GGGTCTGGCG	6400
AGGAGACGGC	ATTATGCACA	CTCGCTGCCA	CTGTGGAGCT	GAGATCAC TG	6900
GACATGTCAA	AAACGGGACG	ATGAGGATCG	TGGTTCCTAG	GACCTGCAGG	6950
AACATGTGGA	GTGGGACGTT	CCCCATTAA	GCCTACACCA	CGGGCCCC TG	6550
TACTCCCCCT	CCTGCGCGCA	ACTATAAGTT	CGCGCTGTGG	AGGGTGTC TG	6600
CAGAGGAATA	CGTGGAGATA	AGCGGGGTGG	GGGACTTCCA	CTACGTATCG	6650
GGTATGACTA	CTGACAATCT	TAAATGCCCG	TGCCAGATCC	CATGCCCCGA	6700
ATTTTTCACA	GAATTGGACG	GGGTGCGGCT	ACACAGGTTT	GCGCCCCCTT	6750
GCAAGCCCCCT	GCTGCGGGAG	GAGGTATCAT	TCAGAGTAGG	ACTCCACGAG	6800
TACCCGGTGG	GGTGGCAATT	ACCTTGGGAG	CCCGAACCGG	ACGTAGCCGT	6850
GTGACGCTCC	ATGCTCACTG	ATCCCTCCCA	TATAACAGCA	GAGGCGGCGG	6900
GGAGAAGGTT	GGCGAGAGGG	TCACCCCTTT	CTATGGCCAG	CTCCTGGGCT	6950
AGCCAGCTGT	CCGCTOCATC	TCTCAAGGCA	ACTTGCACCG	CCAACCATGA	7000
CTCCCTGAC	GCCGAGCTCA	TAGAGGCTAA	CCTCCTGTGG	AGGCAGGAGA	7050
TGGGGGGCAA	CATCACCAGG	GTGAGTTCAG	AGAACAAGT	GGTGATTCTG	7100
GACTCCTTCG	ATCCGCTTGT	GGCAGAGGAG	GATGAGGGGG	AGGTCTCCGT	7150
ACCTGCAGAA	ATTCTGGCGA	AGTCTGGGAG	ATTGGCCCGG	GCCC TGCCCG	7200
TCTGGGCGCG	GCCGGACTAC	AAACCCCGCG	TAGTAGAGAC	GTGGAAAAG	7250
CCTGACTACG	AACCACTGT	GGTCCATGGC	TGCCCCCTAC	CACCTOCACG	7300
GTCCCCCTCT	GTGCCCTCCG	CTCGGAAAAA	GCGTACGGTG	GTCCTCACCG	7350
AATCAACCTT	ATCTACTGCC	TTGGCCGAGC	TTGCCACCAA	AAGTTT TGCC	7400
AGCTCCTCAA	CTTCGGGCAT	TACGGGCGAC	AATAAGACAA	CATCCTCTGA	7450
GCCCCCCCCCT	TCTGGCTGCC	CCCCCGACTC	CGACGTTGAG	TCTATTCTTT	7500
CCATGCCCCC	CCTGGAGGGG	GAGCCTGGGG	ATCCGGATCT	CAGCGACGGG	7550
TCATGGTCGA	CGGTACGTAG	TGGGGCCGAC	ACCGAAGATG	TCGTGTGCTG	7600

FIG. 6D

SUBSTITUTE SHEET (RULE 26)

H77C

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1234567890	1234567890	1234567890	1234567890	1234567890	
CTCAATGICT	TATTCCTGGA	CAGGCGCACT	CGTCACCCCG	TGCGCTGCGG	7650
AAGAACA AAA	ACTGCCCATC	AACGCCTGA	GCAACTCGTT	GCTACGCCAT	7700
CACAATCTGG	TGTATTCCAC	CACCTCAAGC	AGTGCTTGCC	AAAGGCAGAA	7750
GAAAGTCACA	TTTGACAGAC	TGCAAGTICT	GGACAGCCAT	TACCAGGACG	7800
TGCTCAAGGA	GGTCAAGCA	GCGGCGTCAA	AAGTGAAGGC	TAACTTGCTA	7850
TCOGTAGAGG	AAGCTTG CAG	CCTGACGCCC	CCACATT CAG	CCAAATCCAA	7900
GTTTGCGCTAT	GCGGCAAAAG	ACGTCCGTTG	CCATCC CAG	AAGGCGGTAG	7950
CCCACATCAA	CTCCGTGTGG	AAAGACCTTC	TGGAAGACAG	TGTAACACCA	8000
ATAGACACTA	CCATCATGGC	CAAGAACGAG	GTTTTCTGCG	TTCAGCCTGA	8050
GAAGGGGGGT	CGTAAGCCAG	CTCGTCTCAT	CGTGTTCCCC	GACCTGGGCG	8100
TGCGCGTGIG	CGAGAAGATG	GCCCTGTACG	ACGTGGT TAC	CAAGCTCCCC	8150
CTGGCCGIGA	TGGGAAGCTC	CTACGGATT C	CAATACTCAC	CAGGACAGCG	8200
GGTTGAATTC	CTCGTGCAAG	CGTGGAAATC	CAAGAAGACC	CCGATGGGGT	8250
TCTCGTATGA	TACCCGCTGT	TTTGACTCCA	CAGTCACTGA	GAGCGACATC	8300
CGTACGGAGG	AGGCAATTTA	CCAATGTTGT	GACCTGGACC	CCCAAGCCCC	8350
CGTGGCCATC	AAGTCCCTCA	CTGAGAGGCT	TATATGTTGGG	GGCCCTCTTA	8400
CCAATTCAAG	GCGGGA AAAC	TGCGGCTACC	GCAGGTGCCG	CGCGAGCGGC	8450
GTA CTGACAA	CTAGCTGTGG	TAACACCTTC	ACTTGCTACA	TCAAGGCCCC	8500
GGCAGCCTGT	CGAGCCGCAG	GGCTCCAGGA	CTGCACCATG	CTCGTGTGTG	8550
CGGACGACTT	AGTCGTTATC	TGTGAAAGTG	CGGGGGTCCA	GGAGGACCGG	8600
CGGAGCCTGA	GAGCCTTCAC	GGAGGCTATG	ACCAGGTA CT	CCGCCCCCCC	8650
CGGGGACCCC	CCACAACCAG	AATACGACTT	GGAGCTTATA	ACATCATGCT	8700
CCTCCAACGT	GTCAGTCGCC	CACGACGGCG	CTGGAAAGAG	GGTCTACTAC	8750
CTTACCCGTG	ACCTTACAAC	CCCCCTCGCG	AGAGCCGGGT	GGGAGACAGC	8800
AAGACACACT	CCAGTCAATT	CCTGGCTAGG	CAACATAATC	ATGTTTGCCC	8850
CCACACTGTG	GGCGAGGATG	ATACTGATGA	CCATTTCTTT	TAGOGTCTC	8900
ATAGCCAGGG	ATCAGCTTGA	ACAGGCTCTT	AAC TGTGAGA	TCTACGGAGC	8950
CTGCTACTCC	ATAGAACCAC	TGGATCTACC	TCCAATCATT	CAAAGACTCC	9000
ATGGCCTCAG	CGCATTTTCA	CTCCACAGTT	ACTCTCCAGG	TGAAATCAAT	9050
AGGGTGGCCG	CATGCCCTCAG	AAA ACTTGGG	GTCCCGCCCT	TGCGAGCTTG	9100
GAGACACCGG	GCCCCGAGCG	TCCGCGCTAG	GCTTCTGTCC	AGAGGAGGCA	9150
GGGCTGCCAT	ATGTGGCAAG	TACCTCTTCA	ACTGGGCAGT	AAGAACA AAG	9200
CTCAA ACTCA	CTCCAATAGC	GCGCGCTGGC	CGGCTGGACT	TGTCCGGTTG	9250
GTTACAGGCT	GGCTACAGCG	GGGAGACAT	TTATCACAGC	GTGTCTCATG	9300
CCCGGCCCCG	CTGGTTCTGG	TTTTTGCC TAC	TCCTGCTCGC	TGCAGGGGTA	9350
GGCATCTACC	TCCTCCCCAA	CCGATGAAGG	TTGGGGTAAA	CAC TCCGGCC	9400
TCTTAAGCCA	TTTCCTGTTT	TTTTTTTTTT	TTTTTTTTTT	TTTTTCTTTT	9450
TTTTTTTCTT	TCCTTTCTCT	CTTTTTTTTC	TTTCTTTTTC	CCTTCTTTAA	9500

FIG. 6E

SUBSTITUTE SHEET (RULE 26)

H77C

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
TGGTGGCTCC	ATCTTAGCCC	TAGTCACGGC	TAGCTGTGAA	AGGTCCGIGA	9550
GCCGCATGAC	TGCAGAGAGT	GCTGATACTG	GCCTCTCTGC	AGATCATGT	9599

FIG. 6F

H77C

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
MSTNPKPQRK	TKRNINRRPQ	DVKFPGGQI	VGGVYLLPRR	GPRLGVRATR	50
KTSESRQPRG	RRQPIPKARR	PEGRIWAQPG	YFWPLYGNEG	CGWAGWLLSP	100
RGSRPSWGPT	DPRRRSRNLG	KVIDTLTQGF	ADLMGYIPLV	GAPLGGAARA	150
LAHGVRVLED	GVNYATGNLP	GCSFSIFLLA	LLSCLTVPAS	AYQVRNSSGL	200
YHVINDCPNS	SIVYEADAI	LHTPGCVPCV	REGNASRCW	AVTPTVATRD	250
GKLPITQLRR	HIDLLVGSAT	LCSALYVGD	CGSVFLVQQL	FTFSPRRHWT	300
TQDQNCSTYP	GHTTGHMAW	IMMNSPTA	ALVVAQLLRI	PQAIMDMIAG	350
AHWGVLGIA	YFSMVGWAK	VLVVLILFAG	VDAETHVIGG	NAGRTTAGLV	400
GLITPGAKQN	IQLININGSW	HINSTALNCN	ESLNTGWLAG	LFYQHKFNSS	450
GCPERLASCR	RLTDFAQGWG	PISYANGSGL	DERPYCWHYP	PRPOGIVPAK	500
SVCGPVYCFT	PSPVVVGTTD	RSGAPTYSWG	ANDIDVFVLN	NIRPPLGNWF	550
GCTWMNSTGF	TKVCGAPPCV	IGGVGNNTLL	CFIDCFRKHP	EATYSRCGSG	600
PWITPRCMTD	YPYRLWHYPC	TINYTIFKVR	MYVGGVEHRL	EAACNWTIRGE	650
RCDLEDNRDS	ELSPILLSTT	QWQVLPSCFT	TLPALSTGLI	HLHQNTVDVQ	700
YLYGVGSSIA	SWAIKWEYV	LLFLLIADAR	VCSCILWMLL	ISQAEAALEN	750
LVILNAASLA	GTHGLVSFLV	FFCFAWYKLG	RWPGAVYZAL	YGMWPLLLLL	800
LALPQRAYAL	DTEVAASCGG	VVLVGLMALT	LSPYYKRYIS	WCMWWLQYFL	850
TRVEAQLHW	VPPLNVRGGR	DAVILLMCVV	HPITLVFDITK	LLLAIFGPLW	900
ILQASLLKVP	YFVRVQGLLR	ICALARKIAG	GHYVQMAITK	LGALTGTIVY	950
NHILTPLRDA	HNGLRDLAVA	VEPVVFSRME	TKLITWGADT	AACGDIINGL	1000
PVSARRQOEI	LLGPADGMVS	KGWRLAPIT	AYAQQTRGLL	GCTITSLTGR	1050
DKNQVEGEVQ	IVSTATQTFL	ATCINGVCWT	VYHAGITRTI	ASPKGPVIQM	1100
YTINVDQDLVG	WPAPQGSRL	TPCTCGSSDL	YLVIRHADVI	PVRRRGDSRG	1150
SLLSPRPISY	LKGSSGGPLL	CPAGHAVGLF	RAAVCTRGA	KAVDFIPVEN	1200
LGTIMRSPVF	TDNSSPPAVP	QSFOVAHLHA	PTGSGKSTKV	PAAYAAQGYK	1250
VLVLNPSVAA	TLGFGAYMSK	AHGVDPNIRT	GVRTTTTGSP	ITYSTYGKFL	1300
ADGGCSCGAY	DIICDECHS	TDATSIILGIG	TVLDQAEIAG	ARLVVLATAT	1350
PPGSVTVSHP	NIEEVALSIT	GEIPFYGKAI	PLEVIKGRH	LIFCHSKKKC	1400
DELAACLVAL	GINAVAYYRG	LDVSVIPTSG	DVVVVSIDAL	MIGFTGDFDS	1450
VIDQNTCVIQ	TVDFSLDPTF	TIETTTLPQD	AVSRTQRRGR	TGRGKPGIYR	1500
FVAPGERPSG	MFDSSVLCEC	YDAGCAWYEL	TPAETTVRLR	AYMNTPLGFV	1550
CQDHLEFWEG	VFTGLTHIDA	HFLSQIKQSG	ENFPYLVAYQ	ATVCARAQAP	1600
PPSWDQMKC	LIRLKPTLHG	PTFLLYRLGA	VQNEVTLIHP	ITKYIMTCMS	1650
ADLEVVTSTW	VLVGGVLAAL	AAYCLSTGCV	VTVGRIVLSG	KPALIPDREV	1700
LYQEFDEMEE	CSQHLPTYBQ	GMMLAEQFKQ	KALGLLQIAS	RHAEVITPAV	1750
QTNWQKLEVF	WAKHMANFTS	GIQYLAGLST	LPGNPAIASL	MAFTAAVTSP	1800
LTITQITLLFN	ILGGWAAQL	AAPGAATAFV	GAGLAGAAIG	SVGLGKVLVD	1850
ILAGYGAGVA	GALVAFKIMS	GEVPSTEDLV	NLLPAILSPG	ALVVGWVCAA	1900

FIG. 6G

SUBSTITUTE SHEET (RULE 26)

H77C

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
ILRRHVGPGE	GAVQWMNRLI	AFASRGNHVS	PIHYVPESDA	AARVTAILSS	1950
LITVTQLLRRL	HQWISSECTT	PCSGSWLRDI	WJWICEVLSD	FKTWLKAKLM	2000
PQLPGIPFVS	CQRGYRGWWR	GDGIMHIRCH	CGAETTGHVK	NGIMRIVGPR	2050
TCRNWMSGTF	PINAYTTGFC	TPLPAPNYKF	ALWRVSAEEY	VEIRRVGDFH	2100
YVSGMITDNL	KCPCQIPSPE	FFTELDGVRL	HRFAPPCKPL	LREEVSFRVG	2150
LHEYFVGSQJ	PCEPEPDVAV	LTSMLTDP SH	ITAEAGRRRL	ARGSPPSMAS	2200
SSASQLSAPS	LKATCTANHD	SPDAELLEAN	LLWRQEMGGN	ITRVESENKV	2250
VILDSFDELV	AEEDEREVS	PAETLRKSRR	FARALPWAR	PDYNPPLVET	2300
WKKPDYEPV	VHGCPPLPPR	SPPVPPPRKK	RIVVLTESTL	STALAEATK	2350
SFGSSSTSGI	TGENTTTSS	PAPSGCPPDS	DVESYSSMPP	LEGEFGDPL	2400
SDGSWSTVSS	GADTEDVCC	SMSYSWIGAL	VTFCAAEEDK	LPINALSNSL	2450
LRHHNLVYST	TSRSACQROK	KVTFDLRLQL	DSHYQDLVKE	VKAAASKVKA	2500
NLLSVEEACS	LTPPHSAKSK	FGYGAKDVRC	HARKAVAHIN	SVWKDLLEDS	2550
VTPIDTTIMA	KNEVFCVQPE	KGGRKPARLI	VFPDLGVRVC	EKMALYDVVS	2600
KLPLAVMGSS	YGFOYSPGQR	VEFLVQAWKS	KKTFMGFSYD	TRCFDSTVTE	2650
SDIRTEEATY	QCCDLDPQAR	VAIKSLTERL	YVGGPLINSR	GENCGYRRCR	2700
ASGVLTTSCG	NILTCYIKAR	AACRAAGLOD	CIMLVCGDDL	VVICESAGVQ	2750
EDAASLRAFT	EAMTRYSAAP	GDPPOPEYDL	ELITSCSSNV	SVAHDGAGKR	2800
VYYLTRDPTT	PLARAAWETA	RHTFVNSWL	NIIMFAPTLW	ARMILMTHFF	2850
SVLIARDQLE	QALNCETYGA	CYSIEPLDLP	PIIQLRHGLS	AFSLHSYSFG	2900
EINRVAACLR	KLGVPPPLRAW	RHRARSVRAR	LLSRGGRAAI	CGKYLENWAV	2950
RTKLKLTPIA	AAGRLDLSGW	FTAGYSGGDI	YHSVSHARPR	WFWFCLLLLA	3000
AGVGITYLLN	R				3011

FIG. 6H

HC-J4

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
GCCAGCCCC	TGATGGGGG	GACACTCCAC	CATGAATCAC	TCCCTGTGGA	50
GGAACACTG	TCTTCACGA	GAAAGOGTCT	AGCCATGGCG	TTAGTATGAG	100
TGTCTGTGAG	CCTCCAGGAC	CCCCCTCC	GGGAGAGCCA	TAGTGGTCTG	150
CGGAACCGGT	GAGTACACCG	GAATTGCCAG	GACGAACGGG	TCTTTCTTG	200
GATCAACCCG	CTCAATGCCCT	GGAGATTTCG	GCGTGCCCC	GCGAGACTGC	250
TAGCCGAGTA	GTGTTGGGTC	GCGAAAGGCC	TTGTGGTACT	GCTTGATAGG	300
GTGCTTGCGA	GTGCCCCGGG	AGGTCTCGTA	GACCGTGCAC	CATGAGCACG	350
AATCCTAAAC	CTCAAAGAAA	AACCAAACGT	AACACCAACC	GCCGCCACCA	400
GGACGTCAAG	TTCCTGGGG	GTGGTCAGAT	CGTTGGTGGG	GTTTACCTGT	450
TGCCGCGCAG	GGGCCCCAGG	TTGGGTGTGC	GCGGACTAG	GAAGGCTTCC	500
GAGCGGTGCG	AACCTCGTGG	AAGGCGACAA	CCTATCCCAA	AGGCTCGCCG	550
ACCCGAGGGC	AGGGCCTGGG	CTCAGCCCGG	GTACCCCTTG	CCCTCTATG	600
GCAATGAGGG	CCTGGGGTGG	GCAGGATGGC	TCTGTTCACC	CCGGGGCTCC	650
CGGCTAGT	GGGGCCCCAC	GGACCCCGG	CGTAGGTGCG	GTAACCTGGG	700
TAAGGTGATC	GATACCTTA	CATGCGGCTT	CGCCGATCTC	ATGGGGTACA	750
TTCCGCTCGT	CGGCGCCCC	CTAGGGGGCG	CTGCCAGGGC	CTTGGCACAC	800
GGTGTCCGGG	TTCTGGAGGA	CGGCGTGAAC	TATGCAACAG	GGAACCTGCC	850
CGGTGTCTCT	TTCTCTATCT	TCTCTTGGC	TCTGTCTGCC	TGTTTGACCA	900
TCCAGCTTC	CGCTTATGAA	GTGCGCAACG	TGTCCGGGAT	ATACCATGTC	950
ACGAACGACT	GCTCCAACTC	AAGCATTTGTG	TATGAGGCAG	CGGACGTGAT	1000
CATGCATACT	CCCGGGTGCG	TGCCCTGTGT	TCAGGAGGGT	AACAGCTCCC	1050
GTGTCTGGGT	AGCGCTCACT	CCACGCTCG	CGGCCAGGAA	TGCCAGCGTC	1100
CCCACTACGA	CAATACGACG	CCACGTGAC	TGTCTCGTTG	GGACGGCTGC	1150
TTTCTGCTCC	GCTATGTACG	TGGGGGATCT	CTGCCGATCT	ATTTTCTCTG	1200
TCTCCAGCT	GTCACCTTC	TGGCTCGCC	GGCATGAGAC	AGTGCAGGAC	1250
TGCAACTGCT	CAATCTATCC	CGGCCATGTA	TCAGGTCCACC	GCAATGGCTTG	1300
GGATATGATG	ATGAACTGGT	CACCTACAAC	AGCCCTAGTG	GTGTGCGAGT	1350
TGCTCCGGAT	CCCACAAGCT	GTCGTGGACA	TGGTGGCGGG	GGCCCACTGG	1400
GGAGTCCCTGG	CGGGCCTTGC	CTACTATTCC	ATGGTAGGGA	ACTGGGCTAA	1450
GGTTCTGATT	GTGGCGCTAC	TCTTTGCCCG	CGTTGACGGG	GAGAACCA	1500
CGACGGGGAG	GGTGGCCGGC	CACACCACCT	CCGGGTTCAC	GTCCTTTTC	1550
TCATCTGGGG	CGTCTCAGAA	AATCCAGCTT	GTGAATACCA	ACGGCAGCTG	1600
GCACATCAAC	AGGACTGCCC	TAAATTGCAA	TGACTCCCTC	CAAACTGGGT	1650
TCTTTGCCGC	GCTGTTTTAC	GCACACAAGT	TCAACTCGTC	CGGGTGCCCC	1700
GAGCGCATGG	CCAGCTGCCG	CCCATTTGAC	TGGTTCCGCC	AGGGGTGGGG	1750
CCCATCACC	TATACTAAGC	CTAACAGCTC	GGATCAGAGG	CCTTATTGCT	1800
GGCATTACGC	GCTTCGACCG	TGTGGTGTGC	TACCCGCGTC	GCAGGTGTGT	1850
GGTCCAGTGT	ATTGTTTCAC	CCCAAGCCCT	GTTGTGGTGG	CGACCAACGA	1900

FIG. 7A

SUBSTITUTE SHEET (RULE 26)

HC-J4

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
TCGTTCCGGT	GTCCCTACGT	ATAGCTGGGG	GGAGAATGAG	ACAGACGTGA	1950
TGCTCCTCAA	CAACACGGGT	CCGCCACAAG	GCAACTGGTT	CGGCTGTACA	2000
TGGATGAATA	GTACTGGGTT	CAC TAAGACG	TGCGGAGGTC	CCCCGTGTAA	2050
CATCGGGGGG	GTCGGTAACC	GCACCTTGAT	CTGCCCCACG	GACTGCTTCC	2100
GGAAGCACCC	CGAGGCTACT	TACACAAAAT	GTCGGCTGGG	GOCCGTGGTTG	2150
ACACCTAGGT	GCCTAGTAGA	CTAACCATAAC	AGGCTTTGGC	ACTACCCCTG	2200
CACCTCTCAAT	TTTTCCATCT	TTAAGGTTAG	GATGTATGTG	GGGGGGGTGG	2250
AGCACAGGCT	CAATGCCGCA	TGCAATTGGA	CTCGAGGAGA	GCGCTGTAAAC	2300
TTGGAGGACA	GGGATAGGTC	AGAACTCAGC	CCGCTGCTGC	TGTCCTACAAC	2350
AGAGTGGCAG	ATACTGCOCT	GTGCTTTTAC	CACCTTACCG	GCTTTTATCCA	2400
CTGGTTTIGAT	CCATCTCCAT	CAGAACATCG	TGGAAGTGCA	ATACTGTATC	2450
GGTGTAGGGT	CAGCGTTTGT	CTCCTTTGCA	ATCAAATGGG	AGTACATCCT	2500
GTGTCTTTTC	CTTCTCCTGG	CAGACGGCGG	CGTGTGTGCC	TGCTTGTGGA	2550
TGATGCTGCT	GATAGCCGAG	GCTGAGGCGG	CCTTAGAGAA	CTTGGTGGTC	2600
CTCAATGCCG	CGTCCGTGGC	CGGAGCGCAT	GGTATTCTCT	CCTTTCTTGT	2650
GTCTTTCTGC	GCCGCTTGGT	ACATTAAAGG	CAGGCTGGCT	CCTGGGGGGG	2700
CGTATGCTTT	TTATGGCGTA	TGGCCGCTGC	TCCGTCTCCT	ACTGGCGTTA	2750
CCACCACGAG	CTTACGCCTT	GGACCGGGAG	ATGGCTGCAT	CGTGGGGGGG	2800
TGCGGTTCTT	GTAGGTCTGG	TATTCTTGAC	CTTGTCAACA	TACTACAAAG	2850
TGTTTCTCAC	TAGGCTCATA	TGGTGGTTAC	AATACTTTAT	CACCAGAGCC	2900
GAGGCGCACA	TGCAAGTGTG	GGTCCCCCCC	CTCAACGTTT	GGGGAGGCGG	2950
CGATGCCATC	ATCCTCCTCA	CGTGTGCGGT	TCATCCAGAG	TTAATTTTTTG	3000
ACATCACCAA	ACTCCTGCTC	GCCATACTCG	GCCCGCTCAT	GGTGTCTCAG	3050
GCTGGCATAA	CGAGAGTGCC	GTACTTCGTG	CGCGCTCAAG	GGCTCATTCG	3100
TGCATGCATG	TTAGTGGGAA	AAGTCGCCGG	GGGTCATTAT	GTCCAAATGG	3150
TCCTTCATGAA	GCTGGGCGCG	CTGACAGGTA	CGTACGTTTA	TAACCATCTT	3200
ACCCCACTGC	GGGACTGGGC	CCACGCGGGC	CTACGAGACC	TTGCGGTGGC	3250
GGTAGAGCCC	GTGCTCTTCT	CCGCCATGGA	GACCAAGGTC	ATCACCTGGG	3300
GAGCAGACAC	CGCTGCGTGT	GGGGACATCA	TCTTGGGICT	ACCGTCTTCC	3350
GCCCGAAGGG	GGAAGGAGAT	ATTTTGTGGG	CCGGCTGATA	GTCTOGAAGG	3400
GCAAGGGTGG	CGACTCCTTG	CGCCCATCAC	GGCTACTTCC	CAACAAAGGC	3450
GGGGCGTACT	TGGTTGCATC	ATCACTAGCC	TCACAGGCGG	GGACAAGAAC	3500
CAGGTGGAAG	GGGAGGTTCA	AGTGGTTTCT	ACCGCAACAC	AATCTTTTCT	3550
GGCGACCTGC	ATCAACGGGG	TGTGCTGGAC	TGCTTACCAT	GGCGCTGGCT	3600
CGAAGACCCCT	AGCCGGTCCA	AAAGGTCCAA	TCACCCAAAT	GTACACCAAT	3650
GTAGACCTGG	ACCTCGTCCG	CTGGCAGGCG	CCCCCGGGGG	CGCGCTCCAT	3700
GACACCATGC	AGCTGTGGCA	GCTCGGACCT	TTACTTGGTC	ACGAGACATG	3750
CTGATGTICAT	TCCGGTGGCG	CGGCGAGGCG	ACAGCAGGGG	AAGTCTACTC	3800

FIG. 7B

SUBSTITUTE SHEET (RULE 26)

HC-J4

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
TCCCCAGGC	CCGTCCTCTA	CCTGAAAGGC	TCCTGGGGTG	GTCCATTGCT	3850
TTGCCCCG	GGGACGTCG	TGGGCGTCCT	CCGGGCTGCT	GTGTGCAACC	3900
GGGGGGTGG	GAAGGCGGTG	GACTTCATAC	CCGTTGAGTC	TATGGAAACT	3950
ACCATGCGGT	CTCCGGTCCT	CACAGACAAC	TCAACCCCCC	CGGCTGTACC	4000
GCAGACATTC	CAAGTGGCAC	ATCTGCACGC	TCTTACTGGC	AGCGGCAAGA	4050
GCACCAAGT	GCCGGCTGGG	TATGCAGCCG	AAGGGTACAA	GGTGCTGGTC	4100
CTGAACCCGT	CCGTTGCCGC	CACCTTAGGG	TTTGGGGCGT	ATATGTCCAA	4150
GGCACACGGT	ATCGACCCTA	ACATCAGAAC	TGGGGTAAGG	ACCATTAACA	4200
CGGGCGGCTC	CATTACGTAC	TCCACCTATG	GCAAGTTGCT	TGCCGACGGT	4250
GGCTGTCTTG	GGGGGGCCTA	TGACATCATA	ATATGTGATG	AGTGCCACTC	4300
AACTGACTCG	ACTACCATCT	TGGGCATCGG	CACAGTCTTG	GAOCAAGCGG	4350
AGACGGCTGG	AGCGCGGCTC	GTCGTGCTCG	CCACCGCTAC	ACCTCCGGGA	4400
TCGGTTACCG	TGCCACACCC	CAATATCGAG	GAAATAGGCG	TGTCCAACAA	4450
TGGAGAGATC	CCCTTCTATG	GCAAAGOCAT	CCCATTTGAG	GCCATCAAGG	4500
GGGGGAGGCA	TCTCATTTTC	TGCCATTCCA	AGAAGAAATG	TGACGAGCTC	4550
GCCGCAAAGC	TGACAGGCGT	CGGACTGAAC	GCTGTAGCAT	ATTACCGGGG	4600
CCTTGATGTG	TCCGTCATAC	CGCCTATCGG	AGACGTGGTT	GTCGTGGCAA	4650
CAGACGCTCT	AATGACGGGT	TTCACCGGCG	ATTTTGACTC	AGTGATCGAC	4700
TGCAATACAT	GTGTACCCCA	GACAGTCGAC	TTCAGCTTGG	ATCCACCTTT	4750
CACCATTGAG	ACGACGACCG	TGCCCCAAGA	CGCGGTGTGG	CGCTCGCAAC	4800
GGCGAGGTAG	AACTGGCAGG	GGTAGGAGTG	GCATCTACAG	GTTTGTGACT	4850
CCAGGAGAAC	GGCCCTCGGG	CATGTTGCGT	TCTTCGGTCC	TGTGTGAGTG	4900
CTATGACGCG	GGCTGTGCTT	GGTATGAGCT	CACGCCCGCT	GAGACCTCGG	4950
TTAGGTTGCG	GGCTTACCTA	AATACACCAG	GGTTGCCCGT	CTGCCAGGAC	5000
CATCTGGAGT	TCTGGGAGAG	CGTCTTCACA	GGCCTCACCC	ACATAGATGC	5050
CCACTTCCTG	TCCCAGACTA	AACAGGCAGG	AGACAACTTT	CCTTACCTGG	5100
TGGCATATCA	AGCTACAGTG	TGCGCCAGGG	CTCAAGCTCC	ACCTCCATCG	5150
TGGGACCAA	TGTGGAAGTG	TCTCATACCG	CTGAAACCTA	CACCTGCACGG	5200
GCCAACACCC	CTGCTGTATA	GGCTAGGAGC	CGTCCAAAAT	GAGGTCAATC	5250
TCACACACCC	CATAACTAAA	TACATCATGG	CATGCATGTC	GGCTGAOCTG	5300
GAGGTGCTCA	CTAGCACCTG	GGTGCTGGTA	GGCGGAGTCC	TTCAGGCTTT	5350
GGCCGCATAC	TGCTGACGA	CAGGCAGTGT	GGTCATTGTG	GGCAGGATCA	5400
TCTTGTCGGG	GAAGCCAGCT	GTCGTTCCCG	ACAGGGAAGT	CCTCTACCAG	5450
GAGTTGCGATG	AGATGGAAGA	GTGTGCCCTCA	CAACTTCCTT	ACATCGAGCA	5500
GGGAATGCAG	CTCGCCGAGC	AATTCAAGCA	AAAGGCGCTC	GGGTGTGTGC	5550
AAACGGCCAC	CAAGCAAGCG	GAGGCTGCTG	CTCCCGTGGT	GGAGTCCAAG	5600
TGGCGAGCCC	TTGAGACCTT	CTGGGCGAAG	CACATGTGGA	ATTTTCATCAG	5650
CGGAATACAG	TACCTAGCAG	GCTTATCCAC	TCTGCTGGA	AACCCGCGGA	5700

FIG. 7C

SUBSTITUTE SHEET (RULE 26)

HC-J4

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
TAGCATCATT	GATGGCATT	ACAGCTTCTA	TCACTAGCCC	GCTCACCACC	5750
CAAAACACCC	TCCGTGTTAA	CATCTTGGGG	GGATGGGTGG	CTGCCCCAAT	5800
CGCTCCTCCC	AGCGCTGCGT	CAGCTTTGGT	GGCGCGCGGC	ATCGCGGGAG	5850
CGGCTGTGGG	CAGCATAGGC	CTTGGGAAGG	TGCTCGTGGG	CATCTTGGGG	5900
GGCTATGCGG	CAGGGGTAGC	CGGCGCACTC	GTTGGCTTTA	AGGTCATGAG	5950
CGGCGAGGTG	CCCTCCACCG	AGGACCTGGT	CACTTACTTC	CCTGCCATCC	6000
TCTCTCCTGG	TGCCCCGGTC	GTCGGGGTGG	TGTGGCGAGC	AATACTGGGT	6050
CGGCAAGTGG	GCCCCGGGGA	GGGGGCTGTG	CAGTGGATGA	ACCGGCTGAT	6100
AGCGTTCCGT	TGCGGGGGTA	ACCAGTCTTC	CCCTAAGCAC	TATGTGCTGT	6150
AGAGCGAAGC	TGCAGCAAGT	GTCACCTAGA	TCTCTCTTAG	CCTTACCATC	6200
ACTCAACTGC	TGAAGCGGCT	CCACCAGTGG	ATTAATGAGG	ACTGCTCTAC	6250
GCCATGCTCC	GGCTCGTGGC	TAAGGGATGT	TTGGGATTGG	ATATGCAAGG	6300
TGTTGACTGA	CTTCAAGACC	TGGCTCCAGT	CCAACTCCT	GOOGCGGTTA	6350
CCGGGAGTCC	CTTTCCGTGC	ATGCCAACGC	GGGTACAAGG	GAGTCTGGCG	6400
GGGGGACGGC	ATCATGCAAA	CCACCTGCCC	ATGCGGAGCA	CAGATCGCCG	6450
GACATGTCAA	AAACGGTTCC	ATGAGGATCG	TAGGGCTTAG	AACCTGCAGC	6500
AACACGTGGC	ACGGAACGTT	CCCCATCAAC	GCATACACCA	CGGGACCTTG	6550
CACACCTTCC	CCGGCGCCCA	ACTATTCCAG	GGCGCTATGG	CGGGTGGCTG	6600
CTGAGGAGTA	CGTGGAGGTT	ACCGGTGTGG	GGGATTTCCA	CTACGTGAGC	6650
GGCATGACCA	CTGACAACGT	AAAGTGCCCA	TGCCAGGTTT	CGGCCCCCGA	6700
ATTCTTCACG	GAGGTGGATG	GAGTGCGGTT	GCACAGGTAC	GCTCCGGCGT	6750
GCAAACCTCT	TCTACGGGAG	GACGTACAGT	TCCAGGTGGG	GCTCAACCAA	6800
TACTTGGTGG	GGTGGCAGCT	CCCATGCGAG	CCCGAACCGG	ACGTAAACAGT	6850
GCTTACTTCC	ATGCTCACCG	ATCCCTCCCA	CATTACAGCA	GAGACGGCTA	6900
AGCGTAGGCT	GGCTAGAGGG	TCTCCCCCT	CTTTAGCCAG	CTCATCAGCT	6950
AGCCAGTTGT	CTGGCCCTTC	TTTGAAGGGG	ACATGCACTA	CCCACCATGA	7000
CTCCCCGGAC	GCTGACCTCA	TGAGGGCCAA	CCTCTTGTGG	CGGCAGGAGA	7050
TGGGCGGAAA	CATCACTCGC	GTGGAGTCAG	AGAATAAGGT	AGTAATTCTG	7100
GACTCTTTGG	AACCGCTTCA	CGCGGAGGGG	GATGAGAGGG	AGATATCCGT	7150
CGCGGCGGAG	ATCCTGGGAA	AATCCAGGAA	GTTCCCTTCA	GCGTTGCCCC	7200
TATGGGCACG	CCCGGACTAC	AATCCTCCAC	TGCTAGAGTC	CTGGAAGGAC	7250
CCGGACTACG	TCCCTCCGGT	GGTACACGGA	TGCCCCATTG	CACCTACCAA	7300
GGCTCCTCCA	ATACCACCTC	CACGGAGAAA	GAGGACGGTT	GTCCTGACAG	7350
AATCCAATGT	GTCTTCTGCC	TTGGCGGAGC	TGCGCACTAA	GACCTTGGGT	7400
AGCTCCGGAT	CGTGGGCCGT	TGATAGCGGC	ACGGCGACCG	CCCTTCTCTA	7450
CCTGGCCTCC	GACGACGGTG	ACAAAGGATC	CGACGTTGAG	TGCTACTCCT	7500
CCATGCCCCC	CCTTGAAGGG	GAGCCGGGGG	ACCCCGATCT	CAGCGACGGG	7550
TCTTGGTCTA	CCGTGAGTGA	GGAGGCTAGT	GAGGATGTGG	TCTGCTGCTC	7600

FIG. 7D

SUBSTITUTE SHEET (RULE 26)

HC-J4

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
AATGTCTAT	ACGTGGACAG	GCGCCCTGAT	CACGCCATGC	GCTGCGGAGG	7650
AAAGTAAAGCT	GCCCCATCAAC	CCGTTTGAGCA	ACTCTTTTGCT	GCGTCACCAC	7700
AACATGGTCT	ACGCCACAAC	ATCCCCGAGC	GCAAGCCTCC	GGCAGAAGAA	7750
GGTCACCTTT	GACAGATTGC	AAGTOCTGGA	TGATCATTAC	CGGGACGTAC	7800
TCAAGGAGAT	GAAGGCGAAG	GCGTCCACAG	TTAAGGCTAA	GCTTCTATCT	7850
ATAGAGGAGG	CCTGCAAGCT	GACGCCCCCA	CATTGGGCGA	AATOCAAATT	7900
TGGCTATGGG	GCAAAGGACG	TCCGGAACCT	ATCCAGCAGG	GCCGTTAACC	7950
ACATCCGCTC	CGTGTGGGAG	GACTTGCTGG	AAGACACTGA	AACAACAATT	8000
GACACCAACA	TCATGGCAAA	AAGTGAGGTT	TTCTGGGTCC	AACCAGAGAA	8050
GGGAGGCCGC	AAGCCAGCTC	GCCTTATCGT	ATTCCAGAC	CTGGGAGTTC	8100
GTGTATGCGA	GAAGATGGCC	CTTTACGAAG	TGGTCTCCAC	CCTTCTCAG	8150
GCCGTGATGG	GCTCCTCATA	CGGATTTCAA	TACTCCCCCA	AGCAGCGGGT	8200
CGAGTTCCCTG	GTGAATAAAT	GGAAATCAAA	GAAATGCCCT	ATGGGCTTCT	8250
CATATGACAC	CCGCTGTTTT	GACTCAACGG	TACTGAGAG	TGACATTCTT	8300
GTTGAGGAGT	CAATTTACCA	ATGTTGTGAC	TTGGCCCCCG	AGGCCAGACA	8350
GGCCATAAGG	TGGCTCACAG	AGCGGCTTTA	CATCGGGGGT	CCCCTGACTA	8400
ACTCAAAAGG	GCAGAACTGC	GGTTATCGCC	GGTGCCGGGC	AAGTGGCGTG	8450
CTGACGACTA	GCTGCGGTAA	TACCCCTACA	TGTTACTTGA	AGGCCACTGC	8500
AGCCTGTGGA	GCTGCAAAGC	TCCAGGACTG	CACGATGCTC	GTGAACGGAG	8550
ACGACCTTGT	CGTTATCTGT	GAAAGCGCGG	GAACCCAGGA	GGATGCGGGC	8600
GCCCTACGAG	CCTTCACGGA	GGCTATGACT	AGGTATTCCG	CCCCCCCCCG	8650
GGATCCGCCC	CAACCAGAAT	ACGACCTGGA	GCTGATAACA	TCATGTTTCT	8700
CCAATGTGTC	AGTCGCGCAC	GATGCATCTG	GCAAAAGGGT	ATACTACCTC	8750
ACCCGTGACC	CCACCACCCC	CCTTGCAACG	GCTGGGTGGG	AGACAGCTAG	8800
ACACACTCCA	ATCAACTCTT	GGCTAGGCAA	TATCATCATG	TATGCGCCCA	8850
CCCTATGGGC	AAGGATGATT	CTGATGACTC	ACTTTTCTC	CATCTTTCTA	8900
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CTACTCCATT	GAGCCACTTG	ACCTACCTCA	GATCATTGAA	CGACTCCATG	9000
GTCTTAGCGC	ATTTACACTC	CACAGTTACT	CTCCAGGTGA	GATCAATAGG	9050
GTGGCTTCAT	GCCTCAGGAA	ACTTGGGGTA	CCACCCCTGC	GAACTGGAG	9100
ACATCGGGCC	AGAAGTGTCC	GCGCTAAGCT	ACTGTCCAG	GGGGGGAGGG	9150
CCGCCACTTG	TGGCAGATAC	CTCTTTAACT	GGGCAGTAAG	GACCAAGCTT	9200
AAACTCACTC	CAATCCCGGC	CGCGTCCAG	CTGGACTTGT	CTGGCTGGTT	9250
CGTGGCTGGT	TACAGCGGGG	GAGACATATA	TCACAGCCTG	TCTGTGCCCC	9300
GACCCCGCTG	GTTTCCGTTG	TGCCCTACTCC	TACTTTCTGT	AGGGGTAGGC	9350
ATTTACCTGC	TCCCCAACCG	ATGAACGGGG	AGCTAACCCAC	TCCAGGCTTT	9400
AAGCCATTTC	CTGTTTTTTT	TTTTTTTTTT	TTTTTTTTTT	TCTTTTTTTT	9450
TTTCTTCTCT	TCTCTCTTT	TTTTCTTTTC	TTTTTCCCTT	CTTTAATGGT	9500

FIG. 7E

SUBSTITUTE SHEET (RULE 26)

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
GGCTCCATCT	TAGCCCTAGT	CACGGCTAGC	TGTGAAAGGT	CCGTGAGCOG	9550
CATGACTGCA	GAGAGTGCTG	ATACTGGCCT	CTCTGCAGAT	CATGT	9595

FIG. 7F

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RGSRPSWGPT	DPRRRSRNLG	KVIDTLTIGF	ADLMGYIPLV	GAPLGGAARA	150
LAHGVRLLED	GVNYATGNLP	GCSFSIFLLA	LLSCLITPAS	AYEVRNVSGI	200
YHVINDCSNS	SIVYEAADVI	MHTPGCVFCV	QEGNSSROW	ALITPILARN	250
ASVPTTIRR	HVDLLVGTAA	FCSAMYVGD	CGSIFLVSQL	FTFSRRHET	300
VQDNCSTYP	GHSVGHMAW	DMMTWSPTT	ALVVSQLLRI	PQAVDMVAG	350
AHWGLAGLA	YYSMVGNWAK	VLIVALLFAG	VDGETHTTGR	VAGHTTSGFT	400
SLFSSGASQK	IQLVNINGSW	HINRTALNCH	DSLQTGFFAA	LFYAHKFNS	450
GCPERMASCR	PIDWFAQGW	PITYTKFNSS	DQRPYCWHA	PRPGVVPAS	500
QVCGPVYCF	PSPVVGTTD	RSVPTYSWG	ENEIDVMLIN	NIRPPQGNWF	550
GCTWMNSTGF	TKTCGGPPON	IGGVGNRTLI	CPIDCFRHP	EATYIKCGSG	600
PWLTPRCLVD	YPYRLWHYFC	TLNFSIFKVR	MYVGVVHRL	NAAQNWIRGE	650
RONLEDRDRS	ELSPILLSTT	EWQILPCAFT	TLPALSTGLI	HLHQNVDMQ	700
YLYGVGSAFV	SFAIKWEYIL	LLFLLADAR	VCACIWMMLL	IAQAEAALEN	750
LVVLNAAVA	GAHGILSFLV	FFCAAWYIKG	RLAPGAAYAF	YGVWPLLLLL	800
LALPPRAYAL	DREMAASCGG	AVLVGLVFLT	LSPYKVFLT	RLIWWLQYFI	850
TRAEAHMQW	VPPLNVRGGR	DAIILLTCAV	HPELIFDITK	LLAILGLPLM	900
VLQAGITRVP	YFVRAQGLIR	ACMLVRKVAG	GHYVQVFMK	LGALTGTIVY	950
NHILPLRDA	HAGLRDLAVA	VEPVVFSAME	TKVITWGADT	AACGDIILGL	1000
PVSARRGKEI	FLGPADSLEG	QGWRLAPIT	AYSQQTRGVL	GCIITSLTGR	1050
DKNQVEGEVQ	VVSTATQSFL	ATCINGVCWT	VYHAGSKTL	AGPKGPITQM	1100
YTINVDLILVG	WQAPFGARSM	TPCSCGSSDL	YLVIRHADVI	PVRRRGDSRG	1150
SLSPRPVSY	LKGSSGGPLL	CPSGHVGVF	RAAVCTRGVA	KAVDFIPVES	1200
METIMRSPVF	TINSTPPAVP	QTFQVAHLHA	PTGSGKSTKV	PAAYAAQGYK	1250
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DELAAKLTGL	GLNAVAYYRG	LDVSVIPPIG	DVVVATDAL	MIGFTGDFDS	1450
VIDQNTCVTQ	TVDFSLDPTF	TIETTTVPQD	AVRSQRRGR	TGRGRSGIYR	1500
FVTPGERPSG	MFDSSVLCEC	YDAGCAWYEL	TPAETSVRLR	AYLNTPLFV	1550
CQDHLEFWES	VFTGLTHIDA	HFLSQIKQAG	DNFPYLVAYQ	ATVCARAQAP	1600
PPSWDQWAKC	LIRLKPTLHG	PTPLLYRLGA	VQNEVILTHP	ITKYIMACMS	1650
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LYQEFDEEMEE	CASQLPYIEQ	GMQLAEQFKQ	KALGLIQTAT	KQAEAAAPVV	1750
ESKWRALETF	WAKHMANFIS	GIQYLAGLST	LPGNPATASL	MAFTASTTSP	1800
LTTQNTLLFN	ILGGWAAQL	APPSAASAFV	GAGIAGAAGV	SIGLGKVLVD	1850
ILAGYGAGVA	GALVAFKVM	GEVPSTEDLV	NLLPAILSPG	ALVGVVCAA	1900

FIG. 7G

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PRLPGVPFLS	CQRGYKGWR	GDGIMQITCP	CGAQLAGHVK	NGSMRIVGPR	2050
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LLSTEEACKL	TPPHSAKSKF	GYGAKDVRNL	SSRAVNHIRS	WEDLLEDTE	2550
TPIDITTIMAK	SEVFCVQPEK	GGRKPARLIV	FPLDGVVCE	KMALYDVVST	2600
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FIG. 7H

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 Purcell, Robert

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<150> 60/137,693

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<170> PatentIn Ver. 2.1

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Gly Val Tyr Leu Leu Pro Arg Arg Gly Pro Arg Leu Gly Val Arg Ala
          35              40              45

Thr Arg Lys Thr Ser Glu Arg Ser Gln Pro Arg Gly Arg Arg Gln Pro
  50              55              60

Ile Pro Lys Asp Arg Arg Ser Thr Gly Lys Ser Trp Gly Lys Pro Gly
  65              70              75              80

Tyr Pro Trp Pro Leu Tyr Gly Asn Glu Gly Leu Gly Trp Ala Gly Trp
          85              90              95

Leu Leu Ser Pro Arg Gly Ser Arg Pro Ser Trp Gly Pro Asn Asp Pro
          100             105             110

Arg His Arg Ser Arg Asn Val Gly Lys Val Ile Asp Thr Leu Thr Cys
          115             120             125

Gly Phe Ala Asp Leu Met Gly Tyr Ile Pro Val Val Gly Ala Pro Leu
          130             135             140

Gly Gly Val Ala Arg Ala Leu Ala His Gly Val Arg Val Leu Glu Asp
          145             150             155             160

Gly Val Asn Phe Ala Thr Gly Asn Leu Pro Gly Cys Ser Phe Ser Ile
          165             170             175

Phe Leu Leu Ala Leu Leu Ser Cys Ile Thr Thr Pro Val Ser Ala Ala
          180             185             190

Glu Val Lys Asn Ile Ser Thr Gly Tyr Met Val Thr Asn Asp Cys Thr
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Asn Asp Ser Ile Thr Trp Gln Leu Gln Ala Ala Val Leu His Val Pro
          210             215             220

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	245	250	255
Gly Leu Arg Thr His Ile Asp Met Val Val Met Ser Ala Thr Leu Cys			
	260	265	270
Ser Ala Leu Tyr Val Gly Asp Leu Cys Gly Gly Val Met Leu Ala Ala			
	275	280	285
Gln Met Phe Ile Val Ser Pro Gln His His Trp Phe Val Gln Asp Cys			
	290	295	300
Asn Cys Ser Ile Tyr Pro Gly Thr Ile Thr Gly His Arg Met Ala Trp			
305	310	315	320
Asp Met Met Met Asn Trp Ser Pro Thr Ala Thr Met Ile Leu Ala Tyr			
	325	330	335
Ala Met Arg Val Pro Glu Val Ile Ile Asp Ile Ile Ser Gly Ala His			
	340	345	350
Trp Gly Val Met Phe Gly Leu Ala Tyr Phe Ser Met Gln Gly Ala Trp			
	355	360	365
Ala Lys Val Val Val Ile Leu Leu Leu Ala Ala Gly Val Asp Ala Arg			
	370	375	380
Thr His Thr Val Gly Gly Ser Ala Ala Gln Thr Thr Gly Arg Leu Thr			
385	390	395	400
Ser Leu Phe Asp Met Gly Pro Arg Gln Lys Ile Gln Leu Val Asn Thr			
	405	410	415
Asn Gly Ser Trp His Ile Asn Arg Thr Ala Leu Asn Cys Asn Asp Ser			
	420	425	430
Leu His Thr Gly Phe Ile Ala Ser Leu Phe Tyr Thr His Ser Phe Asn			
	435	440	445
Ser Ser Gly Cys Pro Glu Arg Met Ser Ala Cys Arg Ser Ile Glu Ala			
	450	455	460
Phe Arg Val Gly Trp Gly Ala Leu Gln Tyr Glu Asp Asn Val Thr Asn			
465	470	475	480

Pro Glu Asp Met Arg Pro Tyr Cys Trp His Tyr Pro Pro Arg Gln Cys
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Gly Val Val Ser Ala Lys Thr Val Cys Gly Pro Val Tyr Cys Phe Thr
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Pro Ser Pro Val Val Val Gly Thr Thr Asp Arg Leu Gly Ala Pro Thr
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Tyr Thr Trp Gly Glu Asn Glu Thr Asp Val Phe Leu Leu Asn Ser Thr
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Arg Pro Pro Leu Gly Ser Trp Phe Gly Cys Thr Trp Met Asn Ser Ser
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Gly Tyr Thr Lys Thr Cys Gly Ala Pro Pro Cys Arg Thr Arg Ala Asp
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Phe Asn Ala Ser Thr Asp Leu Leu Cys Pro Thr Asp Cys Phe Arg Lys
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His Pro Asp Thr Thr Tyr Leu Lys Cys Gly Ser Gly Pro Trp Leu Thr
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Pro Arg Cys Leu Ile Asp Tyr Pro Tyr Arg Leu Trp His Tyr Pro Cys
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Thr Val Asn Tyr Thr Ile Phe Lys Ile Arg Met Tyr Val Gly Gly Val
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Glu His Arg Leu Thr Ala Ala Cys Asn Phe Thr Arg Gly Asp Arg Cys
 645 650 655

Asn Leu Glu Asp Arg Asp Arg Ser Gln Leu Ser Pro Leu Leu His Ser
 660 665 670

Thr Thr Glu Trp Ala Ile Leu Pro Cys Ser Tyr Ser Asp Leu Pro Ala
 675 680 685

Leu Ser Thr Gly Leu Leu His Leu His Gln Asn Ile Val Asp Val Gln
 690 695 700

Phe Met Tyr Gly Leu Ser Pro Ala Leu Thr Lys Tyr Ile Val Arg Trp
 705 710 715 720

Glu Trp Val Ile Leu Leu Phe Leu Leu Leu Ala Asp Ala Arg Val Cys
 725 730 735

Ala Cys Leu Trp Met Leu Ile Leu Leu Gly Gln Ala Glu Ala Ala Leu	740	745	750
Glu Lys Leu Val Ile Leu His Ala Ala Ser Ala Ala Ser Cys Asn Gly	755	760	765
Phe Leu Tyr Phe Val Ile Phe Phe Val Ala Ala Trp Tyr Ile Lys Gly	770	775	780
Arg Val Val Pro Leu Ala Thr Tyr Ser Leu Thr Gly Leu Trp Ser Phe	785	790	795
Ser Leu Leu Leu Leu Ala Leu Pro Gln Gln Ala Tyr Ala Tyr Asp Ala	805	810	815
Ser Val His Gly Gln Ile Gly Ala Ala Leu Leu Val Met Ile Thr Leu	820	825	830
Phe Thr Leu Thr Pro Gly Tyr Lys Thr Leu Leu Ser Arg Phe Leu Trp	835	840	845
Trp Leu Cys Tyr Leu Leu Thr Leu Gly Glu Ala Met Val Gln Glu Trp	850	855	860
Ala Pro Pro Met Gln Val Arg Gly Gly Arg Asp Gly Ile Ile Trp Ala	865	870	875
Val Ala Ile Phe Tyr Pro Gly Val Val Phe Asp Ile Thr Lys Trp Leu	885	890	895
Leu Ala Val Leu Gly Pro Ala Tyr Leu Leu Lys Gly Ala Leu Thr Arg	900	905	910
Val Pro Tyr Phe Val Arg Ala His Ala Leu Leu Arg Met Cys Thr Met	915	920	925
Ala Arg His Leu Ala Gly Gly Arg Tyr Val Gln Met Ala Leu Leu Ala	930	935	940
Leu Gly Arg Trp Thr Gly Thr Tyr Ile Tyr Asp His Leu Thr Pro Met	945	950	955
Ser Asp Trp Ala Ala Ser Gly Leu Arg Asp Leu Ala Val Ala Val Glu	965	970	975
Pro Ile Ile Phe Ser Pro Met Glu Lys Lys Val Ile Val Trp Gly Ala	980	985	990

Glu Thr Ala Ala Cys Gly Asp Ile Leu His Gly Leu Pro Val Ser Ala
 995 1000 1005

Arg Leu Gly Arg Glu Val Leu Leu Gly Pro Ala Asp Gly Tyr Thr Ser
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Lys Gly Trp Ser Leu Leu Ala Pro Ile Thr Ala Tyr Ala Gln Gln Thr
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Arg Gly Leu Leu Gly Thr Ile Val Val Ser Met Thr Gly Arg Asp Lys
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Pro Val Glu Thr Leu Asp Ile Val Thr Arg Ser Pro Thr Phe Ser Asp
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His Ala Pro Thr Gly Ser Gly Lys Ser Thr Lys Val Pro Val Ala Tyr
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Asn Ile Arg Thr Gly Val Arg Thr Val Thr Thr Gly Ala Pro Ile Thr
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Tyr Ser Thr Tyr Gly Lys Phe Leu Ala Asp Gly Gly Cys Ala Gly Gly
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Ala Tyr Asp Ile Ile Ile Cys Asp Glu Cys His Ala Val Asp Ser Thr
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Thr Ile Leu Gly Ile Gly Thr Val Leu Asp Gln Ala Glu Thr Ala Gly
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Val Arg Leu Thr Val Leu Ala Thr Ala Thr Pro Pro Gly Ser Val Thr
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Thr Pro His Pro Asn Ile Glu Glu Val Ala Leu Gly Gln Glu Gly Glu
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Ile Pro Phe Tyr Gly Arg Ala Ile Pro Leu Ser Tyr Ile Lys Gly Gly
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Asp Cys Asn Val Ala Val Thr Gln Val Val Asp Phe Ser Leu Asp Pro
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Thr Phe Thr Ile Thr Thr Gln Ile Val Pro Gln Asp Ala Val Ser Arg
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Ser Gln Arg Arg Gly Arg Thr Gly Arg Gly Arg Leu Gly Ile Tyr Arg
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Tyr Val Ser Thr Gly Glu Arg Ala Ser Gly Met Phe Asp Ser Val Val
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Leu Cys Glu Cys Tyr Asp Ala Gly Ala Ala Trp Tyr Glu Leu Thr Pro
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Ser Glu Thr Thr Val Arg Leu Arg Ala Tyr Phe Asn Thr Pro Gly Leu
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Pro Val Cys Gln Asp His Leu Glu Phe Trp Glu Ala Val Phe Thr Gly
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Leu Thr His Ile Asp Ala His Phe Leu Ser Gln Thr Lys Gln Ser Gly
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Glu Asn Phe Ala Tyr Leu Thr Ala Tyr Gln Ala Thr Val Cys Ala Arg
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Ala Lys Ala Pro Pro Pro Ser Trp Asp Val Met Trp Lys Cys Leu Thr
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Arg Leu Lys Pro Thr Leu Val Gly Pro Thr Pro Leu Leu Tyr Arg Leu
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Gly Ser Val Thr Asn Glu Val Thr Leu Thr His Pro Val Thr Lys Tyr
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Val Leu Ala Gly Gly Val Leu Ala Ala Val Ala Ala Tyr Cys Leu Ala
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Thr Gly Cys Val Cys Ile Ile Gly Arg Leu His Ile Asn Gln Arg Ala
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Val Val Ala Pro Asp Lys Glu Val Leu Tyr Glu Ala Phe Asp Glu Met
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Glu Glu Cys Ala Ser Arg Ala Ala Leu Ile Glu Glu Gly Gln Arg Ile
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Ala Glu Met Leu Lys Ser Lys Ile Gln Gly Leu Leu Gln Gln Ala Ser
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Lys Gln Ala Gln Asp Ile Gln Pro Thr Val Gln Ala Ser Trp Pro Lys
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Gln Tyr Leu Ala Gly Leu Ser Thr Leu Pro Gly Asn Pro Ala Val Ala
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Ser Met Met Ala Phe Ser Ala Ala Leu Thr Ser Pro Leu Ser Thr Ser
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Thr Thr Ile Leu Leu Asn Ile Leu Gly Gly Trp Leu Ala Ser Gln Ile
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Ala Pro Pro Ala Gly Ala Thr Gly Phe Val Val Ser Gly Leu Val Gly
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Ala Ala Val Gly Ser Ile Gly Leu Gly Lys Val Leu Val Asp Ile Leu
 1845 1850 1855

Ala Gly Tyr Gly Ala Gly Ile Ser Gly Ala Leu Val Ala Phe Lys Ile
 1860 1865 1870

Met Ser Gly Glu Lys Pro Ser Met Glu Asp Val Val Asn Leu Leu Pro
 1875 1880 1885

Gly Ile Leu Ser Pro Gly Ala Leu Val Val Gly Val Ile Cys Ala Ala
 1890 1895 1900

Ile Leu Arg Arg His Val Gly Pro Gly Glu Gly Ala Val Gln Trp Met
 1905 1910 1915 1920

Asn Arg Leu Ile Ala Phe Ala Ser Arg Gly Asn His Val Ala Pro Thr
 1925 1930 1935

His Tyr Val Thr Glu Ser Asp Ala Ser Gln Arg Val Thr Gln Leu Leu
 1940 1945 1950

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Trp Asp Trp Val Cys Thr Ile Leu Thr Asp Phe Lys Asn Trp Leu Thr
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Ser Lys Leu Phe Pro Lys Met Pro Gly Leu Pro Phe Val Ser Cys Gln
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<211> 3015

<212> PRT

<213> Hepatitis C virus

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Gly Val Tyr Leu Leu Pro Arg Arg Gly Pro Arg Leu Gly Val Arg Ala
35 40 45

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Thr Arg Lys Thr Ser Glu Arg Ser Gln Pro Arg Gly Arg Arg Gln Pro
50 55 60

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Ile Pro Lys Asp Arg Arg Ser Thr Gly Lys Ser Trp Gly Lys Pro Gly
65 70 75 80

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Tyr Pro Trp Pro Leu Tyr Gly Asn Glu Gly Leu Gly Trp Ala Gly Trp

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21

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Trp Gly Val Met Phe Gly Leu Ala Tyr Phe Ser Met Gln Gly Ala Trp		
355	360	365
Ala Lys Val Val Val Ile Leu Leu Leu Ala Ala Gly Val Asp Ala Arg		
370	375	380
Thr His Thr Val Gly Gly Ser Ala Ala Gln Thr Thr Gly Arg Leu Thr		
385	390	395
Ser Leu Phe Asp Met Gly Pro Arg Gln Lys Ile Gln Leu Val Asn Thr		
405	410	415
Asn Gly Ser Trp His Ile Asn Arg Thr Ala Leu Asn Cys Asn Asp Ser		
420	425	430
Leu His Thr Gly Phe Ile Ala Ser Leu Phe Tyr Thr His Ser Phe Asn		
435	440	445
Ser Ser Gly Cys Pro Glu Arg Met Ser Ala Cys Arg Ser Ile Glu Ala		
450	455	460
Phe Arg Val Gly Trp Gly Ala Leu Gln Tyr Glu Asp Asn Val Thr Asn		
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Pro Glu Asp Met Arg Pro Tyr Cys Trp His Tyr Pro Pro Arg Gln Cys		
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Pro Ser Pro Val Val Val Gly Thr Thr Asp Arg Leu Gly Ala Pro Thr		
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Tyr Thr Trp Gly Glu Asn Glu Thr Asp Val Phe Leu Leu Asn Ser Thr		
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Arg Pro Pro Leu Gly Ser Trp Phe Gly Cys Thr Trp Met Asn Ser Ser		
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Gly Tyr Thr Lys Thr Cys Gly Ala Pro Pro Cys Arg Thr Arg Ala Asp		
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Phe Asn Ala Ser Thr Asp Leu Leu Cys Pro Thr Asp Cys Phe Arg Lys		
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His Pro Asp Thr Thr Tyr Leu Lys Cys Gly Ser Gly Pro Trp Leu Thr		

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Pro Arg Cys Leu Ile Asp Tyr Pro Tyr Arg Leu Trp His Tyr Pro Cys		
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Glu His Arg Leu Thr Ala Ala Cys Asn Phe Thr Arg Gly Asp Arg Cys		
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Asn Leu Glu Asp Arg Asp Arg Ser Gln Leu Ser Pro Leu Leu His Ser		
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Thr Thr Glu Trp Ala Ile Leu Pro Cys Ser Tyr Ser Asp Leu Pro Ala		
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Phe Met Tyr Gly Leu Ser Pro Ala Leu Thr Lys Tyr Ile Val Arg Trp		
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Leu Thr Leu Ser Pro Tyr Tyr Lys Arg Tyr Ile Ser Trp Cys Met Trp		
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Trp Leu Gln Tyr Phe Leu Thr Arg Val Glu Ala Gln Leu His Val Trp		

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Leu Ala Ile Phe Gly	Pro Leu Trp Ile Leu Gln Ala Ser	Leu Leu Lys
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Val Pro Tyr Phe Val	Arg Val Gln Gly Leu Leu Arg Ile Cys Ala	Leu
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Ala Arg Lys Ile Ala	Gly Gly His Tyr Val Gln Met Ala Ile Ile	Lys
930	935	940
Leu Gly Ala Leu Thr	Gly Thr Tyr Val Tyr Asn His Leu Thr Pro	Leu
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Pro Val Val Phe Ser	Arg Met Glu Thr Lys Leu Ile Thr Trp Gly Ala	
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Asp Thr Ala Ala Cys	Gly Asp Ile Ile Asn Gly Leu Pro Val Ser	Ala
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Arg Arg Gly Gln Glu	Ile Leu Leu Gly Pro Ala Asp Gly Met Val	Ser
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Asn Gln Val Glu Gly	Glu Val Gln Ile Val Ser Thr Ala Thr Gln	Thr
1060	1065	1070
Phe Leu Ala Thr Cys	Ile Asn Gly Val Cys Trp Thr Val Tyr His	Gly
1075	1080	1085
Ala Gly Thr Arg Thr	Ile Ala Ser Pro Lys Gly Pro Val Ile Gln	Met
1090	1095	1100
Tyr Thr Asn Val Asp	Gln Asp Leu Val Gly Trp Pro Ala Pro Gln	Gly

1105	1110	1115	1120
Ser Arg Ser Leu Thr Pro Cys Thr Cys Gly Ser Ser Asp Leu Tyr Leu	1125	1130	1135
Val Thr Arg His Ala Asp Val Ile Pro Val Arg Arg Arg Gly Asp Ser	1140	1145	1150
Arg Gly Ser Leu Leu Ser Pro Arg Pro Ile Ser Tyr Leu Lys Gly Ser	1155	1160	1165
Ser Gly Gly Pro Leu Leu Cys Pro Ala Gly His Ala Val Gly Leu Phe	1170	1175	1180
Arg Ala Ala Val Cys Thr Arg Gly Val Ala Lys Ala Val Asp Phe Ile	1185	1190	1195
Pro Val Glu Asn Leu Gly Thr Thr Met Arg Ser Pro Val Phe Thr Asp	1205	1210	1215
Asn Ser Ser Pro Pro Ala Val Pro Gln Ser Phe Gln Val Ala His Leu	1220	1225	1230
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Ala Ala Gln Gly Tyr Lys Val Leu Val Leu Asn Pro Ser Val Ala Ala	1250	1255	1260
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Ser Ser Glu Cys Thr Thr Pro Cys Ser Gly Ser Trp Leu Arg Asp Ile		
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Ala Lys Leu Met Pro Gln Leu Pro Gly Ile Pro Phe Val Ser Cys Gln		
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Arg Ile Val Gly Pro Arg Thr Cys Arg Asn Met Trp Ser Gly Thr Phe		
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Asn Tyr Lys Phe Ala Leu Trp Arg Val Ser Ala Glu Glu Tyr Val Glu		
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Ile Arg Arg Val Gly Asp Phe His Tyr Val Ser Gly Met Thr Thr Asp		
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Leu Asp Gly Val Arg Leu His Arg Phe Ala Pro Pro Cys Lys Pro Leu		

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Gln Leu Ser Ala Pro Ser Leu Lys Ala Thr Cys Thr Ala Asn His Asp			
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Ser Pro Asp Ala Glu Leu Ile Glu Ala Asn Leu Leu Trp Arg Gln Glu			
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Gly Val Tyr Leu Leu Pro Arg Arg Gly Pro Arg Leu Gly Val Arg Ala
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Thr Arg Lys Thr Ser Glu Arg Ser Gln Pro Arg Gly Arg Arg Gln Pro
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Ile Pro Lys Asp Arg Arg Ser Thr Gly Lys Ser Trp Gly Lys Pro Gly
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Tyr Pro Trp Pro Leu Tyr Gly Asn Glu Gly Leu Gly Trp Ala Gly Trp
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Leu Leu Ser Pro Arg Gly Ser Arg Pro Ser Trp Gly Pro Asn Asp Pro
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Arg His Arg Ser Arg Asn Val Gly Lys Val Ile Asp Thr Leu Thr Cys
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Gly Phe Ala Asp Leu Met Gly Tyr Ile Pro Val Val Gly Ala Pro Leu
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Gly Gly Val Ala Arg Ala Leu Ala His Gly Val Arg Val Leu Glu Asp
 145 150 155 160

Gly Val Asn Phe Ala Thr Gly Asn Leu Pro Gly Cys Ser Phe Ser Ile
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Phe Leu Leu Ala Leu Leu Ser Cys Ile Thr Thr Pro Val Ser Ala Ala
 180 185 190

Glu Val Lys Asn Ile Ser Thr Gly Tyr Met Val Thr Asn Asp Cys Thr
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Asn Asp Ser Ile Thr Trp Gln Leu Gln Ala Ala Val Leu His Val Pro
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Gly Cys Val Pro Cys Glu Lys Val Gly Asn Ala Ser Gln Cys Trp Ile
 225 230 235 240

Pro Val Ser Pro Asn Val Ala Val Gln Arg Pro Gly Ala Leu Thr Gln
 245 250 255

Gly Leu Arg Thr His Ile Asp Met Val Val Met Ser Ala Thr Leu Cys
 260 265 270

Ser Ala Leu Tyr Val Gly Asp Leu Cys Gly Gly Val Met Leu Ala Ala
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Gln Met Phe Ile Val Ser Pro Gln His His Trp Phe Val Gln Asp Cys
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Asn Cys Ser Ile Tyr Pro Gly Thr Ile Thr Gly His Arg Met Ala Trp
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Asp Met Met Met Asn Trp Ser Pro Thr Ala Thr Met Ile Leu Ala Tyr
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Ala Met Arg Val Pro Glu Val Ile Ile Asp Ile Ile Ser Gly Ala His
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Trp Gly Val Met Phe Gly Leu Ala Tyr Phe Ser Met Gln Gly Ala Trp
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Ala Lys Val Val Val Ile Leu Leu Leu Ala Ala Gly Val Asp Ala Arg
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Thr His Thr Val Gly Gly Ser Ala Ala Gln Thr Thr Gly Arg Leu Thr
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Ser Leu Phe Asp Met Gly Pro Arg Gln Lys Ile Gln Leu Val Asn Thr
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Asn Gly Ser Trp His Ile Asn Arg Thr Ala Leu Asn Cys Asn Asp Ser
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Leu His Thr Gly Phe Ile Ala Ser Leu Phe Tyr Thr His Ser Phe Asn
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Ser Ser Gly Cys Pro Glu Arg Met Ser Ala Cys Arg Ser Ile Glu Ala
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Phe Arg Val Gly Trp Gly Ala Leu Gln Tyr Glu Asp Asn Val Thr Asn
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Pro	Glu	Asp	Met	Arg	Pro	Tyr	Cys	Trp	His	Tyr	Pro	Pro	Arg	Gln	Cys	485	490	495	
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Pro	Ser	Pro	Val	Val	Val	Gly	Thr	Thr	Asp	Arg	Leu	Gly	Ala	Pro	Thr	515	520	525	
Tyr	Thr	Trp	Gly	Glu	Asn	Glu	Thr	Asp	Val	Phe	Leu	Leu	Asn	Ser	Thr	530	535	540	
Arg	Pro	Pro	Leu	Gly	Ser	Trp	Phe	Gly	Cys	Thr	Trp	Met	Asn	Ser	Ser	545	550	555	560
Gly	Tyr	Thr	Lys	Thr	Cys	Gly	Ala	Pro	Pro	Cys	Arg	Thr	Arg	Ala	Asp	565	570	575	
Phe	Asn	Ala	Ser	Thr	Asp	Leu	Leu	Cys	Pro	Thr	Asp	Cys	Phe	Arg	Lys	580	585	590	
His	Pro	Asp	Thr	Thr	Tyr	Leu	Lys	Cys	Gly	Ser	Gly	Pro	Trp	Leu	Thr	595	600	605	
Pro	Arg	Cys	Leu	Ile	Asp	Tyr	Pro	Tyr	Arg	Leu	Trp	His	Tyr	Pro	Cys	610	615	620	
Thr	Val	Asn	Tyr	Thr	Ile	Phe	Lys	Ile	Arg	Met	Tyr	Val	Gly	Gly	Val	625	630	635	640
Glu	His	Arg	Leu	Thr	Ala	Ala	Cys	Asn	Phe	Thr	Arg	Gly	Asp	Arg	Cys	645	650	655	
Asn	Leu	Glu	Asp	Arg	Asp	Arg	Ser	Gln	Leu	Ser	Pro	Leu	Leu	His	Ser	660	665	670	
Thr	Thr	Glu	Trp	Ala	Ile	Leu	Pro	Cys	Ser	Tyr	Ser	Asp	Leu	Pro	Ala	675	680	685	
Leu	Ser	Thr	Gly	Leu	Leu	His	Leu	His	Gln	Asn	Ile	Val	Asp	Val	Gln	690	695	700	
Phe	Met	Tyr	Gly	Leu	Ser	Pro	Ala	Leu	Thr	Lys	Tyr	Ile	Val	Arg	Trp	705	710	715	720
Glu	Trp	Val	Ile	Leu	Leu	Phe	Leu	Leu	Leu	Ala	Asp	Ala	Arg	Val	Cys	725	730	735	

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Met Cys Val Val His Pro Thr Leu Val Phe Asp Ile Thr Lys Leu Leu
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Thr Leu Gly Phe Gly Ala Tyr Met Ser Lys Ala His Gly Val Asp Pro
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Asn Ile Arg Thr Gly Val Arg Thr Ile Thr Thr Gly Ser Pro Ile Thr
 1285 1290 1295

Tyr Ser Thr Tyr Gly Lys Phe Leu Ala Asp Gly Gly Cys Ser Gly Gly
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Ser Ile Leu Gly Ile Gly Thr Val Leu Asp Gln Ala Glu Thr Ala Gly
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gcagatcatg t 9611

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<211> 3015

<212> PRT

<213> Hepatitis C virus

<400> 8

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```

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Arg Arg Pro Gln Asp Val Lys Phe Pro Gly Gly Gly Gln Ile Val Gly
      20              25              30

```

```

Gly Val Tyr Leu Leu Pro Arg Arg Gly Pro Arg Leu Gly Val Arg Ala
      35              40              45

```

```

Thr Arg Lys Thr Ser Glu Arg Ser Gln Pro Arg Gly Arg Arg Gln Pro
      50              55              60

```

```

Ile Pro Lys Asp Arg Arg Ser Thr Gly Lys Ser Trp Gly Lys Pro Gly
      65              70              75              80

```

```

Tyr Pro Trp Pro Leu Tyr Gly Asn Glu Gly Leu Gly Trp Ala Gly Trp
      85              90              95

```

Leu Leu Ser Pro Arg Gly Ser Arg Pro Ser Trp Gly Pro Asn Asp Pro
 100 105 110

Arg His Arg Ser Arg Asn Val Gly Lys Val Ile Asp Thr Leu Thr Cys
 115 120 125

Gly Phe Ala Asp Leu Met Gly Tyr Ile Pro Val Val Gly Ala Pro Leu
 130 135 140

Gly Gly Val Ala Arg Ala Leu Ala His Gly Val Arg Val Leu Glu Asp
 145 150 155 160

Gly Val Asn Phe Ala Thr Gly Asn Leu Pro Gly Cys Ser Phe Ser Ile
 165 170 175

Phe Leu Leu Ala Leu Leu Ser Cys Ile Thr Thr Pro Val Ser Ala Ala
 180 185 190

Glu Val Lys Asn Ile Ser Thr Gly Tyr Met Val Thr Asn Asp Cys Thr
 195 200 205

Asn Asp Ser Ile Thr Trp Gln Leu Gln Ala Ala Val Leu His Val Pro
 210 215 220

Gly Cys Val Pro Cys Glu Lys Val Gly Asn Ala Ser Gln Cys Trp Ile
 225 230 235 240

Pro Val Ser Pro Asn Val Ala Val Gln Arg Pro Gly Ala Leu Thr Gln
 245 250 255

Gly Leu Arg Thr His Ile Asp Met Val Val Met Ser Ala Thr Leu Cys
 260 265 270

Ser Ala Leu Tyr Val Gly Asp Leu Cys Gly Gly Val Met Leu Ala Ala
 275 280 285

Gln Met Phe Ile Val Ser Pro Gln His His Trp Phe Val Gln Asp Cys
 290 295 300

Asn Cys Ser Ile Tyr Pro Gly Thr Ile Thr Gly His Arg Met Ala Trp
 305 310 315 320

Asp Met Met Met Asn Trp Ser Pro Thr Ala Thr Met Ile Leu Ala Tyr
 325 330 335

Ala Met Arg Val Pro Glu Val Ile Ile Asp Ile Ile Ser Gly Ala His
 340 345 350

Trp Gly Val Met Phe Gly Leu Ala Tyr Phe Ser Met Gln Gly Ala Trp
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Ala Lys Val Val Val Ile Leu Leu Ala Ala Gly Val Asp Ala Arg
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Thr His Thr Val Gly Gly Ser Ala Ala Gln Thr Thr Gly Arg Leu Thr
 385 390 395 400

Ser Leu Phe Asp Met Gly Pro Arg Gln Lys Ile Gln Leu Val Asn Thr
 405 410 415

Asn Gly Ser Trp His Ile Asn Arg Thr Ala Leu Asn Cys Asn Asp Ser
 420 425 430

Leu His Thr Gly Phe Ile Ala Ser Leu Phe Tyr Thr His Ser Phe Asn
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Ser Ser Gly Cys Pro Glu Arg Met Ser Ala Cys Arg Ser Ile Glu Ala
 450 455 460

Phe Arg Val Gly Trp Gly Ala Leu Gln Tyr Glu Asp Asn Val Thr Asn
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Pro Glu Asp Met Arg Pro Tyr Cys Trp His Tyr Pro Pro Arg Gln Cys
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Gly Val Val Ser Ala Lys Thr Val Cys Gly Pro Val Tyr Cys Phe Thr
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Pro Ser Pro Val Val Val Gly Thr Thr Asp Arg Leu Gly Ala Pro Thr
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Tyr Thr Trp Gly Glu Asn Glu Thr Asp Val Phe Leu Leu Asn Ser Thr
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Arg Pro Pro Leu Gly Ser Trp Phe Gly Cys Thr Trp Met Asn Ser Ser
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Gly Tyr Thr Lys Thr Cys Gly Ala Pro Pro Cys Arg Thr Arg Ala Asp
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Phe Asn Ala Ser Thr Asp Leu Leu Cys Pro Thr Asp Cys Phe Arg Lys
 580 585 590

His Pro Asp Thr Thr Tyr Leu Lys Cys Gly Ser Gly Pro Trp Leu Thr
 595 600 605

Pro Arg Cys Leu Ile Asp Tyr Pro Tyr Arg Leu Trp His Tyr Pro Cys
 610 615 620
 Thr Val Asn Tyr Thr Ile Phe Lys Ile Arg Met Tyr Val Gly Gly Val
 625 630 635 640
 Glu His Arg Leu Thr Ala Ala Cys Asn Phe Thr Arg Gly Asp Arg Cys
 645 650 655
 Asn Leu Glu Asp Arg Asp Arg Ser Gln Leu Ser Pro Leu Leu His Ser
 660 665 670
 Thr Thr Glu Trp Ala Ile Leu Pro Cys Ser Tyr Ser Asp Leu Pro Ala
 675 680 685
 Leu Ser Thr Gly Leu Leu His Leu His Gln Asn Ile Val Asp Val Gln
 690 695 700
 Phe Met Tyr Gly Leu Ser Pro Ala Leu Thr Lys Tyr Ile Val Arg Trp
 705 710 715 720
 Glu Trp Val Ile Leu Leu Phe Leu Leu Leu Ala Asp Ala Arg Val Cys
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 Ala Cys Leu Trp Met Leu Ile Leu Leu Gly Gln Ala Glu Ala Ala Leu
 740 745 750
 Glu Lys Leu Val Ile Leu His Ala Ala Ser Ala Ala Ser Cys Asn Gly
 755 760 765
 Phe Leu Tyr Phe Val Ile Phe Phe Val Ala Ala Trp Tyr Ile Lys Gly
 770 775 780
 Arg Val Val Pro Leu Ala Thr Tyr Ser Leu Thr Gly Leu Trp Ser Phe
 785 790 795 800
 Ser Leu Leu Leu Leu Ala Leu Pro Gln Gln Ala Tyr Ala Leu Asp Thr
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 Glu Val Ala Ala Ser Cys Gly Gly Val Val Leu Val Gly Leu Met Ala
 820 825 830
 Leu Thr Leu Ser Pro Tyr Tyr Lys Arg Tyr Ile Ser Trp Cys Met Trp
 835 840 845
 Trp Leu Gln Tyr Phe Leu Thr Arg Val Glu Ala Gln Leu His Val Trp
 850 855 860

Val Pro Pro Leu Asn Val Arg Gly Gly Arg Asp Ala Val Ile Leu Leu			
865	870	875	880
Met Cys Val Val His Pro Thr Leu Val Phe Asp Ile Thr Lys Leu Leu			
	885	890	895
Leu Ala Ile Phe Gly Pro Leu Trp Ile Leu Gln Ala Ser Leu Leu Lys			
	900	905	910
Val Pro Tyr Phe Val Arg Val Gln Gly Leu Leu Arg Ile Cys Ala Leu			
	915	920	925
Ala Arg Lys Ile Ala Gly Gly His Tyr Val Gln Met Ala Ile Ile Lys			
	930	935	940
Leu Gly Ala Leu Thr Gly Thr Tyr Val Tyr Asn His Leu Thr Pro Leu			
945	950	955	960
Arg Asp Trp Ala His Asn Gly Leu Arg Asp Leu Ala Val Ala Val Glu			
	965	970	975
Pro Val Val Phe Ser Arg Met Glu Thr Lys Leu Ile Thr Trp Gly Ala			
	980	985	990
Asp Thr Ala Ala Cys Gly Asp Ile Ile Asn Gly Leu Pro Val Ser Ala			
	995	1000	1005
Arg Arg Gly Gln Glu Ile Leu Leu Gly Pro Ala Asp Gly Met Val Ser			
1010	1015	1020	
Lys Gly Trp Arg Leu Leu Ala Pro Ile Thr Ala Tyr Ala Gln Gln Thr			
1025	1030	1035	1040
Arg Gly Leu Leu Gly Cys Ile Ile Thr Ser Leu Thr Gly Arg Asp Lys			
	1045	1050	1055
Asn Gln Val Glu Gly Glu Val Gln Ile Val Ser Thr Ala Thr Gln Thr			
	1060	1065	1070
Phe Leu Ala Thr Cys Ile Asn Gly Val Cys Trp Thr Val Tyr His Gly			
	1075	1080	1085
Ala Gly Thr Arg Thr Ile Ala Ser Pro Lys Gly Pro Val Ile Gln Met			
	1090	1095	1100
Tyr Thr Asn Val Asp Gln Asp Leu Val Gly Trp Pro Ala Pro Gln Gly			
1105	1110	1115	1120

Ser Arg Ser Leu Thr Pro Cys Thr Cys Gly Ser Ser Asp Leu Tyr Leu
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Val Thr Arg His Ala Asp Val Ile Pro Val Arg Arg Arg Gly Asp Ser
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Arg Gly Ser Leu Leu Ser Pro Arg Pro Ile Ser Tyr Leu Lys Gly Ser
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Ser Gly Gly Pro Leu Leu Cys Pro Ala Gly His Ala Val Gly Leu Phe
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Arg Ala Ala Val Cys Thr Arg Gly Val Ala Lys Ala Val Asp Phe Ile
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Pro Val Glu Asn Leu Gly Thr Thr Met Arg Ser Pro Val Phe Thr Asp
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Asn Ser Ser Pro Pro Ala Val Pro Gln Ser Phe Gln Val Ala His Leu
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His Ala Pro Thr Gly Ser Gly Lys Ser Thr Lys Val Pro Ala Ala Tyr
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Ala Ala Gln Gly Tyr Lys Val Leu Val Leu Asn Pro Ser Val Ala Ala
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Thr Leu Gly Phe Gly Ala Tyr Met Ser Lys Ala His Gly Val Asp Pro
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Asn Ile Arg Thr Gly Val Arg Thr Ile Thr Thr Gly Ser Pro Ile Thr
 1285 1290 1295

Tyr Ser Thr Tyr Gly Lys Phe Leu Ala Asp Gly Gly Cys Ser Gly Gly
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Ala Tyr Asp Ile Ile Ile Cys Asp Glu Cys His Ser Thr Asp Ala Thr
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Ser Ile Leu Gly Ile Gly Thr Val Leu Asp Gln Ala Glu Thr Ala Gly
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Ala Arg Leu Val Val Leu Ala Thr Ala Thr Pro Pro Gly Ser Val Thr
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Val Ser His Pro Asn Ile Glu Glu Val Ala Leu Ser Thr Thr Gly Glu
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Ile Pro Phe Tyr Gly Lys Ala Ile Pro Leu Glu Val Ile Lys Gly Gly	1380	1385	1390
Arg His Leu Ile Phe Cys His Ser Lys Lys Lys Cys Asp Glu Leu Ala	1395	1400	1405
Ala Lys Leu Val Ala Leu Gly Ile Asn Ala Val Ala Tyr Tyr Arg Gly	1410	1415	1420
Leu Asp Val Ser Val Ile Pro Thr Ser Gly Asp Val Val Val Val Ser	1425	1430	1435 1440
Thr Asp Ala Leu Met Thr Gly Phe Thr Gly Asp Phe Asp Ser Val Ile	1445	1450	1455
Asp Cys Asn Thr Cys Val Thr Gln Thr Val Asp Phe Ser Leu Asp Pro	1460	1465	1470
Thr Phe Thr Ile Glu Thr Thr Thr Leu Pro Gln Asp Ala Val Ser Arg	1475	1480	1485
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Phe Val Ala Pro Gly Glu Arg Pro Ser Gly Met Phe Asp Ser Ser Val	1505	1510	1515 1520
Leu Cys Glu Cys Tyr Asp Ala Gly Cys Ala Trp Tyr Glu Leu Thr Pro	1525	1530	1535
Ala Glu Thr Thr Val Arg Leu Arg Ala Tyr Met Asn Thr Pro Gly Leu	1540	1545	1550
Pro Val Cys Gln Asp His Leu Glu Phe Trp Glu Gly Val Phe Thr Gly	1555	1560	1565
Leu Thr His Ile Asp Ala His Phe Leu Ser Gln Thr Lys Gln Ser Gly	1570	1575	1580
Glu Asn Phe Pro Tyr Leu Val Ala Tyr Gln Ala Thr Val Cys Ala Arg	1585	1590	1595 1600
Ala Gln Ala Pro Pro Pro Ser Trp Asp Gln Met Trp Lys Cys Leu Ile	1605	1610	1615
Arg Leu Lys Pro Thr Leu His Gly Pro Thr Pro Leu Leu Tyr Arg Leu	1620	1625	1630

Gly Ala Val Gln Asn Glu Val Thr Leu Thr His Pro Ile Thr Lys Tyr
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Ile Met Thr Cys Met Ser Ala Asp Leu Glu Val Val Thr Ser Thr Trp
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Val Leu Val Gly Gly Val Leu Ala Ala Leu Ala Ala Tyr Cys Leu Ser
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Thr Gly Cys Val Val Ile Val Gly Arg Ile Val Leu Ser Gly Lys Pro
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Ala Ile Ile Pro Asp Arg Glu Val Leu Tyr Gln Glu Phe Asp Glu Met
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Glu Glu Cys Ser Gln His Leu Pro Tyr Ile Glu Gln Gly Met Met Leu
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Ala Glu Gln Phe Lys Gln Lys Ala Leu Gly Leu Leu Gln Thr Ala Ser
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Arg His Ala Glu Val Ile Thr Pro Ala Val Gln Thr Asn Trp Gln Lys
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Leu Glu Val Phe Trp Ala Lys His Met Trp Asn Phe Ile Ser Gly Ile
 1765 1770 1775

Gln Tyr Leu Ala Gly Leu Ser Thr Leu Pro Gly Asn Pro Ala Ile Ala
 1780 1785 1790

Ser Leu Met Ala Phe Thr Ala Ala Val Thr Ser Pro Leu Thr Thr Gly
 1795 1800 1805

Gln Thr Leu Leu Phe Asn Ile Leu Gly Gly Trp Val Ala Ala Gln Leu
 1810 1815 1820

Ala Ala Pro Gly Ala Ala Thr Ala Phe Val Gly Ala Gly Leu Ala Gly
 1825 1830 1835 1840

Ala Ala Ile Gly Ser Val Gly Leu Gly Lys Val Leu Val Asp Ile Leu
 1845 1850 1855

Ala Gly Tyr Gly Ala Gly Val Ala Gly Ala Leu Val Ala Phe Lys Ile
 1860 1865 1870

Met Ser Gly Glu Val Pro Ser Thr Glu Asp Leu Val Asn Leu Leu Pro
 1875 1880 1885

Ala Ile Leu Ser Pro Gly Ala Leu Val Val Gly Val Val Cys Ala Ala
1890 1895 1900

Ile Leu Arg Arg His Val Gly Pro Gly Glu Gly Ala Val Gln Trp Met
1905 1910 1915 1920

Asn Arg Leu Ile Ala Phe Ala Ser Arg Gly Asn His Val Ser Pro Thr
1925 1930 1935

His Tyr Val Pro Glu Ser Asp Ala Ala Ala Arg Val Thr Ala Ile Leu
1940 1945 1950

Ser Ser Leu Thr Val Thr Gln Leu Leu Arg Arg Leu His Gln Trp Ile
1955 1960 1965

Ser Ser Glu Cys Thr Thr Pro Cys Ser Gly Ser Trp Leu Arg Asp Ile
1970 1975 1980

Trp Asp Trp Ile Cys Glu Val Leu Ser Asp Phe Lys Thr Trp Leu Lys
1985 1990 1995 2000

Ala Lys Leu Met Pro Gln Leu Pro Gly Ile Pro Phe Val Ser Cys Gln
2005 2010 2015

Arg Gly Tyr Arg Gly Val Trp Arg Gly Asp Gly Ile Met His Thr Arg
2020 2025 2030

Cys His Cys Gly Ala Glu Ile Thr Gly His Val Lys Asn Gly Thr Met
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Pro Ile Asn Ala Tyr Thr Thr Gly Pro Cys Thr Pro Leu Pro Ala Pro
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Asn Tyr Lys Phe Ala Leu Trp Arg Val Ser Ala Glu Glu Tyr Val Glu
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Leu Arg Glu Glu Val Ser Phe Arg Val Gly Leu His Glu Tyr Pro Val
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Gly Ser Gln Leu Pro Cys Glu Pro Glu Pro Asp Val Ala Val Leu Thr
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Ser Met Leu Thr Asp Pro Ser His Ile Thr Ala Glu Ala Ala Gly Arg
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Arg Leu Ala Arg Gly Ser Pro Pro Ser Met Ala Ser Ser Ser Ala Ser
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Gln Leu Ser Ala Pro Ser Leu Lys Ala Thr Cys Thr Ala Asn His Asp
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Met Gly Gly Asn Ile Thr Arg Val Glu Ser Glu Asn Lys Val Val Ile
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Leu Asp Ser Phe Asp Pro Leu Val Ala Glu Glu Asp Glu Arg Glu Val
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Ser Val Pro Ala Glu Ile Leu Arg Lys Ser Arg Arg Phe Ala Arg Ala
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Trp Lys Lys Pro Asp Tyr Glu Pro Pro Val Val His Gly Cys Pro Leu
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Pro Pro Pro Arg Ser Pro Pro Val Pro Pro Pro Arg Lys Lys Arg Thr
 2325 2330 2335

Val Val Leu Thr Glu Ser Thr Leu Ser Thr Ala Leu Ala Glu Leu Ala
 2340 2345 2350

Thr Lys Ser Phe Gly Ser Ser Ser Thr Ser Gly Ile Thr Gly Asp Asn
 2355 2360 2365

Thr Thr Thr Ser Ser Glu Pro Ala Pro Ser Gly Cys Pro Pro Asp Ser
 2370 2375 2380

Asp Val Glu Ser Tyr Ser Ser Met Pro Pro Leu Glu Gly Glu Pro Gly
 2385 2390 2395 2400

Asp Pro Asp Leu Ser Asp Gly Ser Trp Ser Thr Val Ser Ser Gly Ala			
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	370	375		380		
Thr His Thr Val Gly Gly Ser Ala Ala Gln Thr Thr Gly Arg Leu Thr						
	385	390		395		400
Ser Leu Phe Asp Met Gly Pro Arg Gln Lys Ile Gln Leu Val Asn Thr						
	405	410		415		
Asn Gly Ser Trp His Ile Asn Arg Thr Ala Leu Asn Cys Asn Asp Ser						
	420	425		430		
Leu His Thr Gly Phe Ile Ala Ser Leu Phe Tyr Thr His Ser Phe Asn						
	435	440		445		
Ser Ser Gly Cys Pro Glu Arg Met Ser Ala Cys Arg Ser Ile Glu Ala						
	450	455		460		
Phe Arg Val Gly Trp Gly Ala Leu Gln Tyr Glu Asp Asn Val Thr Asn						
	465	470		475		480
Pro Glu Asp Met Arg Pro Tyr Cys Trp His Tyr Pro Pro Arg Gln Cys						

69

70

995	1000	1005
Arg Arg Gly Gln Glu Ile Leu Leu Gly Pro Ala Asp Gly Met Val Ser		
1010	1015	1020
Lys Gly Trp Arg Leu Leu Ala Pro Ile Thr Ala Tyr Ala Gln Gln Thr		
1025	1030	1035 1040
Arg Gly Leu Leu Gly Cys Ile Ile Thr Ser Leu Thr Gly Arg Asp Lys		
1045	1050	1055
Asn Gln Val Glu Gly Glu Val Gln Ile Val Ser Thr Ala Thr Gln Thr		
1060	1065	1070
Phe Leu Ala Thr Cys Ile Asn Gly Val Cys Trp Thr Val Tyr His Gly		
1075	1080	1085
Ala Gly Thr Arg Thr Ile Ala Ser Pro Lys Gly Pro Val Ile Gln Met		
1090	1095	1100
Tyr Thr Asn Val Asp Gln Asp Leu Val Gly Trp Pro Ala Pro Gln Gly		
1105	1110	1115 1120
Ser Arg Ser Leu Thr Pro Cys Thr Cys Gly Ser Ser Asp Leu Tyr Leu		
1125	1130	1135
Val Thr Arg His Ala Asp Val Ile Pro Val Arg Arg Arg Gly Asp Ser		
1140	1145	1150
Arg Gly Ser Leu Leu Ser Pro Arg Pro Ile Ser Tyr Leu Lys Gly Ser		
1155	1160	1165
Ser Gly Gly Pro Leu Leu Cys Pro Ala Gly His Ala Val Gly Leu Phe		
1170	1175	1180
Arg Ala Ala Val Cys Thr Arg Gly Val Ala Lys Ala Val Asp Phe Ile		
1185	1190	1195 1200
Pro Val Glu Asn Leu Gly Thr Thr Met Arg Ser Pro Val Phe Thr Asp		
1205	1210	1215
Asn Ser Ser Pro Pro Ala Val Pro Gln Ser Phe Gln Val Ala His Leu		
1220	1225	1230
His Ala Pro Thr Gly Ser Gly Lys Ser Thr Lys Val Pro Ala Ala Tyr		
1235	1240	1245
Ala Ala Gln Gly Tyr Lys Val Leu Val Leu Asn Pro Ser Val Ala Ala		

1250	1255	1260
Thr Leu Gly Phe Gly Ala Tyr Met Ser Lys Ala His Gly Val Asp Pro		
1265	1270	1275 1280
Asn Ile Arg Thr Gly Val Arg Thr Ile Thr Thr Gly Ser Pro Ile Thr		
	1285	1290 1295
Tyr Ser Thr Tyr Gly Lys Phe Leu Ala Asp Gly Gly Cys Ser Gly Gly		
	1300	1305 1310
Ala Tyr Asp Ile Ile Ile Cys Asp Glu Cys His Ser Thr Asp Ala Thr		
	1315	1320 1325
Ser Ile Leu Gly Ile Gly Thr Val Leu Asp Gln Ala Glu Thr Ala Gly		
	1330	1335 1340
Ala Arg Leu Val Val Leu Ala Thr Ala Thr Pro Pro Gly Ser Val Thr		
	1345	1350 1355 1360
Val Ser His Pro Asn Ile Glu Glu Val Ala Leu Ser Thr Thr Gly Glu		
	1365	1370 1375
Ile Pro Phe Tyr Gly Lys Ala Ile Pro Leu Glu Val Ile Lys Gly Gly		
	1380	1385 1390
Arg His Leu Ile Phe Cys His Ser Lys Lys Lys Cys Asp Glu Leu Ala		
	1395	1400 1405
Ala Lys Leu Val Ala Leu Gly Ile Asn Ala Val Ala Tyr Tyr Arg Gly		
	1410	1415 1420
Leu Asp Val Ser Val Ile Pro Thr Ser Gly Asp Val Val Val Val Ser		
	1425	1430 1435 1440
Thr Asp Ala Leu Met Thr Gly Phe Thr Gly Asp Phe Asp Ser Val Ile		
	1445	1450 1455
Asp Cys Asn Thr Cys Val Thr Gln Thr Val Asp Phe Ser Leu Asp Pro		
	1460	1465 1470
Thr Phe Thr Ile Glu Thr Thr Thr Leu Pro Gln Asp Ala Val Ser Arg		
	1475	1480 1485
Thr Gln Arg Arg Gly Arg Thr Gly Arg Gly Lys Pro Gly Ile Tyr Arg		
	1490	1495 1500
Phe Val Ala Pro Gly Glu Arg Pro Ser Gly Met Phe Asp Ser Ser Val		

1505	1510	1515	1520
Leu Cys Glu Cys Tyr Asp Ala Gly Cys Ala Trp Tyr Glu Leu Thr Pro	1525	1530	1535
Ala Glu Thr Thr Val Arg Leu Arg Ala Tyr Met Asn Thr Pro Gly Leu	1540	1545	1550
Pro Val Cys Gln Asp His Leu Glu Phe Trp Glu Gly Val Phe Thr Gly	1555	1560	1565
Leu Thr His Ile Asp Ala His Phe Leu Ser Gln Thr Lys Gln Ser Gly	1570	1575	1580
Glu Asn Phe Pro Tyr Leu Val Ala Tyr Gln Ala Thr Val Cys Ala Arg	1585	1590	1595
Ala Gln Ala Pro Pro Pro Ser Trp Asp Gln Met Trp Lys Cys Leu Ile	1605	1610	1615
Arg Leu Lys Pro Thr Leu His Gly Pro Thr Pro Leu Leu Tyr Arg Leu	1620	1625	1630
Gly Ala Val Gln Asn Glu Val Thr Leu Thr His Pro Ile Thr Lys Tyr	1635	1640	1645
Ile Met Thr Cys Met Ser Ala Asp Leu Glu Val Val Thr Ser Thr Trp	1650	1655	1660
Val Leu Val Gly Gly Val Leu Ala Ala Leu Ala Ala Tyr Cys Leu Ser	1665	1670	1675
Thr Gly Cys Val Val Ile Val Gly Arg Ile Val Leu Ser Gly Lys Pro	1685	1690	1695
Ala Ile Ile Pro Asp Arg Glu Val Leu Tyr Gln Glu Phe Asp Glu Met	1700	1705	1710
Glu Glu Cys Ser Gln His Leu Pro Tyr Ile Glu Gln Gly Met Met Leu	1715	1720	1725
Ala Glu Gln Phe Lys Gln Lys Ala Leu Gly Leu Leu Gln Thr Ala Ser	1730	1735	1740
Arg His Ala Glu Val Ile Thr Pro Ala Val Gln Thr Asn Trp Gln Lys	1745	1750	1755
Leu Glu Val Phe Trp Ala Lys His Met Trp Asn Phe Ile Ser Gly Ile			

	1765	1770	1775
Gln Tyr Leu Ala Gly Leu Ser Thr Leu Pro Gly Asn Pro Ala Ile Ala	1780	1785	1790
Ser Leu Met Ala Phe Thr Ala Ala Val Thr Ser Pro Leu Thr Thr Gly	1795	1800	1805
Gln Thr Leu Leu Phe Asn Ile Leu Gly Gly Trp Val Ala Ala Gln Leu	1810	1815	1820
Ala Ala Pro Gly Ala Ala Thr Ala Phe Val Gly Ala Gly Leu Ala Gly	1825	1830	1835
Ala Ala Ile Gly Ser Val Gly Leu Gly Lys Val Leu Val Asp Ile Leu	1845	1850	1855
Ala Gly Tyr Gly Ala Gly Val Ala Gly Ala Leu Val Ala Phe Lys Ile	1860	1865	1870
Met Ser Gly Glu Val Pro Ser Thr Glu Asp Leu Val Asn Leu Leu Pro	1875	1880	1885
Ala Ile Leu Ser Pro Gly Ala Leu Val Val Gly Val Val Cys Ala Ala	1890	1895	1900
Ile Leu Arg Arg His Val Gly Pro Gly Glu Gly Ala Val Gln Trp Met	1905	1910	1915
Asn Arg Leu Ile Ala Phe Ala Ser Arg Gly Asn His Val Ser Pro Thr	1925	1930	1935
His Tyr Val Pro Glu Ser Asp Ala Ala Ala Arg Val Thr Ala Ile Leu	1940	1945	1950
Ser Ser Leu Thr Val Thr Gln Leu Leu Arg Arg Leu His Gln Trp Ile	1955	1960	1965
Ser Ser Glu Cys Thr Thr Pro Cys Ser Gly Ser Trp Leu Arg Asp Ile	1970	1975	1980
Trp Asp Trp Ile Cys Glu Val Leu Ser Asp Phe Lys Thr Trp Leu Lys	1985	1990	1995
Ala Lys Leu Met Pro Gln Leu Pro Gly Ile Pro Phe Val Ser Cys Gln	2005	2010	2015
Arg Gly Tyr Arg Gly Val Trp Arg Gly Asp Gly Ile Met His Thr Arg			

2020	2025	2030
Cys His Cys Gly Ala Glu Ile Thr Gly His Val Lys Asn Gly Thr Met 2035	2040	2045
Arg Ile Val Gly Pro Arg Thr Cys Arg Asn Met Trp Ser Gly Thr Phe 2050	2055	2060
Pro Ile Asn Ala Tyr Thr Thr Gly Pro Cys Thr Pro Leu Pro Ala Pro 2065	2070	2075 2080
Asn Tyr Lys Phe Ala Leu Trp Arg Val Ser Ala Glu Glu Tyr Val Glu 2085	2090	2095
Ile Arg Arg Val Gly Asp Phe His Tyr Val Ser Gly Met Thr Thr Asp 2100	2105	2110
Asn Leu Lys Cys Pro Cys Gln Ile Pro Ser Pro Glu Phe Phe Thr Glu 2115	2120	2125
Leu Asp Gly Val Arg Leu His Arg Phe Ala Pro Pro Cys Lys Pro Leu 2130	2135	2140
Leu Arg Glu Glu Val Ser Phe Arg Val Gly Leu His Glu Tyr Pro Val 2145	2150	2155 2160
Gly Ser Gln Leu Pro Cys Glu Pro Glu Pro Asp Val Ala Val Leu Thr 2165	2170	2175
Ser Met Leu Thr Asp Pro Ser His Ile Thr Ala Glu Ala Ala Gly Arg 2180	2185	2190
Arg Leu Ala Arg Gly Ser Pro Pro Ser Met Ala Ser Ser Ser Ala Ser 2195	2200	2205
Gln Leu Ser Ala Pro Ser Leu Lys Ala Thr Cys Thr Ala Asn His Asp 2210	2215	2220
Ser Pro Asp Ala Glu Leu Ile Glu Ala Asn Leu Leu Trp Arg Gln Glu 2225	2230	2235 2240
Met Gly Gly Asn Ile Thr Arg Val Glu Ser Glu Asn Lys Val Val Ile 2245	2250	2255
Leu Asp Ser Phe Asp Pro Leu Val Ala Glu Glu Asp Glu Arg Glu Val 2260	2265	2270
Ser Val Pro Ala Glu Ile Leu Arg Lys Ser Arg Arg Phe Ala Arg Ala		

2275	2280	2285
Leu Pro Val Trp Ala Arg Pro Asp Tyr Asn Pro Pro Leu Val Glu Thr		
2290	2295	2300
Trp Lys Lys Pro Asp Tyr Glu Pro Pro Val Val His Gly Cys Pro Leu		
2305	2310	2315 2320
Pro Pro Pro Arg Ser Pro Pro Val Pro Pro Pro Arg Lys Lys Arg Thr		
2325	2330	2335
Val Val Leu Thr Glu Ser Thr Leu Ser Thr Ala Leu Ala Glu Leu Ala		
2340	2345	2350
Thr Lys Ser Phe Gly Ser Ser Ser Thr Ser Gly Ile Thr Gly Asp Asn		
2355	2360	2365
Thr Thr Thr Ser Ser Glu Pro Ala Pro Ser Gly Cys Pro Pro Asp Ser		
2370	2375	2380
Asp Val Glu Ser Tyr Ser Ser Met Pro Pro Leu Glu Gly Glu Pro Gly		
2385	2390	2395 2400
Asp Pro Asp Leu Ser Asp Gly Ser Trp Ser Thr Val Ser Ser Gly Ala		
2405	2410	2415
Asp Thr Glu Asp Val Val Cys Cys Ser Met Ser Tyr Ser Trp Thr Gly		
2420	2425	2430
Ala Leu Val Thr Pro Cys Ala Ala Glu Glu Gln Lys Leu Pro Ile Asn		
2435	2440	2445
Ala Leu Ser Asn Ser Leu Leu Arg His His Asn Leu Val Tyr Ser Thr		
2450	2455	2460
Thr Ser Arg Ser Ala Cys Gln Arg Gln Lys Lys Val Thr Phe Asp Arg		
2465	2470	2475 2480
Leu Gln Val Leu Asp Ser His Tyr Gln Asp Val Leu Lys Glu Val Lys		
2485	2490	2495
Ala Ala Ala Ser Lys Val Lys Ala Asn Leu Leu Ser Val Glu Glu Ala		
2500	2505	2510
Cys Ser Leu Thr Pro Pro His Ser Ala Lys Ser Lys Phe Gly Tyr Gly		
2515	2520	2525
Ala Lys Asp Val Arg Cys His Ala Arg Lys Ala Val Ala His Ile Asn		

2530	2535	2540
Ser Val Trp Lys Asp Leu Leu Glu Asp Ser Val Thr Pro Ile Asp Thr		
2545	2550	2555 2560
Thr Ile Met Ala Lys Asn Glu Val Phe Cys Val Gln Pro Glu Lys Gly		
2565	2570	2575
Gly Arg Lys Pro Ala Arg Leu Ile Val Phe Pro Asp Leu Gly Val Arg		
2580	2585	2590
Val Cys Glu Lys Met Ala Leu Tyr Asp Val Val Ser Lys Leu Pro Leu		
2595	2600	2605
Ala Val Met Gly Ser Ser Tyr Gly Phe Gln Tyr Ser Pro Gly Gln Arg		
2610	2615	2620
Val Glu Phe Leu Val Gln Ala Trp Lys Ser Lys Lys Thr Pro Met Gly		
2625	2630	2635 2640
Phe Ser Tyr Asp Thr Arg Cys Phe Asp Ser Thr Val Thr Glu Ser Asp		
2645	2650	2655
Ile Arg Thr Glu Glu Ala Ile Tyr Gln Cys Cys Asp Leu Asp Pro Gln		
2660	2665	2670
Ala Arg Val Ala Ile Lys Ser Leu Thr Glu Arg Leu Tyr Val Gly Gly		
2675	2680	2685
Pro Leu Thr Asn Ser Arg Gly Glu Asn Cys Gly Tyr Arg Arg Cys Arg		
2690	2695	2700
Ala Ser Gly Val Leu Thr Thr Ser Cys Gly Asn Thr Leu Thr Cys Tyr		
2705	2710	2715 2720
Ile Lys Ala Arg Ala Ala Cys Arg Ala Ala Gly Leu Gln Asp Cys Thr		
2725	2730	2735
Met Leu Val Cys Gly Asp Asp Leu Val Val Ile Cys Glu Ser Ala Gly		
2740	2745	2750
Val Gln Glu Asp Ala Ala Ser Leu Arg Ala Phe Thr Glu Ala Met Thr		
2755	2760	2765
Arg Tyr Ser Ala Pro Pro Gly Asp Pro Pro Gln Pro Glu Tyr Asp Leu		
2770	2775	2780
Glu Leu Ile Thr Ser Cys Ser Ser Asn Val Ser Val Ala His Asp Gly		

2785	2790	2795	2800
Ala Gly Lys Arg Val Tyr Tyr Leu Thr Arg Asp Pro Thr Thr Pro Leu	2805	2810	2815
Ala Arg Ala Ala Trp Glu Thr Ala Arg His Thr Pro Val Asn Ser Trp	2820	2825	2830
Leu Gly Asn Ile Ile Met Phe Ala Pro Thr Leu Trp Ala Arg Met Ile	2835	2840	2845
Leu Met Thr His Phe Phe Ser Val Leu Ile Ala Arg Asp Gln Leu Glu	2850	2855	2860
Gln Ala Leu Asn Cys Glu Ile Tyr Gly Ala Cys Tyr Ser Ile Glu Pro	2865	2870	2875
Leu Asp Leu Pro Pro Ile Ile Gln Arg Leu His Gly Leu Ser Ala Phe	2885	2890	2895
Ser Leu His Ser Tyr Ser Pro Gly Glu Ile Asn Arg Val Ala Ala Cys	2900	2905	2910
Leu Arg Lys Leu Gly Val Pro Pro Leu Arg Ala Trp Arg His Arg Ala	2915	2920	2925
Arg Ser Val Arg Ala Arg Leu Leu Ser Arg Gly Gly Arg Ala Ala Ile	2930	2935	2940
Cys Gly Lys Tyr Leu Phe Asn Trp Ala Val Arg Thr Lys Leu Lys Leu	2945	2950	2955
Thr Pro Ile Ala Ala Ala Gly Arg Leu Asp Leu Ser Gly Trp Phe Thr	2965	2970	2975
Ala Gly Tyr Ser Gly Gly Asp Ile Tyr His Ser Val Ser His Ala Arg	2980	2985	2990
Pro Arg Trp Phe Trp Phe Cys Leu Leu Leu Leu Ala Ala Gly Val Gly	2995	3000	3005
Ile Tyr Leu Leu Pro Asn Arg	3010	3015	

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24

<210> 12

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ttgttcttgt cgggttaatg gcgc

24

<210> 13

<211> 24

<212> DNA

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<400> 13

gggtgtacta cacacatgag taag

24

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<212> DNA

<213> Hepatitis C virus

<400> 14

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22

<210> 15

<211> 40

<212> DNA

<213> Hepatitis C virus

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cgatcatgat acctcagcgg gcatatgcac tggacacgga

40

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<400> 16

gtccagtgcataatgcccgctgagg

24

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<211> 32

<212> DNA

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catgcaccagctgatatagcgtttgtaataatg

32

<210> 18

<211> 30

<212> DNA

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tccgtagaggaaagcttgacgcctgacgccc

30

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cagaggaggcagggtgctaataatgggcaatgtac

34

<210> 20

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43

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aatgc 65

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28

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41

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38

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27

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